Protective Effect of *Lepidium sativum L*. Seeds Powder and Extract on Hypercholesterolemic Rats

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Abstract: The present study was designed to investigate the effects of Lepidium sativum L (LS) on the nutritional status and some biochemical analyses of serum and liver in hypercholesterolemic rats. Forty-two adult albino male rats Sprague Dawley strain were classified into six groups. One was fed on standard diet and kept as control (-ve) group. The other five hypercholesterolemic rat groups were control (+ve), drug, LS extract, 5 % or 10 % LS powder rat groups. In comparison to control (- ve) group, the control (+ve) group showed a significant higher value of weight gain, feed efficiency ratio (FER), serum cholesterol, triglycerides, LDL-c, VLDL-c, cholesterol/ HDL-c and LDL-c/ HDL-c, serum (AST& ALT) ,creatinine ,urea, liver cholesterol and total lipids but significant decrease in HDL-c, globulin and liver triglycerides .Also, LS extract and 5% LS powder rat groups showed a significant increase in weight gain, cholesterol/ HDL-c and LDL-c/ HDL-c, serum (AST& ALT) however, drug and 10% LS powder rat groups showed a significant increase in cholesterol/ HDL-c. On the other hand, The drug, LS extract, 5% and 10% LS powder rat groups showed a significant increase in serum cholesterol, triglycerides VLDL-c, LDL-c serum creatinine and urea level when compared to control (- ve) group. In comparing with control (+ ve) group, The drug, LS extract, 5% and 10% LS powder rat groups showed a significant lower value of weight gain, feed efficiency ratio, serum cholesterol ,triglycerides VLDL-c , LDL-c level, cholesterol/ HDL-c , LDL-c/ HDL-c , serum (AST& ALT), serum creatinine, urea, liver cholesterol and total lipids with a significant increase in both serum globulin and liver triglycerides.

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Key wards : Lepidium sativum – aqua extract- cholesterol and rats

1. Introduction:

Hypercholesterolemia refers to elevated serum LDL cholesterol or a combination of high levels of LDL cholesterol and triglycerides. Hypercholesterolemia, a significant cardiovascular risk factor, is one of the major oxidative stresses that generate excess of highly reactive free radicals. This exacerbates the development and progression of atherogenesis Hypercholesterolemia increased the risk of increased LDL-c or more occurately LDL-c/ HDL-c ratio. The atherogenic index decreased as a result of the reduction in LDL-c and increment in HDL-c (Durrington, 1995 and Abd El-Ghanny et al., 2007).

Plants still remain a major source for drug discovery in spite of the great development of synthetic molecules. Consequently, the uses of traditional plant extract in the treatment of various diseases have been flourished (Jouad et al., 2001). The garden cress seed oil, Lepidium sativum L.,(LS) is a fast growing annual herb belonging to the Brassicaceae family that is native to Egypt and west Asia The seeds are wildly consumed as salad and spice (Gokavi et al .,2004). Previous studies have demonstrated the protective action of LS against

carcinogenic compounds and growth inhibition of *Pseudomonas aeruginosa*, a bacteria strain with a potent antibiotic resistance (Abuja et al., 2001 and Kassie et al., 2003). LS recommended in the treatment of hypertension, diabetes and renal disease (Kirtkar and Basu 2005 and Tahraoui et al., 2007).

The present study was designed to investigate the effect of powder or aqua extract from Lepidium sativum L. seeds (LS) either on the nutritional status and some biochemical analyses of serum and liver in hypercholesterolemic rats.

2. Materials and methods

I.- Materials:

1- Garden cress (Lepidium sativum L.) seeds:

Lepidium sativum L. seeds (LS) were purchased from Agricultural Research Center. Garden cress seeds were dried with hot air (40–60 °C) and grinded to powder. Garden cress seeds powder was used in preparation of aquatic extract and also added to the diet as 5 % and 10% of the constituent of fiber.

2- Gemfiibrozil capsules:

It was obtained from Amoun Pharmaceutical Industries Company. Each tablet contains 100 mg. It is lipid regulating agent which decrease lipid elevated serum lipids by lowering serum triglycerides and total cholesterol. Human therapeutic dose was 100 mg which converted to rat dose that was 9 mg/kg body weight daily which dissolved in distilled water and given to rats by oral intubations according to Paget and Barnes (1964).

3-Biochemical kits:

BioMeriuex Kits were purchased from Alkan Co. for Chemicals and Biodignostics, Dokki,, Egypt.

4-Experimental animals:

A total of forty-two Sprague –Dawley adult male rats were purchased from the Agricultural Research Center, Giza, Egypt. The average weight was 205 \pm 5 g. The animals were kept under observation for five days before experiment and supplied with standard diet and water *ad libitum*. The standard diet comprised of casein (200g/kg), corn starch (497g/kg), sucrose (100g/kg) , cellulose (30 g/kg) ,corn oil (50g/kg), mineral mixture (100g/kg) , vitamin mixture (20g/kg) and DL-methionine (3g/kg).The standard diet was performed according to NRC (1995).

II. -Methods:

1. Preparation of the garden cress aqueous extract:

The aqueous extract was prepared in a standardized manner by boiling 1 g of dried powdered seeds of LS in 100 ml of distilled water for 10 min and left for 15 min to infuse then cooled and filtered. The filtrate was lyophilized and the desired dose was then prepared and reconstituted in 10 ml of distilled water per kilogram body weight just before oral administration. The aqueous extract dose was 20 mg/kg body weight daily by oral intubations (Eddouks et al., 2005).

2- Grouping of rats and experimental design:

The experimental rats were divided into six groups (n= 7 rats). The first group which kept as normal control (-ve) group which fed on standard diet only. The rest of rats were fed on standard diet with 2 % cholesterol for 3 weeks to be hypercholesterolemic, then classified into 5 groups and remained fed on hypercholeslerolemic diet during the experimental period (8 weeks). One of them acted as control (+ve) and the other groups were drug, LS seeds extract, 5% and 10% LS powder.

3 – Calculation of some parameters:

Feeding and growth performance were carried out by determination of daily feed intake, body weight gain and feed efficiency ratio (FER) according to Chapman et al., (1950) using the following Formula

FER = Body weight gain / Feed intake.

5 –Collection of samples:

The rats were sacrificed at the end of the experiment (8 weeks). The collected blood samples were centrifuged at 3000 rpm/ 10 minutes to obtain serum. Livers of rats were also collected for some biochemical analysis.

5 -Biochemical analysis:

A- Serum analysis

Serum cholesterol, triglycerides (TG) and high density lipoprotein cholesterol (HDL-c) were determined by using enzymatic colorimetric methods (Allain et al., 1974, Buccolo and David (1973) and Kostener, 1977, respectively). Very low density lipoprotein cholesterol (VLDL-c) was calculated as TG/5 while low density lipoprotein cholesterol (LDLc) was calculated as following $[LDL-_C = Total]$ cholesterol -HDL-_C -VLDL-_C] according to Fruchart, (1982). Serum aspartate and alanine amino transferase (AST&ALT) enzymes, total protein, and albumin were estimated according to Reitman and Frankel (1957), Weichselbaum (1946) and Bartholomev and Delany (1966), respectively. Serum globulin (G) value was determined by subtracting the albumin from the total proteins according to Coles (1974). In addition, creatinine and urea were estimated according to Bonsens and Taussky, (1984), and Patton and Crouch, (1977) respectively. Atherogenic indexes (cholesterol/ HDL-c & LDL-c/ HDL-c) were calculated according to Castelli and levitar, (1977).

B- Liver analysis:

Livers of rats were rapidly removed and parts of them perfuse with 50 to 100 of ice cold 0.9%NaCL solution for estimation of cholesterol, triglycerides, and total lipids, according to Abell et al., (1952), Seheletter and Nussel, (1975), Folch et al., (1957), respectively.

III.- Statistical analysis:

Collected data were subjected to analysis according to SPSS Program Differences were considered significant at p < 0.05 (Artimage and Berry, 1987).

3. Results:

Data recorded in table (1)showed that the control (+ve) group showed a significant higher value of weight gain and feed efficiency ratio (PER) at p<0.01 while the LS extract, 5% and 10% LS powder rat groups showed a significant higher value of weight gain at p<0.05 compared to control (- ve) group. The drug, LS extract , 5% and 10% LS powder rat groups showed a significant lower value of weight gain and feed efficiency ratio compared to control (+ ve) group.

Table (2) showed that the control (+ve) group showed a significant increase in serum cholesterol, triglycerides, LDL-c and VLDL-c (p< 0.001) but significant decrease in HDL-c (p<0.001) in comparison with control (-ve). On the other hand, the drug, LS extract , 5% and 10% LS powder rat groups showed a significant increase in serum cholesterol, triglycerides LDL-c and VLDL-c level (p< 0.05&0.01) compared to control (- ve) group. The drug, LS extract , 5% and 10% LS powder rat groups showed a significant decrease in serum cholesterol, triglycerides LDL-c and VLDL-c level (p< 0.05&0.01) compared to control (- ve) group. The drug, LS extract , 5% and 10% LS powder rat groups showed a significant decrease in serum cholesterol, triglycerides LDL-c and VLDL-c level in comparison with control (+ve).

Table (3) showed that, control (+ve), LS extract and 5% LS powder rat groups showed a significant increase in cholesterol/ HDL-c and LDL-c/ HDL-c (p < 0.01 & 0.05) while drug and 10% LS powder rat groups showed a significant increase in cholesterol/ HDL-c (p < 0.05) compared to control (-ve) group. The drug, LS extract, 5% and 10% LS powder rat groups showed a significant decrease in value of cholesterol/ HDL-c and LDL-c/ HDL-c in comparison with control (+ve).

From data presented in table (4), it could be noticed that control (+ve) ,drug, LS extract , and 5% LS powder rat groups showed a significant increase in serum alanine and aspartate aminotransferase (ALT &AST) at p<0.01 compared to control (- ve) group. The drug, LS extract , 5% and10% LS powder rat groups showed a significant decrease in serum alanine and aspartate aminotransferase enzymes in comparison with control (+ ve) group .

As shown in table (5), the control (+ve) showed a significant decrease in globuline (p<0.05) but a significant increase in creatinine and urea (p<0.01) compared to control (- ve) group. The drug, LS extract , 5% and 10% LS powder rat groups showed a significant increase in serum creatinine and urea (p<0.05) compared to control (- ve) group. The drug, LS extract , 5% and 10% LS powder rat groups showed a significant increase in serum globulin and a significant decrease in serum creatinine and urea compared to control (+ ve) group.

As recorded in table (6) showed that control (+ve) group showed a significant increase in liver cholesterol and total lipids (p<0.01) but significant decrease in liver triglycerides (p<0.05) in comparison with control (- ve). The drug, LS extract, 5% and 10% LS powder rat groups showed a non significanjt difference in liver cholesterol, triglycerides and total lipids in comparison with control (- ve) group. The drug, LS extract, 5% and 10% LS powder rat groups showed a significant decrease in liver cholesterol and total lipids in comparison with control (- ve) group. The drug, LS extract, 5% and 10% LS powder rat groups showed a significant decrease in liver cholesterol and total lipids but significant increase in liver triglycerides in comparison with control (+ ve) group.

Table (1): Mean values ± SD of bo	weight gain, feed intake and feed efficiency ratio (FER) of experimental
rat groups.	

Groups		Hypercholesterolemic						
	Control	Control	D		LS			
Variables	(-ve)	(+ ve)	Drug	extract	5%	10%		
Initial	115.33±	116.25±	117.35±	117.50±	116.81±	116.75 ± 4.71^{a}		
weight(g)	3.41 ^a	3.77 ^a	4.12 ^a	4.81 ^a	4.99 ^a			
Final	190.53±	217.42±	198.12±	210.64±	214.01±13	211.78±		
weight(g)	11.22 ^a	10.33 ^a	11.44 ^a	13.77 ^a	.25 ^a	14.61 ^a		
Weight	75.20±	101.17±8.	80.77±	93.14±	97.20±	95.03±		
gain(g)	5.68°	24 ^{a**}	7.69 ^{bc}	8.21 ^{b*}	9.11 ^{b*}	8.61 ^{b*}		
Feed	15.11±	16.22±	15.75±	16.53±	16.31±	16.41±		
intake(g/d)	1.24 ^a	1.18 ^a	2.01 ^a	2.11 ^a	1.89 ^a	1.49 ^a		
FER	0.082 ± 0.003^{b}	$0.103 \pm 0.002^{a^{**}}$	0.085 ± 0.001^{b}	0.093 ± 0.001^{b}	0.099 ± 0.004^{b}	0.096± 0.005 ^b		

Significant with control group * P<0.05 ** P<0.01 *** P<0.001 Mean values in each row having different superscript (a, b &c,) are significant.

Groups		Hypercholesterolemic						
	Control (-ve)	Control	Dmug	LS				
Variables		(+ v e)	Drug	extract	5%	10%		
Cholesterol	80.34±	199.77±11	106.78±10	110.01±	113.24±12	105.35±		
(mg/dl)	7.88 ^c	.43 ^{a****}	.18 ^{b*}	11.33 ^{b*}	.14 ^{b*}	10.61 ^{b*}		
Triglyceride	70.31 ± 6.12 °	155.14±	95.67±	98.01±	96.18±	94.31±		
(mg/dl)		18.48 ^{a***}	10.11 ^{b*}	11.31 ^{b*}	10.15 ^{b*}	9.96 ^{b*}		
HDLc	32.32±	20.11±	29.75±	28.88±	31.14±	30.91±		
(mg/dl)	3.47 ^a	2.87 ^{b***}	2.78 ^a	3.11 ^a	2.99 ^a	3.03 ^a		
LDLc	33.06±	104.01±	57.90±	61.53±	62.87±	55.58 ±		
(mg/dl)	4.01 °	10.22 ^{a***}	6.08 ^{b**}	7.18 ^{b**}	7.33 ^{b**}	6.14 ^{b**}		
VLDLc	14.01±	31.02±	19.13±	19.60±	19.23±	18.86±		
(mg/dl)	1.81 [°]	3.17 ^{a***}	2.15 ^{b*}	2.11 ^{b*}	2.16 ^{b*}	2.22 ^{b*}		

Table (2) The Mean values ± SD of some serum lipid patterns of experimental rat groups.

Significant with control group * P<0.05 ** P<0.01 *** P<0.001 Mean values in each row having different superscript (a, b &c,) are significant.

Table (3) The Me	ean values ±	SD of atherogenic	indexes of e	experimental rat gr	oups.
C					

Groups	Control		Hypercholesterolemic					
Variables	(-ve)	Control		LS				
Variables		(+ ve)	Drug	extract	5%	10%		
Cholesterol/H DLc	2.42± 0.38 [°]	9.93± 1.77 ^{a***}	3.58± 0.77 ^{b*}	3.80± 0.75 ^{b*}	3.63± 0.65 ^{b*}	$3.40 \pm 0.54^{b^*}$		
LDLc/ HDLc	1.05± 0.16°	5.17± 1.14 ^{a***}	1.94± 0.18 ^{bc}	2.13± 0.47 ^{b*}	2.01± 0.32 ^{b*}	1.79± 0.18 ^{bc}		

Significant with control group * P<0.05 ** P<0.01 *** P<0.001

Mean values in each row having different superscript (a, b &c,) are significant.

Table (4) T	he Mean v	values ±	SD of s	erum am	ino transfera	se (ALT	& AST	Γ) enzymes o	of experimental
r	at groups.								

Groups Variables	Control	Hypercholesterolemic						
	(-ve)	Control	D		LS	1		
		(+ ve)	Drug	extract	5%	10%		
ALT (µ /ml)	27.69± 2.81 ^c	49.14± 4.31 ^{a**}	34.41± 3.27 ^{b*}	33.47± 3.61 ^{b*}	34.21± 3.11 ^{b*}	31.78± 3.18 ^{bc}		
AST (µ /ml)	41.27± 4.22 ^C	67.71± 5.91 ^{a**}	51.17± 5.15 ^{b*}	49.39± 5.31 ^{b*}	48.11± 3.92 ^{b*}	40.31± 4.16 [°]		

Significant with control group * P<0.05 ** P<0.01 *** P<0.001 Mean values in each row having different superscript (a, b &c,) are significant.

Groups	Control	Hypercholesterolemic					
	(-ve)	Control	Drug		LS		
Variables		(+ve)	Drug	extract	5%	10%	
T. protein	7.30±	6.11±	6.81±	6.87±	6.91±	6.90±	
(g/dl)	1.21 ^a	1.31 ^a	1.30 ^a	1.41 ^a	1.35 ^a	1.40 ^a	
Albumin	3.71±	3.99±	3.59±	3.24±	3.49 ±	3.11±	
(g/dl)	0.55 ^a	0.45 ^a	0.13 ^a	0.19 ^a	0.23 ^a	0.16 ^a	
Globulin	3.59±	2.12±	3.22±	3.63±	3.42±	3.79±	
(g/dl)	0.22 ^a	0.13 ^{b*}	0.66 ^a	0.45 ^a	0.48^{a}	0.55 ^a	
Creatinine	0.75±	1.01±	0.98±	0.97±	0.95±	0.92±	
(mg/dl)	0.01 ^c	0.15 ^{a**}	0.06^{b^*}	0.11 ^{b*}	0.12 ^{b*}	0.14 ^{b*}	
Urea	40.14±	55.79±	50.17±	49.31±	48.20±	47.32±	
(mg/dl)	4.77 ^c	7.11 ^{a**}	6.18 ^{ab*}	5.11 ^{b*}	4.25 ^{b*}	4.60 ^{b*}	

Table (5) The Mean values ± SD of serum total protein, albumin, globulin, creatinine and urea of experimental rat groups.

Significant with control group * P<0.05 ** P<0.01 *** P<0.001 Mean values in each row having different superscript (a, b &c,) are significant.

Table (6)	The Mean	values	± SD	of liver	cholesterol,	total lipids	and	triglyceride	of experimental r	at
	groups.									

Groups	Control (-ve)	Hypercholesterolemic					
		Control	Drug		LS		
Variables		(+ ve)		extract	5%	10%	
Cholesterol (mg/g)	3.66± 0.88 ^b	6.96± 1.41 ^{a**}	4.01± 0.89 ^b	4.35± 0.95 ^b	4.44± 0.58 ^b	4.25± 0.77 ^b	
Total lipids (mg/g)	34.52± 3.21 ^b	48.99± 4.11 ^{a**}	36.11± 3.41 ^b	38.20± 2.71 ^b	38.77± 2.91 ^b	37.43± 3.11 ^b	
Triglyceride (mg/g)	2.44± 0.11 ^a	$1.65 \pm 0.36^{b^*}$	2.11± 0.33 ^a	2.31± 0.35 ^a	2.01± 0.31 ^a	2.21 ± 0.30^{a}	

Significant with control group * P<0.05 ** P<0.01 *** P<0.001 Mean values in each row having different superscript (a, b &c,) are significant.

4. Discussion:

Our investigation revealed that, consumption of Lepidium sativum L.(LS) seeds increase weight gain as LS seeds are found to contain 18–24% of fat which 34% of total fatty acids is alpha linolenic acid .It contains good amount of lignans and antioxidants, which can stabilize the n-3 polyunsaturated fatty acids in its seed oil. LS oil has alpha linoleic acid which could give it nutritional advantages (Gunstone, 2004 and Diwakara et al., 2008). The primary fatty acids in LS oil were oleic (30.6 wt %) and linolenic acids (29.3 wt %). LS seeds oil contained high concentrations of tocopherols. The primary phytosterols in L S were sitosterol and campesterol, with avenasterol (Bryan et al., 2009). The results of lipid profile were agreed with results obtained by Das et al., (1997) who reported that the lipid profile of hypercholesterolemic animals were significantly higher than control rats for total lipid, total cholesterol , triglyceride, LDL-c and VLDL-c while only HDL-c was significantly lower in hypercholesterolemic rats than in control rats.

The increased plasma cholesterol, particularly LDL-c is one of the most important risk factor for coronary vascular disease. LDL-c particle are taken up by macrophage cells after oxidized or modified and then deposited in the arterial intima leading to formation of atheroma (Durrington, 1995). Low HDL-c levels are considered as a strong risk factor for coronary heart disease as HDL-c act as antioxidant and protect LDL-c from oxidation so that reduce LDL-c from circulation (Boden and Pearson, 2000 and Glass and Witztum, 2001). Lepidium sativum L. seeds have revealed presence of glycoside, alkaloids, tannin (Phenolic compound), flavonoids, and amino acids like glutamine, cysteine, and glycine. The tannin and flavonoids may have antioxidant activity whenever glutamate, cysteine, glycine are intermediates for synthesis of the endogenous antioxidant glutathione. Diets rich in alpha linolenic acid have been associated with a reduced risk of fatal ischemic heart disease, a reduction in heart attacks and mortality from chronic vascular disease. Feeding alpha linolenic acid has also been shown to decrease platelet aggregation, total cholesterol, LDL cholesterol and triglycerides in humans and rats (Olsson, and Yuan 1996, Kirtkar and Basu 2005 and Hamer and Steptoe, 2006).

Protective and curative treatment of ethanolic extract of Lepidium sativum seeds L. in renal failure of rats significantly reduced the level of urea and creatinine that indicates increase glomerular filtration rate. Extract of Lepidium sativum L. seeds may be having nephroprotective and curative activity. Daily oral administration of aqueous LS extract for 3 weeks exhibited antihypertensive and diuretic activities. ALT and AST are closely correlated in most cases of liver diseases. Excessive storage of fat in the liver effects on liver functions and increases the susceptibility to free radical attack in hyporcholesterolemic rats resulting in liver damage as described by Tahri et al .,(2000), Mhamed et al .,(2005) and Yadav et al .,(2009). Feeding rats with 10% Garden cress seed oil lowered hepatic cholesterol by 12.3% and serum triglycerides by 40.4% compared to SFO fed group. Very low density lipoprotein cholesterol (VLDL-C) and low density lipoprotein cholesterol (LDL-C) levels decreased by 9.45% in serum of 10% LS oil fed rats, while HDL remained unchanged among LS oil fed rats (Diwakar et al .,2008)

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6. References:

- 1. Abd El-Ghanny, M. A, Magda, K. El-Shaer and Maaly, Y. Gaber (2007): Therapeutic Effect of Some Fat Soluble Vitamins ON Hyperlipidemic Rats. J of Home Economics, Minufia Univ., 17(1):31-42.
- Allain, C.C., Richmond, N and Rosechloy, P. (1974): Cholesterol enzymatic colorimetric test. Clin. Chem., clin, 19 (20): 1350-470.

- Abell, L.L., Levy, B.B., Brodie, B.B and Kendal, R. (1952): A simplified method for the estimation of total cholesterol in serum and demonstration of its specificity. J .Biol .Chem: 357-366.
- Aburjai, T., Darwish, R.M., Al-Khalil, S., Mahafzah, A., Al-Abbadi, A., (2001): Screening of antibiotic resistant inhibitors from local plant materials against different starins of *Pseudomonas aeruginosa*. Journal of Ethnopharmacology 76, 39–44.
- 5. Artimage, G.Y and Berry, W.G (1987): Statistical Methods. 7th Ed. Ames, Iowa Stata University Press, 39-63.
- 6. Bartholomev, R.J. and Delany, A. (1966): Proc Aust. Assoc. Biochemists. 1, 214.
- Boden, W.E. and Pearson, T.A. (2000): Raising low of high-density lipoprotein cholesterol is an important target of therapy. Am J Cardiol. March 1, 85(5): 645-650.
- 8. Bonsens, K. E. and Taussky, D. H. (1984): Determination of serum creatinine .J Chem Inv, 27: 648-660.
- Bryan, R. M., Shailesh, N. S., Jill K. W., Steven F. V and Roque, L. E(2009): Composition and physical properties of cress (Lepidium sativum L.) and field pennycress (Thlaspi arvense L.) oils. Industrial Crops and Products 30, 199–205.
- 10. Buccolo, G. and David, H. (1973): Ouantitative determinarion of serum triglycerides by use enzymes. Clin. Chem., 19: 419-32.
- 11. Castelli T and levitar Y (1977): Atherogenic index, Curr presc P. 39.
- Chapman, D.G.; Gastilla, R. and Campbell, T.A. (1950): Evaluation of protein in food. I. A. Method for the determination of protein efficiency ratio. Can. J. Biochem. Physio. I (37) 679-686.
- 13. Coles, E. H. (1974): Veterinary Clinical Pathology. Saunders Company, Philadelphia and London.
- Das, S.; Snehlata, M.P. and Srivastava, L. M. (1997): Effect of ascorbic acid on lipid profile and lipid peroxidation in hypercholesterolemic rabbits. Nutr. Res. 17 (2): 231-241.
- 15. Diwakara, B.T., Duttaa, P.K., Lokeshb, B.R. and Naidu, K.A. (2008): Bioavailability and metabolism of n-3 fatty acid rich garden cress (Lepidium sativum) seed oil in albino rats. Prostaglandins, Leukotrienes and Essential Fatty Acids. 78, 123– 130.
- Durrington, P.N. (1995): Hyperlipidaemia, Diagnosis and Management.2nd Edition, P. 25-74. Butterworth-Heinemann, Cambridge, Britain.

- 17. Eddouks, M., Maghrani, M., Zeggwagh, N.A and Michel, J. B (2005): Study of the hypoglycaemic activity of Lepidium sativum L. aqueous extract in normal and diabetic rats. J Ethnopharmacol. , Feb 28; 97 (2):391-5.
- Folch, J., lees, M., and Stanley, G.H. (1957): A simple method for isolation and purification of total lipid from animal tissue. J Biol Chem 266: 497-509.
- 19. Fruchart, G.G. (1982): LDL-Cholesterol determination after separation of low density lipoprotein. Rev. Fr. Des. Laboratories, 103: 7:117.
- 20. Glass, C.K. and Witztum, J. L. (2001): Atherosclarosis: The Road Ahead Cell. 104(4): 503-516.
- 21. Gokavi, S.S., Malleshi, N.G.and Guo, M(2004): Chemical composition of garden cress (Lepidium sativum) seeds and its fractions and use of bran as a functional ingredient.Plant Food. Hum. Nutr. 59, 105–111.
- 22. Gunstone, F.D (2004): The Chemistry of Oils and Fats. Sources, Composition, Properties and Uses. CRC Press, Boca Raton.
- 23. Hamer, M. and Steptoe, A. (2006): Influence of specific nutrients on progression of atherosclerosis, vascular function, haemostasis and inflammation in coronary heart disease patients: a systematic review. Br. J. Nutr. 95, 849–859.
- Jouad, H., Haloui, M., Rhiouani, H., El Hilaly, J and Eddouks, M. (2001): Ethnobotanical survey of medicinal plants used for the treatment of diabetes, cardiac and renal diseases in the North centre region of Morocco (Fez-Boulemane). J. Ethnopharmacol, 77: 175 – 182.
- 25. Kassie, F., Laky, B., Gminski, R., Mersch-Sundermann, V., Scharf, G., Lhoste, E. and Kansmuller, S. (2003): Effects of garden and water cress juices and their constituents, benzyl and phenethyl isothiocyanates, towards benzo(a)pyreneinduced DNA damage: a model study with the single cell gel electrophoresis/Hep G2 assay. Chemical Biology Interactions 3, 285–296
- 26. Kirtkar, K. M. and Basu, B.D. (2005): Indian medicinal Plants. I: 174.
- Kostener, C. M. (1977): Enzymatic determination of cholesterol high density lipoprotein fraction prepared by polyanion precipitation. J. Clin. Chem., 22:695.
- Mhamed, M, Naoufel-A.Z., Jean-Baptiste M.and Mohamed, E (2005): Antihypertensive effect of *Lepidium sativum* L. in spontaneously hypertensive rats. Journal of Ethnopharmacology ,100 , 193–197

- 29. NRC (1995): National Research council: Nutrient requirements of laboratory animals. Fourth revised edition, PP.29-30. National Academy Press. Washington, DC.
- Olsson, A.G. and Yuan, X.M. (1996): Antioxidants in the Prevention of Altherosclerosis, Curr. Opin Lipidol., 7(6): 374-380.
- 31. Paget, G.E. and Barnes, J.M. (1964): Inter species dosages conversion scheme in evaluation of results and quantitative application in different species toxicity test. 135-165. Academic Press London and NY.
- Patton, C.J. and Crouch, S.R. (1977): Enzymatic colorimetric method to determination urea in serum. Anal. Chem., 49: 464.
- Reitman, S. and Frankel, S. (1957): Determination of glutamate pyruvat transaminase and glutamate oxaloacetate transaminase. Amer. J. Clin. Path., 28:56-63.
- 34. Scheletter, G and Nussel, E. (1975): Arbeitsmed Sozialmed Praventimed, 10: 25.
- 35. Tahri, A., Yamani, S., Legssyer, A., Aziz, M., Mekhfi, H., Bnouham, M., and Ziyyat, A., (2000): Acute diuretic, natriuretic and hypotensive effects of a continuous perfusion of aqueous extract of Urtica dioica in the rat. Journal of Ethnopharmacology 73, 95–100.
- 36. Tahraoui, A., El Hilaly, J., Israili, Z.H., Lyoussi, B., (2007): Ethnopharmacological survey of plants used in the traditional treatment of hypertension and diabetes in south-eastern Morocco (Errachidia province). Journal of Ethnopharmacology 110, 105– 117.
- Weichselbaum , T.F (1946): An accurate and rapid method for the determination of protein in small amount of blood serum and plasma .Am .J. Clin .Path. (16):40.
- 38. Yadav, Y.C., Srivastav, D.N., Seth, A.K., Gupta, V.D and Kuldeep, S.(2009) : Nephroprotective and curative activity of Lepidium Sativum L. seeds in albino rats using cisplatin induced nephrotoxicity. Pharmacology on line 3: 640-646.
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