

Pravastatin Preserves Vasomotor Response in Atherosclerotic Arteries after Balloon Angioplasty

Ma Hongbao*, **, Yang Yan **, Cherng Shen ***

* Bioengineering Department, Zhengzhou University, Zhengzhou, Henan 450001, China,
mahongbao@zzu.edu.cn; hongbao@gmail.com; 01186-137-8342-5354

** Brookdale University Hospital and Medical Center, Brooklyn, New York 11212, USA,
youngjenny2008@yahoo.com

*** Department of Electrical Engineering, Chengshiu University, Niaosong, Taiwan 833, China,
cherngs@csu.edu.tw; 011886-7731-0606 ext 3423

Abstract: Vasodilation response to pharmacological challenge is inhibited following balloon angioplasty. Pravastatin, a 3-hydroxy-3-methylglutaryl coenzyme A reductase inhibitor, has been demonstrated to enhance endothelial cell production of nitric oxide and reduce low-density lipoprotein cholesterol. This study was conducted to evaluate the effect of pravastatin on vasodilation following balloon angioplasty in normal and atherosclerotic arteries. Three normal and 3 atherosclerotic New Zealand White rabbits were used. Atherosclerosis was induced by feeding a high cholesterol diet. Rabbits were sacrificed and carotid arteries were isolated and placed in a dual perfusion chamber. Both arteries from each rabbit were perfused with oxygenated physiologic buffered solution at 37°C and 60 mmHg. One artery was exposed to pravastatin (100 µM) and the other served as control. Balloon angioplasty (BA) was performed in both arteries using a 2.5×15 mm balloon catheter inflated to 10 atm at 3 different sites for one minute each. Pharmacological challenge was given using acetylcholine (2×10^{-5} M) and sodium nitroprusside (2×10^{-5} M) in norepinephrine (2×10^{-6} M) preconstricted arteries. Vessel diameter was measured by a computer planimetry system. After BA in normal rabbit arteries, acetylcholine did not demonstrate significant difference in percent lumen dilation between control and pravastatin (25.5 ± 10.4 vs 16.6 ± 7.5 , p=ns) while atherosclerotic arteries had significantly preserved vasomotor response with pravastatin (16.9 ± 7.2 vs 33.6 ± 18.2 , p<0.005). Similar results were noted with nitroprusside in normal arteries (29.0 ± 14.5 vs 18.0 ± 10.5 , p=ns) and atherosclerotic arteries (18.6 ± 7.4 vs 38.4 ± 19.8 , p<0.003). Pravastatin preserved vasomotor response in atherosclerotic arteries following BA when compared to normal arteries. This effect may be due to an enhanced production of nitric oxide in atherosclerotic arteries. However, pravastatin also appears to influence vasomotor response by either non-endothelial dependent or a combination of endothelial and non-endothelial dependent mechanism. [Journal of American Science 2009;5(4):101-106]. (ISSN: 1545-1003).

Keywords: artery; atherosclerosis; pravastatin; rabbit

Abbreviation: Ach: Acetylcholine; EDTA: Ethylenediamine-tetraacetic acid; G-6-Pase: Glucose 6-phosphatase; HDL: High-density lipoprotein; HMG-CoA: Inhibiting 3-hydroxy-3-methylglutaryl-coenzyme A; LDL: Low-density lipoproteins; NE: Norepinephrine; PBS: Physiologic buffered saline solution; SN: Sodium nitroprusside

1. Introduction

Atherosclerosis is a major reason resulted in cardiovascular disease and stroke (Zhu and Zhang, 2008). Hypercholesterolemia is a recognized independent risk factor for coronary heart disease, and the lack of physical activity in the general population is a public health problem and is recognized as an independent risk factor for the development of coronary disease (Boraita Perez, 2008). Epidemiological studies have shown that hypertriglyceridemia and low HDL-cholesterol were both associated with an increased risk of coronary heart disease (Fruchart and Duriez, 2006). Drug therapy is recommended for patients whose low-

density lipoprotein (LDL) cholesterol concentrations are not adequately lowered by dietary modifications. Statins (atorvastatin, cerivastatin, fluvastatin, lovastatin, pravastatin and simvastatin) are the most effective agents currently available for lowering plasma levels of LDL cholesterol by inhibiting 3-hydroxy-3-methylglutaryl-coenzyme A (HMG-CoA) reductase, a key enzyme in the synthesis of cholesterol (Blumenthal, 2000; Gbelcova et al., 2008; Neuvonen et al., 2006). HMG-CoA reductase is a rate-controlling microsomal enzyme that converts HMG-CoA to mevalonic acid, a precursor of cholesterol (Shitara and Sugiyama, 2006). HMG-CoA reductase inhibitors are the primary

hypolipidemic drug treatment in most countries and statins are the mainstay of therapy for hyperlipidemia (Illingworth and Tobert, 1994). Pharmacological lowering of LDL cholesterol concentration has been shown in several primary and secondary intervention trials to decrease the occurrence of coronary heart disease and to prevent or delay coronary heart disease progression and the regression of atherosclerotic lesions has been demonstrated in some patients (Levy, 1984). Pravastatin is a new HMG-CoA reductase inhibitor for the treatment of hypercholesterolemia, which reduces LDL cholesterol and increases high-density lipoprotein (HDL) cholesterol (Jungnickel et al., 1992).

Pravastatin is formed by microbial transformation by the *microorganism Nocardia autotrophica*. The structural formula of pravastatin is shown in Figure 1 along with those of lovastatin and simvastatin. The dihydroxyheptanoic acid moiety is the substrate analogue that interacts with the active site of HMG-CoA reductase. The decalin ring interacts with the binding site. Pravastatin sodium is chemically designed as [1S-[1 α (β S*, δ S*), 2 α , 6 α , 8 β (R*)], 8 α]-1,2,6,7,8,8a-hexahydro- β , δ ,6-trihydroxy-20methyl-8-(2-methyl-1-oxobutoxy)-1-naphthaleneheptanoic acid, monosodium salt. Its empirical formula is C₂₃H₃₅NaO₇ and its molecular weight is 446.52 (Watanabe et al., 1988). Pravastatin is a hygroscopic, crystalline powder that is freely soluble in water (>300 mg/ml) and methanol, slightly soluble in isopropanol, and practically insoluble in acetone, acetonitrile, chloroform and ether (Watanabe et al., 1988). Atherosclerotic plaque disruptions with subsequent arterial thrombosis are critical causes for acute coronary ischemic syndromes (Bentzon et al., 2007; Libby, 2006). Pharmacological protections of artery are under searching. We suppose that pravastatin will preserve the vasoactivity of the atherosclerotic artery by its affection of lowering LDL cholesterol levels.

The effects of statin drug therapy on the cardiovascular system extend beyond their anti-hyperlipidemic properties. Many studies showed that statins have a pronounced antioxidant effect as well as well documented endothelial protective effect (Namazi, 2004). Statins are 3-hydroxy-3-methylglutaryl-coenzyme A (HMG-CoA) reductase inhibitors, a key enzyme in the synthesis of cholesterol. They have been shown to decrease significantly risk of cardiac events in the setting of primary prevention, secondary prevention or during acute coronary syndrome (MIRACL trial). Since the effect of statin therapy is so diversified we selected to study effect of pravastatin on vasoreactivity of the endothelium following acute physical injury

mimicking the current intravascular interventions.

3-Hydroxy-3-methylglutaryl-coenzyme A reductase (HMGR) A reductase (HMGR) catalyzes the formation of mevalonate - converts HMG-CoA to mevalonic acid (Zhang, Wan et al. 2007). In many classes of organisms, this is the committed step leading to the synthesis of essential compounds, such as cholesterol. However, a high level of cholesterol is an important risk factor for coronary heart disease, for which an effective clinical treatment is to block HMGR using inhibitors like statins. Recently the structures of catalytic portion of human HMGR complexed with six different statins have been determined by a delicate crystallography study (Ma et al, 2008).

2. Materials and Methods

Six, male, New Zealand White rabbits (Harlan-Sprague Dawley, Inc., Indianapolis, IN, USA) weighing between 2.8 and 3.2 kg were used in this study. The control group consisted of 3 normal rabbits that were fed a regular diet (Harlan-Sprague Dawley, Inc., Indianapolis, IN, USA) for six months. To induce atherosclerosis, another 3 rabbits underwent balloon-induced arterial injury, then were maintained on a 1% cholesterol diet (Harlan-Sprague Dawley, Inc., Indianapolis, IN, USA) for one month followed by another month of alternated regular diet and the two months diet cycle was repeated for three times to keep the rabbits for total of six months. Balloon-induced arterial wall injury of the aorta was performed with a 4F Fogarty Arterial Embolectomy catheter (0.9 × 40 cm, Baxter Healthcare Corporation, Irvine, CA, USA) introduced through the right femoral artery cutdown. The catheter was advanced in a retrograde fashion to the aortic valve and then withdrawn 3 cm. The balloon was inflated with 1.5 cm³ of air, and the catheter was retracted down to the iliofemoral artery. This was repeated three times in each rabbit. Rabbits were anesthetized with ketamine (50 mg/kg, IM, Fort Dodge Animal Health, Fort Dodge, Iowa, USA) and xylazine (20 mg/kg, IM, The Butler Company, Columbus, Ohio, USA) in this surgery process.

After intravenous administration of heparin sulfate (1000 IU/rabbit) (Elkins-Sinn, Inc, Cherry Hill, NJ, USA) to prevent postmortem clotting, rabbits were anesthetized by injecting nembutal sodium solution (pentobarbital 50 mg/ml, 1 ml/kg rabbit) (Abbot Laboratories, North Chicago, IL, USA) through a marginal ear vein. Both carotid arteries from each rabbit were removed immediately after rabbits were sacrificed and were immersed in oxygenated physiologic buffered saline solution (NaCl 119 mM, KCl 4.7 mM, CaCl₂ 2 mM, NaH₂PO₄ 1.2 mM, MgSO₄ 1.2 mM, NaHCO₃ 22.6 mM,

glucose 5.5 mM, Na₂EDTA 0.03 mM) (PBS). Arteries were perfused in a dual organ chamber under 60 mmHg flow pressure and 2.5 ml/min flow rate at 37°C and artery diameter vasodilation was measured. Balloon angioplasty was performed in both arteries using a 2.5×15 mm balloon catheter inflated to 10 atm at 3 different sites for 1 minute. One artery was served as non-pravastatin control and the other one was exposed to pravastatin (100 µM). After norepinephrine (NE, 2×10⁻⁶ M, Sigma Chemical Co, St Louis, MO, USA) preconstriction, pharmacological challenge was done with acetylcholine (Ach, 1×10⁻⁵ M, Sigma, St. Louis, MO, USA) and sodium nitroprusside (SN, 1×10⁻⁵ M,

Sigma, St. Louis, MO, USA) (Table 1).

Data were calculated according to the formulas: (PBS-NE) (%)=(PBS-NE)/NE×100, (Ach-NE) (%)=(Ach-NE)/NE×100 and SN-NE (%)=(SN-NE)/NE×100 separately, where Ach, NE, PBS and SN represented average diameter (mm) of arteries that were perfused by PBS containing a corresponding chemical. Balloon angioplasty was performed in both arteries using a 2.5×15 mm balloon catheter inflated to 10 atm at 3 different sites for 1 minute. The Vessel diameter was measured by a computer planimetry system (Figure 1). Procedures were performed according to NIH Animal Care.

Table 1. Artery Perfusion Step for Diameter Measurement

Treatments	Perfusion Steps	Perfusion Steps - Abbreviation in Figures	Perfusion Time (min)
Cycle 1: Before balloon injury Without pravastatin	Buffer 1	B1	10
	NE 1	N1	10
	Ach 1	A1	10
	SN 1	S1	10
	Buffer 2	B2	10
Cycle 2: Before balloon injury Half of arteries with pravastatin (100 µM)	Buffer 3	B3	10
	NE 2	N2	10
	Ach 2	A2	10
	SN 2	S2	10
	Buffer 4	B4	10
Cycle 3: After balloon injury Half* of arteries with pravastatin (100 µM)	Buffer 5	B5	10
	NE 3	N3	10
	Ach 3	A3	10
	SN 3	S3	10
	Buffer 6	B6	10

* Same arteries as in cycle 2

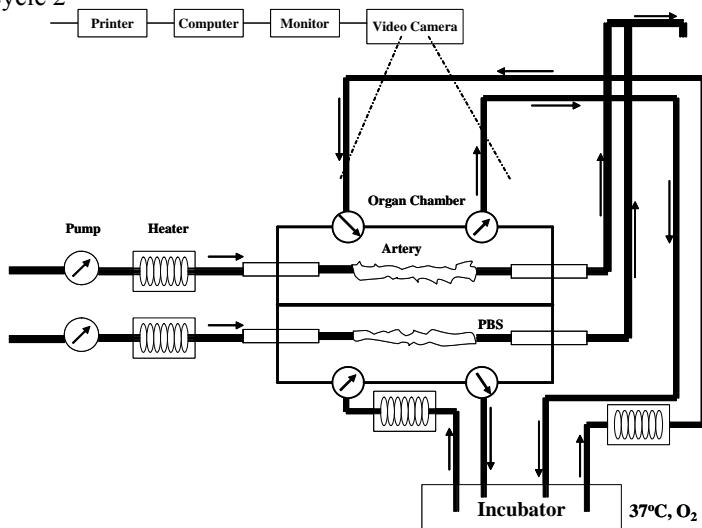


Figure 1. Dual organ chamber with separate perfusion using oxygenated physiological buffered solution at 37°C. Vessel diameter was measured by a computer planimetry system.

Statistical analysis:

With Jandel Scientific program, SigmaStat (Sigma Chemical Co., St. Louis, MO, USA) was used for data statistical analysis. $P<0.05$ was considered statistically significant difference. Measured data were reported as mean \pm SD. The student t-test was used for different studies.

3. Results

From the observation of aorta arteries, all the rabbits were atherosclerosis with the balloon-induced injury and maintained on a 1% cholesterol diet for one month alternatively up to 6 month feeding.

Myocardial infarction in human cases a triggering activity such as physical exertion precipitates the acute onset of the disorder (Mittleman et al., 2005; Muller et al., 1989; Tofler et al., 1990), but it is difficult to be studied in human. Therefore, a suitable animal model is important for the research in this field. This study demonstrated that atherosclerotic rabbit can be induced with balloon induced arterial injury surgery combined with 6 months of alternative 1% cholesterol diet. The rabbits which were balloon induced arterial injury

and then were maintained in an alternative 1% cholesterol diet for a total of 6 months clearly caught atherosclerosis. This model is a useful method to get atherosclerotic animal for the related scientific research purpose.

In this experiment, the vasoactivity of both normal and atherosclerotic rabbit carotid arteries was measured using NE preconstriction and pharmacological challenge with Ach and SN (Figure 2, 3, 4). The measurements were performed with the steps of a perfusion cycle as showed in Table 1.

For normal rabbit arteries under PBS perfusion and pharmacological challenged by Ach and SN to NE, there was no significant difference in percent vasodilation between control and pravastatin in both before balloon injury and after balloon injury cycle (25.5 ± 10.4 vs 16.6 ± 7.5 for Ach, $p=ns$; 4.5 vs 18.0 ± 10.5 for SN, $p = ns$). However, compared to control, pravastatin demonstrated a significantly greater percent vasodilation on atherosclerotic arteries after balloon angioplasty (16.9 ± 7.2 vs 33.6 ± 18.2 for Ach, $p<0.005$; 18.6 ± 7.4 vs 38.4 ± 19.8 for SN, $p<0.003$) (Figure 2, 3, 4).

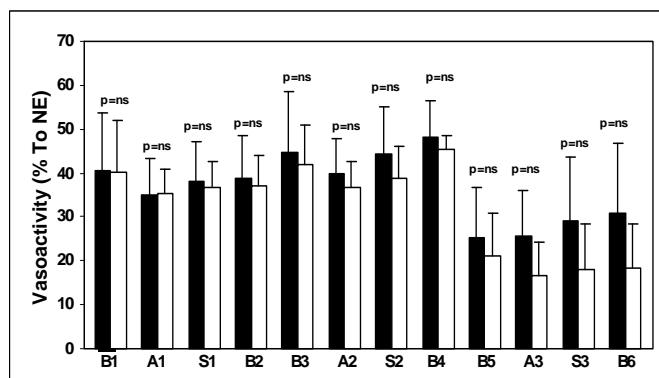


Figure 2. Vasoactivity (% to norepinephrine) of control rabbit.

■ : Control; □ : Pravastatin. Letter meanings are shown in Table 1.

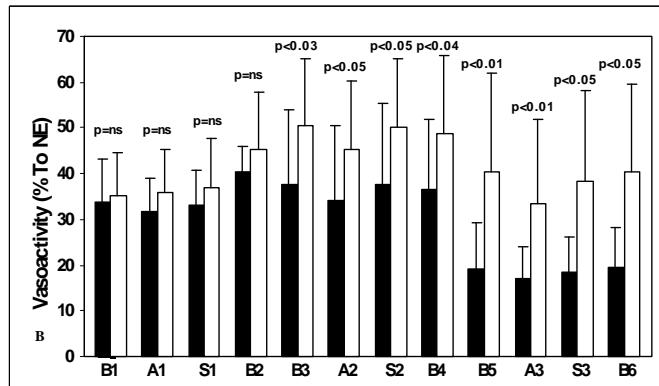


Figure 3. Vasoactivity (% to norepinephrine) of atherosclerotic rabbit.

■ : Control; □ : Pravastatin. Letter meanings are shown in Table 1.

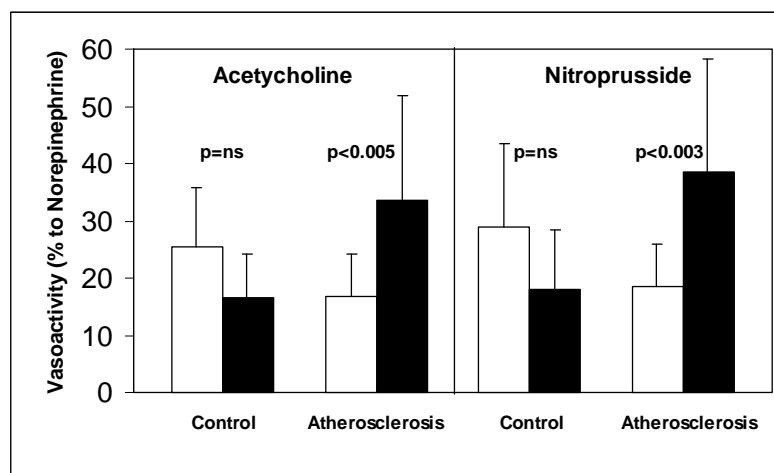


Figure 4. Vasoactivity (% to Norepinephrine) of rabbit carotid balloon injured. □ Perfusion by buffer; ■ Perfusion by buffer plus pravastatin.

Aasodilatation range of ratio to NE of PBS and pharmacological challenge with Ach and SN was 30-50% for before balloon injury and 20-40% for after balloon injury. Pravastatin enhances the vasodilation response in atherosclerotic arteries following balloon angioplasty. Pravastatin may influence vasodilation by a combination of endothelial and non-endothelial dependent mechanism.

Pravastatin enhances both endothelium dependant and independent vasoreactivity of carotid arteries in setting of acute balloon injury. This was true in presence of atherosclerosis. Mechanism of this effect is unlikely to be due to the lipid lowering property of pravachol. This might be due to its antioxidant effect or to some other unidentified process through a direct endothelial process, or the activation of some receptors or the induction of some signal that ultimately relates to the genomic makeup of endothelium. Its effect on the NO system is being established and might be part of the puzzle.

4. Discussions

Atherosclerosis, or "hardening of the arteries", is the process that causes heart attacks and most strokes (Rahmani et al., 2006). It is currently believed that cholesterol, especially the LDL, must be modified or oxidized before they can be taken up to cause foam cells. When cells use oxygen for energy, they produce by-products called free radicals. Free radicals damage cells and tissues during a process called oxidation - a factor in many chronic illnesses, including some forms of cancer, cataracts, arthritis and cardiovascular disease. LDL, known as the "bad cholesterol", is actually a protein that carries cholesterol throughout the body. The cholesterol

carried by LDL deserves its bad reputation, however. It often ends up in our arteries, causing clots that can lead to heart attacks. Oxidation of LDL-cholesterol contributes to the plaque build-up in arteries, a process called atherosclerosis that can cause blockages and reduced blood flow. The process also plays a role in the loss of elasticity in arteries.

Antioxidants help neutralize free radicals and prevent them from causing cellular damage (McDaniel et al., 2003). Once oxidized, cholesterol is less apt to be expelled by body's cleaning mechanisms and more likely to be stored in arteries.

One problem for atherosclerotic rabbits is that their free radical and oxidation condition are changed under disease. Free radical modification of serum that is not the solely increased level of lipoprotein oxidation products in blood lipoproteins is an important cause for cholesterol accumulation in cells, and apparently for their transformation into foam cells during atherosclerosis (Panasenko et al., 1991).

Once altered by free radical oxidation, plasma lipoproteins undergo dramatic change, both in the manner in which they can interact with cells and in the ways in which they influence cell functions (Chisolm, 1991). Pravastatin preserved vasomotor response in atherosclerotic arteries following BA when compared to normal arteries. This effect may be due to an enhanced production of nitric oxide in atherosclerotic arteries. However, pravastatin also appears to influence vasomotor response by either non-endothelial dependent or a combination of endothelial and non-endothelial dependent mechanism. Pravastatin play protection function on vascular activity may through anti-oxidation.

Correspondence to:

Ma Hongbao, PhD, Professor
 Bioengineering Department, Zhengzhou University
 Zhengzhou, Henan 450001, China
hongbao@gmail.com; 01186-137-331-67674

References

1. Bentzon JF, Sondergaard CS, Kassem M, Falk E. Smooth muscle cells healing atherosclerotic plaque disruptions are of local, not blood, origin in apolipoprotein E knockout mice. *Circulation.* 2007;116(18):2053-61.
2. Blumenthal RS. Statins: effective antiatherosclerotic therapy. *Am Heart J.* 2000;139(4):577-83.
3. Boraita Perez A. Exercise as the cornerstone of cardiovascular prevention. *Rev Esp Cardiol.* 2008;61(5):514-28.
4. Chisolm GM, 3rd. Antioxidants and atherosclerosis: a current assessment. *Clin Cardiol.* 1991;14(2 Suppl 1):I25-30.
5. Fruchart JC, Duriez P. Mode of action of fibrates in the regulation of triglyceride and HDL-cholesterol metabolism. *Drugs Today (Barc).* 2006;42(1):39-64.
6. Gbelcova H, Lenicek M, Zelenka J, Knejzlik Z, Dvorakova G, Zadinova M, Pouckova P, Kudla M, Balaz P, Rum T, Vitek L. Differences in antitumor effects of various statins on human pancreatic cancer. *Int J Cancer.* 2008;122(6):1214-21.
7. Illingworth DR, Tobert JA. A review of clinical trials comparing HMG-CoA reductase inhibitors. *Clin Ther.* 1994;16(3):366-85; discussion 365.
8. Jungnickel PW, Cantral KA, Maloley PA. Pravastatin: a new drug for the treatment of hypercholesterolemia. *Clin Pharm.* 1992;11(8):677-89.
9. Levy RI. Causes of the decrease in cardiovascular mortality. *Am J Cardiol.* 1984;54(5):7C-13C.
10. Libby P. Atherosclerosis: disease biology affecting the coronary vasculature. *Am J Cardiol.* 2006;98(12A):3Q-9Q.
11. Ma H, Jenny Y, Cherng S. HMG-CoA reductase (3-hydroxy-3-methyl-glutaryl-CoA reductase) (HMGR). *Journal of American Science.* 2008;4(3):62-4.
12. McDaniel MA, Maier SF, Einstein GO. "Brain-specific" nutrients: a memory cure? *Nutrition.* 2003;19(11-12):957-75.
13. Mittleman MA, Maclure M, Glasser DB. Evaluation of acute risk for myocardial infarction in men treated with sildenafil citrate. *Am J Cardiol.* 2005;96(3):443-6.
14. Muller JE, Tofler GH, Stone PH. Circadian variation and triggers of onset of acute cardiovascular disease. *Circulation.* 1989;79(4):733-43.
15. Namazi MR. Statins: novel additions to the dermatologic arsenal? *Exp Dermatol.* 2004;13(6):337-9.
16. Neuvonen PJ, Niemi M, Backman JT. Drug interactions with lipid-lowering drugs: mechanisms and clinical relevance. *Clin Pharmacol Ther.* 2006;80(6):565-81.
17. Panasenko OM, Vol'nova TV, Azizova OA, Vladimirov YA. Free radical modification of lipoproteins and cholesterol accumulation in cells upon atherosclerosis. *Free Radic Biol Med.* 1991;10(2):137-48.
18. Rahmani M, Cruz RP, Granville DJ, McManus BM. Allograft vasculopathy versus atherosclerosis. *Circ Res.* 2006;99(8):801-15.
19. Shitara Y, Sugiyama Y. Pharmacokinetic and pharmacodynamic alterations of 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase inhibitors: drug-drug interactions and interindividual differences in transporter and metabolic enzyme functions. *Pharmacol Ther.* 2006;112(1):71-105.
20. Tofler GH, Stone PH, Maclure M, Edelman E, Davis VG, Robertson T, Antman EM, Muller JE. Analysis of possible triggers of acute myocardial infarction (the MILIS study). *Am J Cardiol.* 1990;66(1):22-7.
21. Watanabe Y, Ito T, Shiomi M, Tsujita Y, Kuroda M, Arai M, Fukami M, Tamura A. Preventive effect of pravastatin sodium, a potent inhibitor of 3-hydroxy-3-methylglutaryl coenzyme A reductase, on coronary atherosclerosis and xanthoma in WHHL rabbits. *Biochim Biophys Acta.* 1988;960(3):294-302.
22. Zhang QY, J Wan, et al. Structure-based rational quest for potential novel inhibitors of human HMG-CoA reductase by combining CoMFA 3D QSAR modeling and virtual screening. *J Comb Chem.* 2007;9(1):131-8.
23. Zhu Y, Zhang W. Cloning and analyzing of the cDNA sequence of N-terminal region and C-terminal region of zinc finger protein (ZFP580) gene. *Life Science Journal.* 2008; 5(1): 11 – 6.