

## Association of serum Leptin and Adiponectin with Atherosclerosis in obese and non-obese Type 2 Diabetes Mellitus patients

Mohga S Abdalla<sup>1</sup>, Hayat M Sharada<sup>1</sup>, Ashraf I Amin<sup>2</sup>, Nervana Samy<sup>3</sup>, Magda Sayed<sup>3</sup>, Esmat Ashour<sup>3</sup> and Elham M Youssef-Elabd<sup>3</sup>

<sup>1</sup>Biochemistry Dept, Faculty of Science, Helwan University, Egypt;

<sup>2</sup>Clinical Pathology Dept, National Institute of Diabetes & Endocrinology, Cairo, Egypt.

<sup>3</sup>Biochemistry Dept, National Research Centre, Dokki, Giza, Egypt

[Nervana91@hotmail.com](mailto:Nervana91@hotmail.com)

**Abstract:** Obesity is a major risk factor for insulin resistance, type 2 diabetes, heart disease, and many other chronic diseases. The current study was designed to investigate the endogenous mechanism by which obesity may increase the risk of CVD by examining whether serum adiponectin, Leptin or insulin mediate the association of obesity and type 2 diabetes and cardiovascular risk factors in Egyptian adult patients. **Patients and Methods:** This study included 82 subjects, 30 patients suffering from type 2 diabetes and 52 patients suffering from type 2 diabetes together with coronary artery disease (CAD) together with another group having CAD without diabetes. They were classified according to their body mass index (BMI) into obese and non-obese groups, also 25 healthy volunteers were considered as controls. All patients were subjected to anthropometric assessment and laboratory determination of serum Adiponectin, Leptin, insulin and glucose. Insulin resistance was established by homeostasis model assessment (HOMA-IR). Differences in clinical or laboratory parameters among groups were compared by using one-way ANOVA test. **Results** revealed highly significant decrease in Adiponectin levels and highly significant increase in serum Leptin in non obese groups (G1 (T2D), G2 (CAD) and G3 (T2D+ CAD) as compared to controls. However, there were no statistical variations between non obese groups when compared to each others. HOMA-IR showed highly significant increase in non obese groups as compared to both controls and each other. Also, the results showed high significant decrease in Adiponectin and highly significant increase in Leptin in obese groups (G4 (T2D), G5 (CAD) and G6 (T2D+CAD) when compared to controls. However, there were no statistical variations between obese groups when compared to each others as regard Adiponectin, while Leptin showed statistical increase between (G4) and (G5) groups when compared to each others, HOMA-IR showed highly significant increase in the two obese groups only (G4 and G6) as compared to controls, while there was no significant variation in (G5) when compared to controls. Moreover, there was a significant increase in all obese groups when compared to each other. Also, there was significant correlation between serum Adiponectin and Leptin in obese DM patients. **Conclusion:** The coexistence of correlation between serum leptin and Adiponectin levels in addition to increase of serum leptin and decrease serum Adiponectin levels in obese DM patients in the current study; support the hypothesis of their susceptibility to atherosclerosis. [Journal of American Science 2010;6(5):153-164]. (ISSN: 1545-1003).

**Keywords:** Type2 Diabetes, Cardiovascular disease, Adiponectin, Leptin, HOMA-IR

### 1. Introduction

Obesity is a major risk factor for insulin resistance (IR), type 2 diabetes, heart disease and many other chronic diseases. These associations are influenced by adipose tissue (AT) distribution (Marcus et al., 1999). IR is characteristically more severe in T2DM than in similarly obese non-diabetics, but whether this difference is related to differences in body composition is unclear (Azuma et al., 2007; Marinou et al., 2009).

Type 2 diabetes mellitus (T2D) is considered one of the major metabolic diseases of 21st century (Kowalska, 2007, Thévenod 2008). The excessive intake of food, sedentary life style and lack of physical activity are responsible for the growing epidemic of obesity, together with the increasing rate of T2D in many parts of the world (Zimmet, et al., 2001). The

main burden of T2D is connected with development of vascular complications, which are a consequence of accelerated atherogenesis (Cooper et al., 2001; Salas-Salvadó et al., 2007).

Atherosclerotic cardiovascular complications are the major causes of morbidity and mortality in type 2 diabetic patients. Cardiovascular disease is the main cause of premature mortality and two to six times greater morbidity of T2D patients than of non-diabetic people. There are several mechanisms which could play a role in the pathogenesis of vascular complications and most of them are triggered by hyperglycemia and hyperglycemia-induced oxidative stress (Qasim et al., 2008).

However, during the past few years a lot of attention has been paid to the potential role of adipose

tissue in the development of vascular complications of diabetes. As obesity is considered to be a major risk factor for atherosclerosis, understanding of the underlying mechanisms leading to obesity and linking obesity with atherogenesis is necessary, for the development of therapeutic strategies against atherosclerosis. The pathophysiology of CVD linked to obesity is an area of intensive research (Marinou et al., 2009). The relation between obesity and CVD is indeed complicated. Some investigators suggest that the connection is indirect and dependent on the increased prevalence of diabetes, hypertension and dyslipidemia, whereas others have demonstrated an independent association between obesity (and especially abdominal obesity) and CVD risk (Frayn 2005). The relationship between obesity and CVD appears to develop at a relatively young age. Obesity in young men, aged 15 to 34 years, is associated with accelerated coronary atherosclerosis (McGill et al. 2002).

Excessive adiposity is the most important risk factor in the development of insulin resistance and type 2 diabetes mellitus (Bloomgarden.,2002). The pathophysiology linking obesity to type 2 diabetes is not completely understood, but adipokines are thought to be involved (Jazet et al., 2003). Adipose tissue produces several proteins (adipocytokines) such as leptin, adiponectin, resistin, tumor-necrosis factor- $\alpha$ , and interleukin (IL)-6, that modulate insulin sensitivity and appear to play an important role in the pathogenesis of insulin resistance, diabetes, dyslipidemia, inflammation, and atherosclerosis (Trujillo & Scherer , 2005, Arita et al., 1999 Kershaw and Flier 2004; Arner ,2005; Fantuzzi .,2005; Hotamisligil and Spiegelman 1994; Hotamisligil et al., 1993, Hotamisligil et al.,1994). However, the mechanisms by which fat tissue induces insulin resistance and the role of adipocytokines in the pathogenesis of type 2 diabetes mellitus have not been well established.

Adipokines are fat-secreted biomolecules with diverse signaling effects that modulate insulin resistance, hepatic lipoprotein production, and vascular inflammation . Two in particular, Adiponectin and leptin, are almost exclusively fat derived and have antithetic actions in insulin resistance and in vascular signaling. Because of these properties, Adiponectin and leptin have been proposed as biomarkers of adipose function that may add value in predicting cardiovascular disease (CVD) risk and provide targets for therapeutic interventions (Qasim et al., 2008).

Levels of Adiponectin, an insulin-sensitizing hormone with anti-inflammatory properties, are reduced in obesity, type 2 diabetes, and coronary artery disease (CAD) compared with controls (Behre., 2007) Indeed, several (Schulze et al., 2005, Pischon et al., 2004]) but not all (Lawlor et al., 2005, Lindsay et al.,

2005) epidemiologic studies suggest that reduced plasma Adiponectin levels are independent predictors of

Leptin, on the other hand, is a pleiotropic adipokine that modulates innate immune functions and vascular signaling in addition to its central role in regulation of appetite and energy expenditure (Hamann & Matthae, 1996) In contrast to adiponectin, leptin levels directly correlate with insulin resistance, obesity (Chu et al.,2000) and several CVD risk factors (Haynes., 2005) Leptin levels have been associated with CVD beyond body mass index (BMI) in some studies (Reilly et al.,2004) Several large population based studies have suggested strong positive associations between leptin and insulin resistance and its components including hyperlipidemia and hypertension, independent of body mass index (Koerner et al.,2005).

The current study was designed to investigate the endogenous mechanism by which obesity may increase the risk of CVD by examining whether fasting Adiponectin ,leptin or insulin mediate the association of obesity and type2 diabetes, and thrombogenic cardiovascular risk factors in a national population-based cohort of Egyptian adults. A second purpose was to determine whether Adiponectin, leptin and insulin are associated with these CVD risk factors, independent of obesity and type2 diabetes.

## Patients and Methods

### Patients

This study included 81 subjects, 29 patients suffering from type 2 diabetes selected from National Institute of Diabetes and Endocrinology, 52 patients suffering from type 2 diabetes together with coronary artery disease (CAD) and another group with CAD without diabetes were selected from the National Institute of Heart Disease, they were classified according to their body mass index (BMI) into obese and non-obese groups, obesity was defined according to the WHO criterion (BMI>30kg/m<sup>2</sup>), also 25 healthy volunteers were considered as controls.

Type 2 diabetes mellitus was diagnosed based on the criteria of World Health Organization criteria (Alberti and Zimmet, 1998). The duration of diabetes was 9.1+ 0.7 years (mean+ SD). No subject had clinical or laboratory signs of acute infection and none had a history or presence of clinically evident cardiovascular disease. They do not receive insulin therapy.

The CAD inclusion criteria were: angiographic evidence with >50% occlusion of 1 major coronary arteries, old myocardial infarction, or angina pectoris, but any possible non-Atherogenesis occlusions such as osteal stenosis and spasm were excluded. Electrocardiogram (ECG)) abnormality was

defined when the interpretation was of a trial fibrillation, a major ST-T segment change, left ventricular hypertrophy or a ventricular conduction defect.

All subjects provided informed consent and the study protocol was approved by the Ethics Committee of the Institutional Review Board of National Institute of Diabetes & Endocrinology and National Heart Institute.

All patients were classified according to their body mass index (BMI) into obese and non-obese groups, obesity was defined according to the WHO criterion (BMI>30kg/m<sup>2</sup>).

- G1: included 10 non-obese suffering from type II diabetes mellitus patients without history of cardiovascular complications (T2DM)
- G2: included 19 non-obese patients suffer from Coronary Artery Disease without suffering from type II diabetes mellitus (CAD)
- G3: included 10 non-obese patients suffer from T2DM+CAD
- G4 included 19 obese suffering from type II diabetes mellitus patients without history of cardiovascular complications
- G5: included 10 non-obese patients suffer from CAD without suffering from type II diabetes mellitus
- G6: included 13 obese patients suffer from T2DM+CAD

Beside reference group included (25) healthy as controls. Criteria for normal controls included (1) absence of history of CHD. (2) Absence of hypertension, diabetes mellitus, or impaired renal function, and (3) Normal ECG and chest x-ray.

### **Anthropometric assessments**

#### **Body mass index:**

Body mass index calculated as the body weight divided by the square of the height (kg/m<sup>2</sup>) was used as a marker of obesity. Weight and height were measured on the third or fourth day after admission while the subjects were fasting and wearing only their undergarments. Patients were designated as obese where BMI exceeded 30 kg/m<sup>2</sup> and were considered non-obese where BMI was below 30 kg/m<sup>2</sup>.

#### **Blood samples and Laboratory assessments:**

Venous blood samples were obtained from all the patients on admission to the hospital. Venous blood specimens were collected without EDTA-treated and plain tubes after a 12-h fast. The tubes were immediately placed on ice until they arrived at the laboratory room (within 1–3h) and were stored at –70

°C until analysis. The CVs of the biomarker measurements were all <6%.

For glucose tests, blood was collected in fluorinated tubes and plasma was immediately separated and kept refrigerated at 4° C for up to 48 hours. Plasma glucose was determined by the glucose-oxidase method.

#### **Assessment of insulin resistance**

Fasting plasma glucose (FPG) was measured using an enzymatic colorimetric method with glucose oxidase. Insulin was measured by micro particle enzyme assay (Abbott, Chicago, IL). Homeostasis model assessment (HOMA) or (HOMAIR) was used to detect the degree of insulin resistance (Matthews et al., 1985). In each subject, the degree of insulin resistance can be assessed from the fasting glucose and insulin concentrations by the formula: resistance (HOMA-IR.)

HOMA IR (%) = fasting blood glucose (mg/dL)/18 × fasting insulin (μU/mL)/22.5.

#### **Measurement of serum Adiponectin and Leptin**

Serum samples for measurement of Adiponectin and leptin were stored at - 80°C until subsequent analysis. The quantitative determination of human Leptin was conducted in serum by solid phase ELISA techniques, using commercially available kit, purchased from R&D Systems Inc., (Minneapolis, USA). Adiponectin was determined by ELISA (Human Adiponectin ELISA Kit, Linco Research, Inc., St. Charles, MO; intra-assay coefficient of variation, 1.0–7.4%; intra-assay coefficient of variation, 2.4–8.4%; sensitivity, 0.5 ng/ml).

#### **Statistical analyses**

Statistical analyses were conducted using SPSS for Windows, Version 11.0 (SPSS, Chicago, IL). Data are expressed as means ± standard deviations

Significant differences between groups were evaluated using ANOVA with post hoc testing. Possible associations between variables were tested using Pearson's correlation. Differences were considered statistically significant at  $p < 0.05$ .

#### **Results**

Table (1a &b) and figure (1&2) summarizes data and Multiple Comparisons (Post Hoc Tests) among the different studied non-obese groups.

#### **-Serum Adiponectin:**

##### **In non obese groups**

The results revealed that the mean values ± S.E.M of Adiponectin in controls (C) and non obese patients groups T2D (G1), CAD (G2),and T2D+CAD (G3)] were 15.43± 0.57, 10.86 ±0.45, 11.29±0.4, and 11.158±0.68 ng/ml respectively with ranges of 10.8-

20.6, 8.14-12.7, 7.86-14.62, and 7.7-14.38 ng/ml (Table 1). The result showed that the total serum Adiponectin levels were lower than the upper limit in 10 cases (100%), 19 cases (100%) and 10 cases (100%) in group (G1, G2) and (G3) respectively. The statistical analysis revealed highly significant decrease in non obese groups (G1, G2 and G3) as compared to controls ( $P < 0.001$ ). However, there was no statistical variation between non Obese groups when compared to each others (Table 2a).

**In obese groups**

The results revealed that the mean values  $\pm$  S.E.M of Adiponectin in controls (C), obese patient groups [T2D (G4), CAD (G5), and T2D+CAD (G6)] were  $15.43 \pm 0.57$ ,  $9.99 \pm 0.71$ ,  $11.69 \pm 0.67$ , and  $9.9 \pm 0.4$  ng/ml with ranges of 10.8-20.6, 5.18-15.91, 8.3-14.4, and 7.9-12 ng/ml respectively (Table 1a)

The result showed that the total serum Adiponectin levels were lower than the upper limit in 19 cases (100%) 10 cases (100%) and 13 cases (100%) in group G4, G5 and G6. The statistical analysis revealed high significant decrease in obese groups (G4, G5, and G6) when compared to controls ( $P < 0.001$ ). However, there were no statistical variations between obese groups when compared to each others.

**-Serum Leptin:**

**In non Obese**

The results revealed that the mean values  $\pm$  S.E.M of Leptin in controls (C), T2D (G1), CAD (G2), and T2D+CAD (G3) were  $3.87 \pm 0.19$ ,  $5.66 \pm 0.29$ ,  $5.92 \pm 0.45$ , and  $6.645 \pm 0.57$  ng/ml with ranges of 2.17-5.6, 4.1-6.9, 3.4-9.88, and 4.18-9.22 ng/ml. respectively (Table 1a)

The result showed that serum Leptin levels were higher than the upper limit in 5 cases (50%), 10 cases (53%) and 6 cases (32%) in group G1, G2 and G3 respectively.

The statistical analysis revealed highly significant increase in non Obese groups (G1, G2, and G3) as compared to controls ( $P < 0.001$ ). However, there was no statistical variation between obese and non obese.

**In Obese**

The results revealed that the mean values  $\pm$  S.E.M of Leptin in controls (C), obese T2D (G4), obese CAD (G5), and Obese T2D+CAD (G6) were,  $3.87 \pm 0.19$ ,  $5.05 \pm 0.41$ ,  $6.56 \pm 0.99$  and  $7.8 \pm 0.40$  ng/ml respectively with ranges of 2.17-5.6, 2.43-9, 2.36-11.84, and 5.2-10.04 ng/ml (Table 1a). Serum Leptin levels were higher than the upper limit in 11 cases (85%) in group (G4), 5 cases (50%) in group (G5), 7 cases (36%) in group (G6). The statistical analysis revealed high significant increase in obese groups (G4, G5, and G6) as compared to controls ( $P < 0.001$ ). Moreover, there

was statistical increase between (G4) and (G5) groups when compared to each others, but there was no significant variation between (G5) and (G6) groups when compared to each others ( $P > 0.05$ ) (Table variation between non Obese groups when compared to each others (Table 2a).

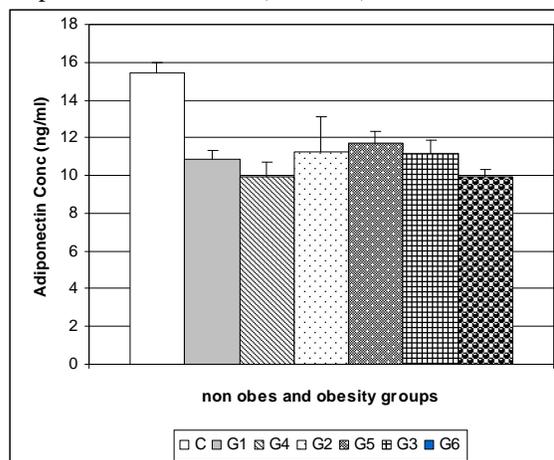


Fig. 1. Adiponectin Conc. (ng/ml) in Different Studied Groups (non obese and obese groups).  
 C: Controls; G1: T2DM (non obese); G4: T2DM (obese); G2: CAD (non obese); G5: CAD (obese); G3: T2DM+CAD (non obese); G6: T2DM+CAD (obese)

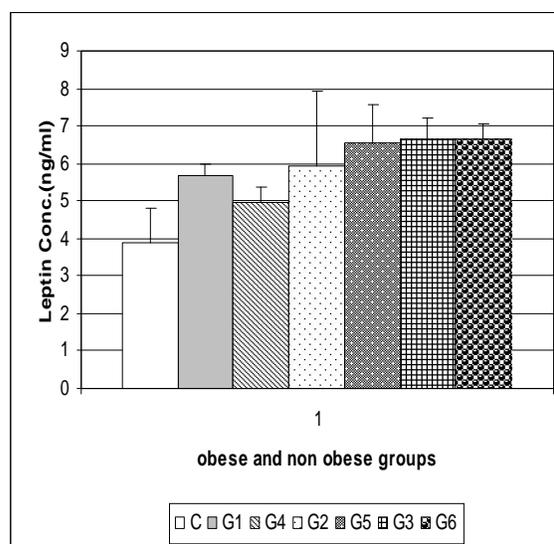


Fig. 2. Leptin Conc. (ng/ml) in Different Studied Groups (non obese and obese groups).  
 C: Controls; G1: T2DM (non obese); G4: T2DM (obese); G2: CAD (non obese); G5: CAD (obese); G3: T2DM+CAD (non obese); G6: T2DM+CAD (obese)

Table (1a). Collective Data for Serum Adipokines (Adiponectin, Leptin) in non Obese, obese and Healthy Control Groups

Parameters	Groups	Controls (C)	T2DM		CAD		T2DM +CAD	
			Non-obese (G1)	Obese (G4)	Non-obese (G2)	Obese (G5)	Non-obese (G3)	Obese (G6)
BMI	N	25	10	19	19	10	10	13
	Range	21.70- 25.10	20.00 -29.50	5.18-15.91	22.00-30.00	31.00-36.00	25.00-30.00	31.51-44.00
	Mean	23.0720	26.0500	9.9579	26.3842	32.6000	27.9330	36.0438
	S.D.	1.04184	2.96695	3.25528	2.13027	1.71270	1.87806	3.88264
	S.E.	.20837	.93823	.74681	.48872	.54160	.59389	1.07685
Adiponectin	N	25	10	19	19	10	10	13
	Range	10.80-20.60	8.14-12.71	2.43-9.00	7.86- 14.62	8.30-14.40	7.68-14.38	7.90-12.00
	Mean	15.4300	10.8560	4.9358	11.2858	11.6900	11.1579	9.9231
	S.D.	2.86669	1.41221	1.81995	1.82027	2.11474	2.14920	1.35040
	S.E.	.57334	.44658	.41752	.41760	.66874	.67964	.37453
Leptin	N	25	10	19	19	10	10	13
	Range	2.17-5.60	4.10-6.93	2.43-9.00	3.40-9.88	2.36-11.84	4.18-9.22	5.20-10.04-
	Mean	3.8696	5.6901	4.9358	5.9247	6.5610	6.6450	7.8477
	S.D.	.95263	.92908	1.81995	1.97883	3.13274	1.79425	1.52043
	S.E.	.19053	.29380	.41752	.45397	.99066	.56739	.42169

Table (1b). Collective Data for Serum Glucose, Insulin and HOMA-IR in non Obese, obese and Healthy Control Groups

Parameters	Groups	Controls (C)	T2DM		CAD		T2DM +CAD	
			Non-obese (G1)	Obese (G4)	Non-obese (G2)	Obese (G5)	Non-obese (G1)	Obese (G4)
Glucose	N	25	10	19	19	10	10	13
	Range	75.00-105.00	123.00-311.00	110.00-357.00	82.00-127.00	75.00-114.78	123.00-228.00	141.25-265.00
	Mean	86.2000	198.6000	230.3684	99.6316	93.1780	169.5000	184.8715
	S.D.	9.21954	51.84421	67.55344	12.20751	12.49912	33.67904	40.68162
	S.E.	1.84391	16.39458	15.49782	2.80060	3.95257	10.65025	11.28305
Insulin	N	25	10	19	19	10	10	13
	Range	3.60-8.90	6.11-19.21	2.70- 10.90	1.09-17.66	4.30-55.31	5.24-17.72	6.42-27.21
	Mean	5.4904	10.1360	5.9842	7.8221	11.6450	11.3830	16.5623
	S.D.	1.48685	4.22863	2.35118	4.81128	15.47841	3.72399	5.92084
	S.E.	.29737	1.33721	.53940	1.10378	4.89470	1.17763	1.64214
HOMA-IR	N	25	10	19	19	10	10	13
	Range	.70-2.20	2.80-9.60	2.70-10.90	.30-4.10	.80-11.50	2.30	4.00
	Mean	1.1840	4.8500	5.9842	1.8632	2.5900	4.7800	7.4923
	S.D.	.41501	2.18187	2.35118	1.12852	3.19668	1.94867	3.43765
	S.E.	.08300	.68997	.53940	.25890	1.01088	.61623	.95343

Table 2 (a): Multiple Comparisons (Post Hoc Tests) Between Age, BMI, Adiponectin and Leptin in non obese Patients and healthy control Groups

Dependent Variable	(I) DIAGNOSIS	(J) DIAGNOSIS	Mean Difference (I-J)	Std. Error	Sig.	95% Confidence Interval	
						Lower Bound	Upper Bound
BMI	C	G1	<b>-2.9780*</b>	.71436	.000	-4.4069	-1.5491
		G2	<b>-3.3122*</b>	.58108	.000	-4.4745	-2.1499
		G3	<b>-4.8610*</b>	.71436	.000	-6.2899	-3.4321
	G1	C	<b>2.9780*</b>	.71436	.000	1.5491	4.4069
		G2	-.3342	.74589	.656	-1.8262	1.1578
		G3	<b>-1.8830*</b>	.85382	.031	-3.5909	-.1751
	G2	C	<b>3.3122*</b>	.58108	.000	2.1499	4.4745
		G1	.3342	.74589	.656	-1.1578	1.8262
		G3	<b>-1.5488*</b>	.74589	.042	-3.0408	-.0568
	G3	C	<b>4.8610*</b>	.71436	.000	3.4321	6.2899

ADIPONEC	C	G1	<b>1.8830*</b>	.85382	<b>.031</b>	.1751	3.5909	
		G2	<b>1.5488*</b>	.74589	<b>.042</b>	.0568	3.0408	
		G3	<b>4.2721*</b>	.92511	<b>.000</b>	2.4051	6.1391	
	G3	G1	<b>4.5740*</b>	.92511	<b>.000</b>	2.7070	6.4410	
		G2	<b>4.1442*</b>	.69890	<b>.000</b>	2.7462	5.5422	
		C	<b>-4.2721*</b>	.92511	<b>.000</b>	-6.1391	-2.4051	
	G1	G1	.3019	1.10572	.786	-1.9295	2.5333	
		G2	-.1279	.89714	.887	-1.9224	1.6667	
		C	<b>-4.5740*</b>	.92511	<b>.000</b>	-6.4410	-2.7070	
	G2	G2	-.4298	.89714	.634	-2.2243	1.3648	
		G3	-.3019	1.10572	.786	-2.5333	1.9295	
		C	<b>2.0551*</b>	.44628	<b>.000</b>	1.1624	2.9478	
	LEPTIN	C	G2	.2346	.57286	.684	-.9113	1.3805
			G3	-.7203	.57286	.214	-1.8662	.4256
			G3	<b>-2.7754*</b>	.44167	<b>.000</b>	-3.6667	-1.8841
G3		G2	<b>-2.0551*</b>	.44628	<b>.000</b>	-2.9478	-1.1624	
		G1	<b>-1.8205</b>	.44167	<b>.000</b>	-2.7118	-.9292	
		C1	<b>2.7754*</b>	.44167	<b>.000</b>	1.8841	3.6667	
G1		G1	.9549	.52790	.078	-.1104	2.0202	
		G2	.7203	.57286	.214	-.4256	1.8662	
		C	<b>1.8205*</b>	.44167	<b>.000</b>	.9292	2.7118	
G2		G3	-.9549	.52790	.078	-2.0202	.1104	
		G3	-.2346	.57286	.684	-1.3805	.9113	
		C	<b>2.0551*</b>	.44628	<b>.000</b>	1.1624	2.9478	
G3		G1	.2346	.57286	.684	-.9113	1.3805	
		G2	-.7203	.57286	.214	-1.8662	.4256	
		G3	-.7203	.57286	.214	-1.8662	.4256	

Based on observed means.

\*The mean difference is significant at the .05 level.

**N.B.:** C=controls  
G2 = CAD

G1= T2DM  
G3= CAD+T2DM

Table 2 (b): Multiple Comparisons (Post Hoc Tests) Between Glucose, Insulin and HOMA.IR in non obese Patients and healthy control Groups

Dependent Variable	(I) DIAGNOSIS	(J) DIAGNOSI	Mean Difference (I-J)	Std. Error	Sig.	95% Confidence Interval	
						Lower Bound	Upper Bound
GLUCOSE	C	G1	<b>-112.400*</b>	9.55422	<b>.000</b>	-131.5113	-93.2887
		G2	-13.4316	7.77160	.089	-28.9771	2.1139
		G3	<b>-83.3000*</b>	9.55422	<b>.000</b>	-102.4113	-64.1887
	G1	C	<b>112.4000*</b>	9.55422	<b>.000</b>	93.2887	131.5113
		G2	<b>98.9684*</b>	9.97593	<b>.000</b>	79.0136	118.9232
		G3	<b>29.1000*</b>	11.41947	<b>.013</b>	6.2576	51.9424
	G2	C	13.4316	7.77160	.089	-2.1139	28.9771
		G1	<b>-98.9684*</b>	9.97593	<b>.000</b>	-118.9232	-79.0136
		G3	<b>-69.8684*</b>	9.97593	<b>.000</b>	-89.8232	-49.9136
	G3	C	<b>83.3000*</b>	9.55422	<b>.000</b>	64.1887	102.4113
		G1	<b>-29.1000*</b>	11.41947	<b>.013</b>	-51.9424	-6.2576
		G2	<b>69.8684*</b>	9.97593	<b>.000</b>	49.9136	89.8232
INSULIN	C	G1	<b>-4.6456*</b>	1.32769	<b>.001</b>	-7.3014	-1.9898
		G2	<b>-2.3317*</b>	1.07997	<b>.035</b>	-4.4920	-.1714
		G3	<b>-5.8926*</b>	1.32769	<b>.000</b>	-8.5484	-3.2368
	G1	C	<b>4.6456*</b>	1.32769	<b>.001</b>	1.9898	7.3014
		G2	2.3139	1.38630	.100	-.4591	5.0869
		G3	-1.2470	1.58690	.435	-4.4213	1.9273
	G2	C	<b>2.3317*</b>	1.07997	<b>.035</b>	.1714	4.4920
		G1	-2.3139	1.38630	.100	-5.0869	.4591
		G3	<b>-3.5609*</b>	1.38630	<b>.013</b>	-6.3339	-.7879
	G3	C	<b>5.8926*</b>	1.32769	<b>.000</b>	3.2368	8.5484
		G1	1.2470	1.58690	.435	-1.9273	4.4213
		G2	<b>3.5609*</b>	1.38630	<b>.013</b>	.7879	6.3339
HOMA	C	G3	<b>-3.5960</b>	.52011	<b>.000</b>	-4.6456	-2.5464
		G1	<b>-3.6660</b>	.52011	<b>.000</b>	-4.7156	-2.6164
		G2	-.6792	.40085	.095	-1.4810	.1227
	G3	C	<b>3.5960</b>	.52011	<b>.000</b>	2.5464	4.6456
		G1	-.0700	.62165	.911	-1.3245	1.1845
		G2	<b>2.9168*</b>	.51455	<b>.000</b>	1.8876	3.9461

G1	C	<b>3.6660</b>	.52011	<b>.000</b>	2.6164	4.7156
	G2	<b>2.9868*</b>	.51455	<b>.000</b>	1.9576	4.0161
	G3	.0700	.62165	.911	-1.1845	1.3245
G2	C	.6792	.40085	.095	-.1227	1.4810
	G1	<b>-2.9868*</b>	.51455	<b>.000</b>	-4.0161	-1.9576
	G3	<b>-2.9168*</b>	.51455	<b>.000</b>	-3.9461	-1.8876

Based on observed means.

\*The mean difference is significant at the .05 level.

**N.B.:** C=controls  
G2 = CAD

**G1= T2DM**  
**G3= CAD+T2DM**

Table 3 (a). Univariate analysis of correlation between Adipokines (Adiponectin & Leptin) in T2DM groups and other variables (all results are two-tailed)

Variable	Adiponectin				Leptin				HOMA-IR			
	Non obese		obese		Non obese		obese		Non obese		obese	
	R	P value	R	P value	R	P value	R	P value	R	P value	R	P value
BM	-.491	.149	.373	.116	.470	.171	.269	.265	<b>.698*</b>	<b>.025</b>	.427	.069
Adiponectin	1	.	1	.	.228	.526	<b>.523*</b>	<b>.021</b>	-.472	.168	.109	.658
Leptin	.228	.526	<b>.523*</b>	<b>.021</b>	1	.	1	.	-.087	.812	.431	.066
Glucose	-.062	.866	-.004	.988	-.185	.608	.142	.561	.424	.222	.412	.080
Insulin	-.349	.323	.102	.676	.081	.823	.218	.370	<b>.786**</b>	<b>.007</b>	<b>.594**</b>	.007
HOMA-IR	-.472	.168	.109	.658	-.087	.812	.431	.066	1	.	1	.

Table 3 (b). Univariate analysis of correlation between Adipokines (Adiponectin & Leptin) in CAD groups and other variables (all results are two-tailed)

Variable	Adiponectin				Leptin				HOMA-IR			
	Non obese		obese		Non obese		obese		Non obese		obese	
	R	P value	R	P value	R	P value	R	P value	R	P value	R	P value
BM	-.328	.170	.014	.969	.038	.876	-.121	.738	.215	.377	-.200	.580
Adiponectin	1	.	1	.	.443	.058	.378	.282	.224	.356	.539	.108
Leptin	.443	.058	.378	.282	1	.	1	.	.418	.075	.246	.493
Glucose	-.250	.303			-	<b>.604*</b>	-.206	.569	-.326	.173	.780	-.102
Insulin	.247	.308	.514	.128	<b>.499*</b>	<b>.030</b>	.272	.447	<b>.985**</b>	<b>.000</b>	.996	.000
HOMA-IR	.224	.356	.539	.108	.418	.075	.246	.493	1	.	1	.

Table 3 (C). Univariate analysis of correlation between Adipokines (Adiponectin & Leptin) in T2DM+CAD groups and other variables (all results are two-tailed)

Variable	Adiponectin				Leptin				HOMA-IR			
	Non obese		obese		Non obese		obese		Non obese		obese	
	R	P value	R	P value	R	P value	R	P value	R	P value	R	P value
BM	.533	.113	-.094	.759	-.222	.538	.113	.713	<b>-.716*</b>	<b>.020</b>	-.202	.508
Adiponectin	1	.	1	.	.376	.285	<b>.660*</b>	<b>.020</b>	-.288	.420	-.202	.509
Leptin	.376	.285	<b>.660*</b>	<b>.020</b>	1	.	1	.	.517	.126	-.046	.882
Glucose	.165	.649	.253	.404	.565	.089	.273	.367	.524	.120	.367	.218
Insulin	-.340	.336	.265	.509	.382	.276	-.197	.520	<b>0.902**</b>	<b>.000</b>	<b>0.868*</b>	<b>.000</b>
HOMA-IR	-.288	.420	-.202	.509	.517	.126	-.046	.882	1	.	1	.

\* Correlation is significant at the 0.05 level (2-tailed).

\*\* Correlation is significant at the 0.01 level (2-tailed).

**HOMA-IR Ratio:**

**In non obese**

The results revealed that the mean values  $\pm$  S.E.M of HOMA-IR Ratio in controls (C),T2D (G1), CAD (G2),and T2D+CAD (G3) were  $1.8\pm 0.082$ ,  $4.85\pm 0.69$ ,  $1.86\pm 0.26$ ,and  $4.78\pm 0.62$  with ranges of, 0.7-2.2, 6.1-19.21, 0.3-4.1,and 2.3-8.2 respectively (Table 1b). The result showed that the HOMA-IR Ratio was higher than the upper limit in 10 cases (100%) ,6 cases (32%) and 6 cases (60%) in group G1,G2and G3 respectively. The statistical analysis revealed highly significant increase in non obese groups (G1,G2and G3) as compared to both controls and each other ( $P<0.001$ ).(Table 2b).

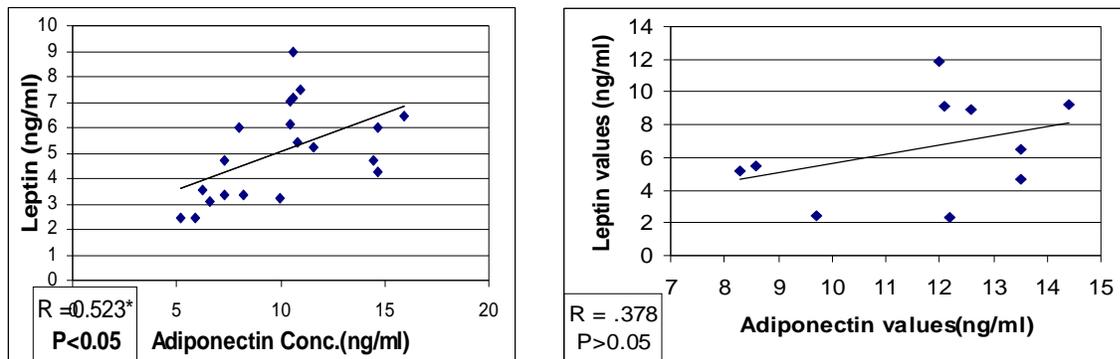
**In Obese**

The results revealed that the mean values  $\pm$  S.E.M of HOMA-IR Ratio in controls (C),T2D (G4), CAD (G5),and T2D+CAD (G6) were  $1.18\pm 0.082$ ,  $5.96\pm 0.51$ ,  $2.59\pm 1.01$ ,and  $7.5\pm 1.0$  respectively with ranges of, 0.7-2.2 , 2.7-10.9,0.8-11.5,and 4-16.5. The result showed that the HOMA-IR Ratio was higher than the upper limit in 19 cases (100%) in group (G4),2 cases (20%) in group (G5),and 13 cases (100%) in group (G6).

The statistical analysis revealed highly significant increase in the two obese groups only (G4, and G6) as compared to controls ( $P<0.001$ ). While there was no significant variation in (G5) when compared to (C) ( $P>0.05$ ).Moreover, there was a significant increase in all obese groups when compared to each others ( $P<0.05$ ). The current study showed also a significant correlation between serum adiponectin and leptin in obese DM patients (Figure2).

**Relationship between adipocytokines and other parameters**

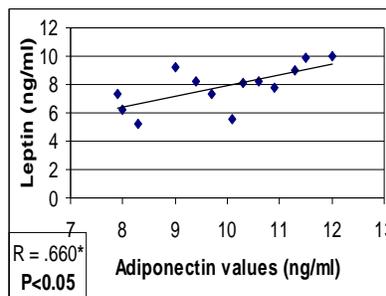
Data on the relationship between Adiponectin and the other variables in non obese and obese patient groups are shown in Tables (2a, b, c). Serum Adiponectin levels were poor associated with Leptin CAD (obese)  $\{r=0.378$ ,  $P=0.282$  (Table (3a, C) & Fig 3) Whereas Adiponectin were associated with Leptin in both of obese T2DM without and CAD ( $r=0.523$ ;  $P=0.0021$ ;  $r=0.660$ ;  $P=0.020$ ) respectively (Table 3b & Fig 3). Leptin Levels were good associated with Insulin in CAD (non obese) ( $r=0.499$ ,  $P<0.03$ ).



**(G4)T2DM (Obese)**

**(G5) CAD (obese)**

Fig 3a. Correlation between Adiponectin and Leptin in obese T2DM and CAD Groups.



**(G6) T2DM+CAD (obese)**

Fig 3b. Correlation between Adiponectin and Leptin in obese T2DM+ CAD Group. Correlation is significant at the 0.05 level.

## Discussion

The current study demonstrated that serum leptin concentration was significantly higher in obese DM + CAD patients than obese patients with CAD alone. These findings suggested that the hyperinsulinaemia that accompanies obesity and diabetes in our DM+CAD patients probably results in increased *ob* gene expression and higher plasma leptin concentration. This agreed with previous studies that showed that leptin is part of the adipoinular axis, as insulin stimulates leptin production in adipocytes and leptin interacts with the pancreatic B-cell through various effects on insulin secretion (Bullo et al. 2002). It has been shown in animal studies, insulin increases the expression of the *ob* gene and results in increased leptin concentration, while relative or absolute deficiency of insulin results in reduced *ob* gene expression (Friedman and Halaas 1998).

The increased leptin levels in obese DM+CAD, obese DM, and obese CAD patients in current study might represent an integrated marker of adiposity, insulin resistance, and vascular dysfunction in Egyptian patients. This integrity agrees with previous studies by (Wallace et al. 2001) who postulated that hyperleptinemia has been recognized as a mediator between obesity and cardiovascular disease. The effect of obesity and high leptin levels on vascular system likely due to several various factors such as increased sympathetic activity, enhanced platelet aggregation, increased oxidative stress, and cardiac hypertrophy (Konstantinides et al. 2001; Bodary et al. 2002; Paolisso et al. 1999). Besides, increased adipose tissue in obesity requires an increased vascular bed to maintain its baseline circulation (Bouloumie et al. 1998; Sierra-Honigmann et al. 1998). Thus this adaptation may, conversely, promote arteriosclerosis over long periods of time.

Higher leptin concentrations were shown to be associated with an impaired arterial distensibility (Singhal et al. 2002), and patients with a restenosis after coronary stenting had higher leptin levels than those without a restenosis (Piatti et al. 2003). Furthermore, the direct influence of leptin on the vascular biology is supported by the *ob/ob* mice, which lacks leptin and consequently becomes hyperphagia and obesity but is nevertheless resistant to atherosclerosis (Schafer et al. 2004). However, the administration of leptin removes this protective effect

against atherosclerosis. The atherosclerosis risk in heterozygote and the control mouse, which suggest a dose-response relation between the leptin levels and the atherosclerotic process (Nishina et al. 1994).

Our findings also showed increased levels of insulin in studied obese patients. The increase in leptin levels in these groups did not show significant correlation with insulin resistance. This agreed with evidence that plasma leptin may vary much more as a function of the circulating insulin concentrations than of the degree of the insulin resistance itself (Mohamed-Ali et al. 1997; Lonnqvist et al. 1995). The current study also showed a significant decrease in serum adiponectin levels in obese and non obese CAD, and DM+CAD patients compared to controls, consistent with our findings, previous studies that showed a decreased levels of adiponectin in a variety of insulin-resistant states, including obesity, diabetes, and cardiovascular diseases (Hotta et al. 2000; Weyer et al. 2001; Kumada et al. 2003; Trujillo and Scherer 2005).

There were some possibilities explained the decrease in serum levels of adiponectin in different studied obese and non obese groups. One of them is elevated insulin levels in diabetic subjects may have been responsible for the decreased adiponectin concentrations as insulin regulates the secretion of various proteins from adipose tissue (Yu et al. 2002; Mohlig et al. 2002). There is evidence that insulin may have direct effect on adiponectin gene expression and adiponectin concentrations in vitro (Fasshauer et al. 2002; Halleux et al. 2001; Motoshima et al. 2002). Another possibility was that accumulation of adiponectin in atherosclerotic vascular walls may accelerate its half-life in plasma, resulting in the reduction of the plasma concentration of adiponectin in subjects with CAD (Hotta et al. 2000). The current study showed significant correlation between serum adiponectin and leptin in obese DM patients, similar results were obtained by Satoh et al. 2004; Kotani et al. 2005).

## Conclusion

The coexistence of a correlation between serum leptin and Adiponectin levels in addition to increase of serum leptin and decrease serum Adiponectin levels in obese DM patients in the current study, support that hypothesis of their susceptibility to atherosclerosis. Although the small sample size does

not enable us to make a definitive conclusion, our study revealed that there were significant differences in plasma levels of Adiponectin between CAD patients with and without DM. DM has an additional effect on CAD, that of decreasing plasma Adiponectin levels. We speculate that people who have very low plasma Adiponectin levels may be at increased risk of developing both CAD and DM.

#### Acknowledgement:

The authors would like to thank Dept. of cardiac catheterization in the National Institute of Heart Disease especially Prof. Dr. Mohamed Badrawy and his staff for the clinical investigation of the patients and for providing the blood samples for the research work.

#### Corresponding Author:

Dr. Nervana Samy  
Department of Biochemistry  
Genetic Engineering & Biotechnology Division.  
National Research Centre  
Dokki, Giza, Egypt  
e.mail: [Nervana91@hotmail.com](mailto:Nervana91@hotmail.com)

#### References

1. Alberti KG, Zimmet PZ. Definition, diagnosis and classification of diabetes mellitus and its complications. Part 1: diagnosis and classification of diabetes mellitus provisional report of a WHO consultation. *Diabet Med.* 1998; 15(7):539-53.
2. Arita Y, Kihara S, Ouchi N, Takahashi M, Maeda K, Miyagawa J, Hotta K, Shimomura I, Nakamura T, Miyaoaka K, Kuriyama H, Nishida M, Yamashita S, Okubo K, Matsubara K, Muraguchi M, Ohmoto Y, Funahashi T, Matsuzawa Y. Paradoxical decrease of an adipose-specific protein, adiponectin, in obesity. *Biochem Biophys Res Commun* 1999; 257:79–83.
3. Arner P. Insulin resistance in type 2 diabetes—role of the adipokines. *Curr Mol Med*; 2005; 5:333–9.
4. Azuma K., Heilbronn L. K., Albu J. B., Smith S. R., Ravussin E., Kelley D. E. and the Look AHEAD Adipose Research Group.. Translational Physiology: Adipose tissue distribution in relation to insulin resistance in type 2 Diabetes mellitus. *Am J Physiol Endocrinol Metab* 2007; 293: E435–E442
5. Behre CJ. Adiponectin, obesity and atherosclerosis. *Scand J Clin Lab Invest*; 2007; 67:449–58.
6. Bloomgarden ZT. Adiposity and diabetes. *Diabetes Care* 2002.;25:2342–9.
7. Bodary, P. F., Westrick R. J., Wickenheiser K. J., Shen Y, and. Eitzman D. T. Effect of leptin on arterial thrombosis following vascular injury in mice. *JAMA* 2002.;287 (13):1706-1709.
8. Bouloumie, A., Drexler H. C., Lafontan M., and Busse R.. Leptin ,the product of Ob gene, promotes angiogenesis. *Circ Res.* 1998; 83 (10):1059-1066.
9. Bullo, M., Garcia-Lorda P., and Salas-Salvado J.. Plasma soluble tumor necrosis factor alpha receptors and leptin levels in normal-weight and obese women: effect of adiposity and diabetes. *Eur J Endocrinol* 2002. 146 (3):325-331.
10. Caro JF, Sinha MK, Kolaczynski JW, Zhang PL, Considine RVLeptin: the tale of an obesity gene. *Diabetes.* 1996.;45:1455–1462.
11. Chu NF, Spiegelman D, Rifai N, Hotamisligil GS, Rimm EB. Glycemic status and soluble tumor necrosis factor receptor levels in relation to plasma leptin concentrations among normal weight and overweight US men. *Int J Obes Relat Metab Disord* 2000; 24:1085–92.
12. Cooper M, Bonnet F, Oldfield M and Jandeleit K: Mechanism of diabetic vasculopathy: an overview. *Am. J. Hypertension.* May; 2001; 14(5 Pt 1): 475-86.
13. Fantuzzi G. Adipose tissue, adipokines, and inflammation. *J.Allergy Clin Immunol*; 2005; 115:911—9.
14. Fasshauer, M., Klein J., Neumann S., Eszlinger M., and Paschke R.. Hormonal regulation of adiponectin gene expression in 3T3-L1 adipocytes. *Biochem Biophys Res Commun* 2002; 290 (3):1084-1089.
15. Frayn KN. Obesity and metabolic disease: is adipose tissue the culprit? *Proc Nutr. Soc*; 2005; 64:7–13.
16. Friedman, J .M., and Halaas J. L. Leptin and the regulation of body weight in mammals. *Nature* 1998; 395 (6704):763-770.
17. Grundy SM. Obesity, metabolic syndrome, and coronary atherosclerosis. *Circulation*; 2002; 105:2696–8.
18. Halleux, C. M., Takahashi M., Delporte M. L., Detry R., Funahashi T., Matsuzawa Y., and Brichard S. M.. Secretion of adiponectin and regulation of apM1 gene expression in human visceral adipose tissue. *Biochem Biophys Res Commun* 2001; 288 (5):1102-1107.
19. Hamann A, Matthaei S. Regulation of energy balance by leptin. *Exp Clin Endocrinol Diabetes*; 1996; 104:293–300.
20. Haynes WG. Role of leptin in obesity-related hypertension. *Exp Physiol*; 2005; 90:683– 8.
21. Hotamisligil GS, Murray DL, Choy LN, Spiegelman BM. TNF-a inhibits signaling from

- the insulin receptor. *Proc Natl Acad Sci U S A.*; 1994; 91:4854–4858.
22. Hotamisligil GS, Shargill NS, Spiegelman BM. Adipose expression of tumor necrosis factor- $\alpha$ : direct role in obesity-linked insulin resistance. *Science*. 1993; 259:87–91.
  23. Hotamisligil GS, Spiegelman BM. Tumor necrosis factor  $\alpha$ : a key component of the obesity-diabetes link. *Diabetes* 1994; 43:1271–1278.
  24. Hotta, K., Funahashi T., Arita Y., Takahashi M., Matsuda M., Okamoto Y., Iwahashi H., Kuriyama H., Ouchi N., Maeda K., Nishida M., Kihara S., Sakai N., Nakajima T., Hasegawa K., Muraguchi M., Ohmoto Y., Nakamura T., Yamashita S., Hanafusa T., and Matsuzawa Y.. Plasma concentrations of a novel, adipose-specific protein, adiponectin, in type 2 diabetic patients. *Arterioscler Thromb Vasc Biol* 2000; 20 :1599-1595:(6)
  25. Jazet IM, Pijl H, Meinders AE, Adipose tissue as an endocrine organ: impact on insulin resistance. *Neth J Med* 2003; 61:194–212
  26. Kershaw EE, Flier JS. Adipose tissue as an endocrine organ. *J.Clin Endocrinol Metab* 2004; 89:2548—56.
  27. Koerner A, Kratzsch J, Kiess W. Adipocytokines: leptin—the classical, resistin—the controversial, adiponectin—the promising, and more to come. *Best Pract Res Clin Endocrinol Metab* 2005; 19:525– 46.
  28. Konstantinides, S., Schafer K., Koschnick S., and Loskutoff D. J. Leptin-dependent platelet aggregation and arterial thrombosis suggests a mechanism for atherothrombotic disease in obesity. *J Clin Invest* 2001; 108 (10):1533-1540.
  29. Kotani, K., Sakane N., Saiga K., and Kurozawa Y. Leptin : adiponectin ratio as an atherosclerotic index in patients with type 2 diabetes : relationship of the index to carotid intima-media thickness. *Diabetologia* 2005; 48 (12):2684-2686.
  30. Kowalska I. Role of adipose tissue in the development of vascular complications in type 2 diabetes mellitus. *Diabetes Research and Clinical Practice* 2007; 78S : S14–S22
  31. Kumada, M., Kihara S., Sumitsuji S., Kawamoto T., Matsumoto S., Ouchi N., Arita Y., Okamoto Y., Shimomura I., Hiraoka H., Nakamura T., Funahashi T., and Matsuzawa Y. Association of hypoadiponectinemia with coronary artery disease in men. *Arterioscler Thromb Vasc Biol* .2003 ; 23 (1.89-85:(
  32. Lawlor DA, Davey Smith G, Ebrahim S, Thompson C, Sattar N. Plasma adiponectin levels are associated with insulin resistance, but do not predict future risk of coronary heart disease in women. *J Clin Endocrinol Metab* 2005; 90:5677–83.
  33. Lindsay RS, Resnick HE, Zhu J, Tun M. L., Howard B. V., Zhang Y., Yeh J. and Lyle G. Adiponectin and coronary heart disease: the Strong Heart Study. *Arterioscler Thromb Vasc Biol*. 2005; 25:e15– 6.
  34. Lonnqvist, F., Arner P., Nordfors L., and Schalling M. Overexpression of the obese (ob) gene in adipose tissue of human obese subjects. *Nat Med* 1995 (9):950-953.
  35. Marcus MA, Murphy L, Pi-Sunyer FX, Albu JB. Insulin sensitivity and serum triglyceride level in obese white and black women: relationship to visceral and truncal subcutaneous fat. *Metabolism* 1999; 48: 194–199
  36. Marinou K., Tousoulis D. , Antonopoulos A. S., Stefanad E. I, Stefanadis C. Review Obesity and cardiovascular disease: From pathoph. *Inter J of Cardiology*. IJCA- 2009; 11988; 6-14.
  37. Matthews D, Hosker J, Rdenski A, Naylor B .Treacher D and Turner R. Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man, *Diabetologia* 1985; 28:412–419.
  38. McGill Jr HC, McMahan CA, Herderick EE, Zieske A. W. ; Malcom G. T.; Tracy R.E.; Strong J. P.; for the Pathobiological Determinants of Atherosclerosis Obesity accelerates the progression of coronary atherosclerosis in young men. *Circulation*; 2002;105(23):2712-8.
  39. Mohamed-Ali, V., J. H. Pinkney, A. Panahloo, S. Goodrick, S. W. Coppack, and J. S .Yudkin. Relationships between plasma leptin and insulin concentrations, but not insulin resistance, in non-insulin-dependent (type 2) diabetes mellitus. *Diabet Med* 1997; 14 (5):376-380.
  40. Mohlig, M., Wegewitz U., Osterhoff M., Isken F., Ristow M., Pfeiffer A. F., and Spranger J. Insulin decrease human adiponectin plasma levels. *Horm Metab Res* 2002; 34 (11-12):655-658.
  41. Motoshima, H., Wu X., Sinha M. K., Hardy V. E., Rosato E. L., Barbot D. J., Rosato F. E., and Goldstein B. J. Differential regulation of adiponectin secretion from cultured human omental and subcutaneous adipocytes: effects of insulin and rosiglitazone. *J Clin Endocrinol Metab* 2002; 87 (12):5662-5667.
  42. Nishina, P. M., Naggert J. K., Verstuyft J., and Paigen B. Atherosclerosis in genetically obese mice: the mutants obese, diabetes, fat, tubby, and lethal yellow. *Metabolism* 1994; 43 (5):554-558.
  43. Paolisso, G., Tagliamonte M. R., Galderisi M., Zito G. A., Petrocelli A., de Divitiis C. Carella, O., and Varricchio M. Plasma leptin level is

- associated with myocardial wall thickness in hypertensive insulin-resistant men. *Hypertension* 34 1999; (5):1047-1052.
44. Piatti, P., Di Mario C., Monti L. D., Fragasso G., Sgura F., Caumo A., Setola E., Lucotti P., Galluccio E., Ronchi C., Origgi A., Zavaroni I., Margonato A., and Colombo A. Association of insulin resistance, hyperleptinemia, and impaired nitric oxide release with in-stent restenosis in patients undergoing coronary stenting. *Circulation* 2003; 108 (17):2074-2081.
  45. Pischon T, Girman CJ, Hotamisligil GS, Rifai N, Hu FB, Rimm EB. Plasma adiponectin levels and risk of myocardial infarction in men. *JAMA* 2004; 291:1730–7.
  46. Qasim A, Mehta N.N., Tadesse M. G., Wolfe M.L., Rhodes T., Girman C., Reilly M. P. Adipokines, Insulin Resistance, and Coronary Artery Calcification. *Journal of the American College of Cardiology* 2008.; 52(3): 231–6.
  47. Reilly MP, Iqbal N, Schutta M, et al. (2004). Plasma leptin levels are associated with coronary atherosclerosis in type 2 diabetes. *J Clin Endocrinol Metab*;89: 3872– 8.
  48. Salas-Salvadó J, Granada M, Bulló M, Corominas A, Casas P, Foz M. Plasma adiponectin distribution in a Mediterranean population and its association with cardiovascular risk factors and metabolic syndrome. *Metabolism*. 2007;56(11):1486-92.
  49. Satoh, N., Naruse M., Usui T., Tagami T., Suganami T., Yamada K., Kuzuya H., Shimatsu A., and Ogawa Y. Leptin-to-adiponectin ratio as a potential atherogenic index in obese type 2 diabetic patients. *Diabetes Care* 2004; 27 (10):2488-2490.
  50. Schafer, K., Halle M., Goeschen C., Dellas C., Pynn M., Loskutoff D. J., and Konstantinides S. Leptin promotes vascular remodeling and neointimal growth in mice. *Arterioscler Thromb Vasc Biol* 2004; 24 (1):112-117.
  51. Schulze MB, Shai I, Rimm EB, Li T, Rifai N, Hu FB. Adiponectin and future coronary heart disease events among men with type 2 diabetes. *Diabetes* 2005; 54:534–9.
  52. Segal KR, Landt M, Klein S. Relationship between insulin sensitivity and plasma leptin concentration in lean and obese men. *Diabetes* 1996;45:988–91.
  53. Sierra-Honigmann, M. R., Nath A. K., Murakami C., Garcia-Cardena G., Papapetropoulos A.; Sessa W. C., Madge L. A., Schechner J. S., Schwabb M. B., Polverini P. J., and Flores-Riveros. J. R. Biological action of leptin as an angiogenic factor. *Science* 1998; 281 (5383):1683-1686.
  54. Singhal, A., Farooqi I. S., Cole T. J., O'Rahilly S., Fewtrell M., Kattenhorn M., Lucas A., and Deanfield J. Influence of leptin on arterial distensibility: a novel link between obesity and cardiovascular disease? *Circulation* 2002; 106 (15):1919-1924.
  55. Thévenod F. Pathophysiology of Diabetes Mellitus Type 2: Roles of Obesity, Insulin Resistance and  $\beta$ -Cell Dysfunction. Masur K, Thévenod F, Z nker KS (eds): *Diabetes and Cancer. Epidemiological Evidence and Molecular Links*. Front Diabetes. Basel, Karger, 2008, 19:1–18
  56. Trujillo, M. E., and Scherer P. E. Adiponectin--journey from an adipocyte secretory protein to biomarker of the metabolic syndrome. *J Intern Med* 2005; 257 (2):167-175.
  57. Wallace AM, McMahon AD, Packard CJ, Kelly A, Shepherd J, Gaw A, Sattar N. (2001). Plasma leptin and the risk of cardiovascular disease in the West Of Scotland Coronary Prevention Study (WOSCOPS). *Circulation*;104:3052-3056..
  58. Weyer, C., T. Funahashi, S. Tanaka, K. Hotta, Y. Matsuzawa, R. E. Pratley, and P. A. Tataranni. Hypoadiponectinemia in obesity and type 2 diabetes: close association with insulin resistance and hyperinsulinemia. *J Clin Endocrinol Metab* 2001; 86 (5):1930-1935.
  59. Zimmet, P., K. G. Alberti, and J. Shaw. Global and societal implications of the diabetes epidemic. *Nature* 2001; 414 (6865):782-787.

3/7/2010