

Production and application of *Spirulina platensis* rich in fatty acids, and vitamins

Aly, M.S.¹, Amber.S. Gad^{*2}, El Sayed M. Kamel

¹Agriculture Microbiology Dept., ²Chemistry of Natural and Microbial Products Dept., National Research Center, Egypt. Faculty of medicine, Pharmacology. Dept. Zagazig Univ

* amber2gad@yahoo.com

Abstract: *Spirulina platensis* is a microscopic blue-green [alga](#) in the shape of a spiral coil, living both in sea and fresh water. It is widely used as health food due to its protein content, vitamins and active substances for immune system. Polyunsaturated fatty acids amount to 46.548 % (w/w) of total lipids. Among the essential fatty acids detected in El Khadra lake water body in Waadi El Natroun micro-alga, cholesterol decreasing -linolenic acid with 0.986% (w/w). Vitamin A amounts to 120.13 µg/100g, vitamin C amounts to 540.34 µg/100g and vitamin D amounts to 105.6 µg/100g were found. Vivo studies revealed *Spirulina* effectiveness on Triglycerides (TG), Total cholesterol (TC), High density lipoprotein-cholesterol (HDL-ch), body weight, serum calcium, serum iron, and serum ferritin after treatment of the experimental rabbits for 30 days.

[Aly, M.S., Amber.S. Gad. El Sayed M. Kamel. **Production and application of *Spirulina platensis* rich in fatty acids, and vitamins.** Journal of American Science 2011;7(2):36-45]. (ISSN: 1545-1003). <http://www.americanscience.org>.

Keywords: *Spirulina platensis*, -linolenic acid, vitamins, hypercholesterolemia, serum calcium, prothrombin, serum iron, serum ferritin.

1. Introduction:

Spirulina is a natural health food as a blue-green algae. It contains beneficial nutrients that are readily digested and absorbed by the body, so none of its nutritional benefits are lost. It is excellent in combating imbalances arising from lifestyle habits and it is effective in overcoming and preventing various disorders arising from a poorly balanced diet, including insufficient intake of vegetables as it supplies several of the vitamins that all living beings need to carry on metabolic processes or prevent some serious diseases; Cingi *et al.* (2008): This cyanobacterium is important for its content of polyunsaturated fatty acids as it is frequently rich in [gamma-linolenic acid](#) (GLA), and also provides [alpha-linolenic acid](#) (ALA), [linoleic acid](#) (LA), [stearidonic acid](#) (SDA), [eicosapentaenoic acid](#) (EPA), [docosahexaenoic acid](#) (DHA), and [arachidonic acid](#) (AA); Babadzhanov *et al.* (2004), beta-carotene and other antioxidants pigments; Ciferri (1983); Gemma *et al.* (2002); anti-virals sulphated polysaccharides; Noaman *et al.* (2004), antimicrobials sterols. *Spirulina* contains a number of vitamins: B1, B2, B3, B6, B9, vitamin C, vitamin D, vitamin E; Babadzhanov, *et al.* (2004) and B12, Watanabe *et al.* (2002).

Spirulina is accepted as functional foods, which are defined as products derived from natural sources,

whose consumption is likely to benefit human health and enhance positive medical properties. These foods are used as a supplement/ingredient or as a complete food to enhance the performance and state of the human body, or improve a specific bodily function. It is being widely studied for its possible antioxidant, antibacterial, and antiparasitic properties, and for several medical conditions such as allergies; Mao *et al.* (2005) ulcers, anemia, heavy-metal poisoning, Barmejo-Bescós *et al.* (2008), and radiation poisoning, Misbahuddin *et al.* (2006); Raj *et al.* (2008). *Spirulina* or its extracts can prevent or inhibit cancer in humans and animals and has other medical effects; Wang *et al.* (2005)

Functional foods are used mainly as products to nourish the human body after physical exertion or as a preventive measure against ailments. *Spirulina* contains unusually high levels of gamma-linolenic acid (GLA), an essential polyunsaturated fatty acid; Otle and Pire (2001); Babadzhanov, *et al.* (2004). The aim of this study is to evaluate fatty acids and vitamins contents of *S. platensis* isolated from Waadi El Natroun and studying its effect on cholesterol TG, TC, HDL levels, body weight, and serum calcium, serum iron and serum ferritin in the blood of experimental animals.

2. Materials and Methods:

Micro-organisms:

Spirulina platensis used in this study were obtained from El-Khadra lake at Wadi El Natroun, Egypt, characterized by extreme conditions of pH 10.5 and salt concentration of 0.55 M, (Aly, 2000).

Maintenance stock media:

Zarrouk's synthetic medium was used for maintaining and batch culture preparation of *S.platensis*(SP) as described according to Ali and Amber (2010).

Harvesting the biomass:

After seven days incubation, the algal biomass were harvested with the 40 µm mesh size cloth, filtered and washed with distilled water to remove the salts from the algal surface then, the washed slurry was divided into two sets, first for determination of vitamins and fatty acids which stored at -20 °C while the second set was dried at 50 °C for 1 h and milled; Ali and Amber (2010). The dried samples were milled and stored in plastic container to be used in feeding experimental animals.

Extraction and Identification of vitamins; Qian and Sheng (1998): Ten grams of SP tissue fresh weight were homogenized with methanol for extraction of water soluble vitamins while acetone–chloroform (30:70 v/v) was used for extraction of fat soluble vitamins. The mixtures were shaken on a vortex mixer for 5 min, centrifuged at 4000 rpm for 5 min and filtered through a Millipore filter (45 µm). The filtrates were evaporated under nitrogen and the residues were re-dissolved in 1 ml water for water soluble vitamins and in 1 ml butanol for fat soluble vitamins were quantified by HPLC. Vitamin C was analyzed by AOAC(1995), -tocopherol (vitamin E) by HPLC; Manz and Vuilleumier(1988), - carotene by spectrophotometric method, AOAC(1995).

Lipids were obtained from lyophilized biomass sample according to Folch *et. al.* (1957) lipids were extracted with chloroform/ methanol (2+1 v/v) purified in methanol/ water (2:1 v/v), containing 9 g Na Cl to remove sugar, salts, protein and concentrated in a rotary evaporator residual solvents were evaporated. Lipids were gravimetrically estimated.

Extraction and Identification of fatty acids (FA): (Isik *et. al.* 2006; Diarman *et. al.* 2009).

FA were analysed by IUPAC (1982) method with Thermoquest Trace GC. FID detector (250 °C) and SP-2330 Fused silica capillary column 30 m.-0.25 mm ID-0.20 µm (film thickness) of cyanopropyl were also used. Air was adjusted 350 ml/min. 35 ml/min H₂ and 30 ml/min He were used. The range, carrier ratio, split flow and split ratio were 1, 0.5 ml/min, 75 ml/min and 1/150, respectively. Oven temperature was 120 °C (up to 220 °C with the adding of 5°C. The sample injection was 0.5 µL. The FA was identified by comparing them in their retention time with standards obtained from Sigma.

Vivo studies.

Healthy adult (1-1.5 kg) white New Zealand rabbits were kindly provided by NRC, Egypt, housed and maintained under a constant temperature of 30±1°C. i.e. animals were acclimatized to laboratory conditions before the experiment with a week. Rabbits were given food and water *ad libitum* along the period of the treatment. Animals were randomly divided into two groups (n=10 per group) and treated for a period of a month as follows: 1) Group 1 (SP-treated group): animals were fed on a standard diet as 100 mg *Spirulina* /kg weight (100mg powder dissolved in 10 ml sterilized water) by a gavage daily for one month and given water for 30 days; 2) Group 2 (untreated control group): animals were fed on a standard diet until the termination of the experiment; Body weight of the animal was recorded every 10 days.

Biochemical analysis

After 15 days fasting following the end of the experimental time, the animals were cut in ears, blood samples were collected from the marginal vein of the ear every 10 days for one month in clean dry test tube. The samples were kept for 30 min at room temp to clot then centrifuged at 3000 rpm for 10 min. Clear serum was divided into aliquots and was used for the biochemical determinations.

Aliquot of serum samples were kept at -20°C until used for determinations.

Total cholesterol (TC) was evaluated according to Richmond (1973).

Triglycerides (TG) were evaluated according to Trinder (1969).

High density lipoprotein cholesterol (HDL-CH) was evaluated according to Lopez-Virella *et. al.* (1977).

Calcium serum was evaluated according to Gindler and King (1977).

Ferritin serum was evaluated according to White *et al.* (1986).

Iron serum. was evaluated according to Dreux (1977).

Atherogenic index (AI)=TC-HDL-ch/HDL-ch.AI when increased than 2 lead to atherosclerosis.

Statistical analysis:

Data are expressed in mean \pm SE of three replicates, results were considered significant when $p < 0.01$.

3. Results and Discussion

Fatty acids content in the local strain of *S.platensis*:

A concentrated source of nutrients such as *Spirulina* is loaded with fats, starches and calories. *Spirulina* contains 7 % lipid, and most of that is in the form of essential fatty acids that promote cholesterol normalization. The essential fatty acids sometimes called vitamin F, include linoleic, linolenic and arachidonic acid. They are used by the body to manufacture prostaglandins, the hormonal regulators of blood pressure and capillary resilience ;Tomaselli *et al.* (1988) .Otle and Pire (2001) determined the composition of *Spirulina platensis* fatty acid composition. The essential fatty acids are involved in respiration in all the cells, and are especially important to oxygen transport. They affect the health of the hair, skin and nails, and help break up cholesterol in the blood stream. Isik *et al.*(2006) found that chemical composition of *Spirulina* related to the environmental cultivation conditions, salinity, light intensity, pH which determines the solubility of CO₂ and minerals in the medium. Data presented in table 1 indicated that PA amounted to 33.452%(w/w). Quoc and Dubacq (1997) ; Romano *et al.* (2000); Mühling *et al.* (2000) and Tomaselli *et al.* (1988) studied the temperature influence on *S. platensis* M2 and determined the fatty acids contents of *S. platensis* at the different temperatures, They observed that the lessening temperature led to the decrease of the C16:0 content. Similarly, Tomaselli *et al.* (1988) and Romano *et al.* (2000)reported the increase in C16:1 level by the low temperatures 26 °C. On the contrary, Quoc and Dubacq (1997) reported the increase in the level of C18:2 with the lowering of the temperature. It was shown that

cyanobacteria responded to a decrease in ambient growth temperature by de-saturating the fatty acids of membrane lipids to compensate for the decrease in membrane fluidity at low temperatures ;Tomaselli *et al.* (1988). In addition, the proportion of de-saturated fatty acids increases by the decrease in temperature . Oliveira *et al.* (1999) reported the increase of the C18:3 with lowering temperature .The percentage of C18:2 decreased at the 26 °C. In agreement with these studies, table 1 indicated the level of C16:0 was found to be 33.452%(w/w), palmitoleic fatty acid (16:1) 1.262, (w/w)% of total lipids which is used to fight weight gain., while C18:2.yield was 5.523% (w/w). Table 1, showed also,the content of C18:3: was small 0.986% (w/w)..;oleic acid (C18:1) yields 37.988%(w/w).; linoleic acid (C18:2) represents 5.523 % (w/w)., LA content was similar to a previous study 10-37% (w/w).;Diraman *et al.*(2009).

GLA (-linolenic) acid (C18:3) that is associated with pharmaceuticals and nutraceuticals amount to 0.986%(w/w) of lipids .However, *S. platensis* is a very rich source in -linolenic acid. It has been found that contents and composition of fatty acids are temperature dependent in *S.platensis*; an increase in temp reduce the composition of fatty acids in membrane lipids, Colla *et al.* (2008). GLA yield depends on dark and light cycles ,indoor or outdoor cultivation , harvest time,age of the culture, This low yield in fatty acids in Wadi El Natroun *S.platensis* may be due to the fact stated by ;Diraman *et al.* (2009) who confirmed that, the mechanism of fatty acids composition are not fully understood ,variation in percentage concentrations may be due to temp.or sodium nitrate concentration in the synthetic media used .Table 1 showed that PA and OA were the most abundant.

Caprylic acid (C8:0) yield is 1.578 (w/w) % known with its strong anti-fungal properties, Capric acid (C10:0) yield is 6.154% w/w)used in making artificial fruit flavors. Lauric acid (C12:0) helps in curing skin infections and dandruff.; myristic acid (C14:0) represents 2.761%, palmitic acid (C16:0) yield is 33.4(w/w) %.; stearic acid (C18:0)represents 5.08 (w/w) % , and arcachedic acid (C20:0) yield is 2.761%. -linolenic acid (GLA and),linoleic fatty acid (LA) are poly and mono- unsaturated fatty acids. Table2 indicated the amount of -linolenic acid was only 0.986% (w/w)). Total saturated fatty acids amount to 53.452.% (w/w). GLA in particular has a role in lowering the decrease blood cholesterol level; Ishikwa *et al.* (1989) used it in treatment of hypercholesterolemia.

Table (1). (%) Fatty acid in *S. platensis* biomass.

Fatty acid	Concentration%(w/w)	Fatty acid	Concentration% (w/w)
Short chain (CA)C8:0	1.578	Unkown	0.789
(CA) C10:0	6.154	(SA)C18:0	5.089
Long chain (IA)C12:0	1.657	(OA)C18:1	37.988
(MA)C14:0	2.761	(LA)C18:2	5.523
(PA)C16:0	33.452	(GLA)C18:3	0.986
(POL)C16:1	1.262	Very long chain (AA) (C20:0)	2.761

Table 2. Concentration % of γ -linolenic poly unsaturated fatty acid in *S.platensis* biomass

GLA% (w/w)	UFA% w/w)	L A% (w/w)	OA% (w/w)	SF% w/w)
0.986	46.548	5.523	37.9	53.452

Total fatty acids TFA, linoleic fatty acid (LA), oleic fatty acid (OA) γ -linolenic fatty acids (GLA) - Unsaturated fatty acids (USf).

Vitamins content in the local strain of *S. platensis* :

S. platensis biomass could be obtained under the optimum laboratory cultivation conditions for the quantification of β -carotene, vitamin A and α -tocopherol. *Spirulina* contains a number of vitamins: B1 (thiamine), B6 (pyridoxine), B9 (folic acid), vitamin C, vitamin D, vitamin E and B12 (cobalamin), biotin, pantothenic acid, beta carotene (source of vitamin A, Elizabeth and Lillian (1968); Watanabe *et al.* (2002); García-Martínez *et al.* (2007), *Spirulina* is rich in vitamin B12. Vitamin B12 (cobalamin), ascorbic acid, (vitamin C), tocopherols (vitamin E), phyloquinone (vitamin K1) and menaquinones (vitamin K2), vitamins: A (β -carotene); B1 (thiamine), B2 (riboflavin), B3 (niacin), were also detected in *Spirulina* extract in a satisfactory amounts; García-Martínez *et al.* (2007).

***S.platensis* and vitamin A:** Table (3) indicated that *Spirulina* contains 120.13 μg / 100g wet weight. Vitamin A helps form and maintains healthy teeth, skeletal and soft tissue, mucous membranes, and skin. It is also known as retinol because it produces the pigments in the [retina](#) of the eye; Duester (2008).

***S.platensis* and vitamin C (ascorbic acid) :** Table (3) indicated that *Spirulina* contains 540.34 μg / 100g wet weight. Ascorbic acid is required for healing of wounds, the production of digestive enzymes and connective tissue, brain and nerve function, formation of teeth and bones, glandular activity. Also vitamin C aids in the absorption of iron and protection of cells, B complex vitamins, vitamin E and vitamin A from oxidation. *Spirulina* helps in curing all immune system problems as well; Quereshi *et al.* (1994).

***S.platensis* and vitamin E:** Table (3) indicated that *Spirulina* contains Tocopherol or vitamin E (105.6µg/100g wet weight). This nutrient protects heart and vascular health, promotes oxygenation of cells; Zingg ,and Azzi(2004), and retards aging. Vitamin E deficiency in humans results in ataxia (poor muscle coordination with shaky movements), decreased sensation to vibration, lack of reflexes, and paralysis of eye muscles. One particularly severe symptom of vitamin E deficiency is the inability to walk.

Table (3). Vitamins in *S. platensis* biomass.

Vitamin	Concentration (µg/100g wet biomass)
(C)Ascorbic	540.34
-carotein(A)	120.13
- tocopherol (E)	105.6

In vivo studies

Effect of *S.platensis* on experimental animals body weight:

Table 4 shows the change recorded in rabbits that had been fed *Spirulina* diet for 4 weeks ,as the body weight of each animal along the experimental period (one month) is given. Kumar *et. al.* (2010) stated that *S.platensis* has diverse biological effect due to high content of highly digestible protein, vitamins, beta-carotene ,phycocyanin and other pigment. Table 4 shows increasing percentage of change in the animals weight with time in all the groups under treatment as compared to control groups, this comes in agreement with Yin, *et.al.* (2008). As the body weight of each animal was recorded every 10 days, the recorded increase in the 3rd period in the experimental animals is 2.10 ± 0.30 (Kg) refers to the "free-feeding" weight of an animal, "The rabbit's *ad libitum* weight was about 300 grams.

Table (4). Mean weight of rabbits over the test period \pm SD

Weight elevation (%)	Test (Kg)	Control (Kg)	Period(10 days)
7.462	1.40±0.11	1.34±0.05	1
6.211	±0.2011.66	1.61±0.10	2
5.376	2.10±0.30	1.86±0.12	3

Effect of *S.platensis* on the cholesterol titres:

Early interest in *S.platensis* focused mainly on its potential as a source of protein and vitamins but recently more attention has been made to study its therapeutic use and number of reports suggested its beneficial effect in acute allergic rhinitis, anti-cardiotoxic ,anti-hepatotoxic, anti-nephrotoxic; Kumar *et. al.* (2010). Table 5 indicated that after 30 days feeding with *Spirulina* , TG reached 48.23 ± 2.11 mg % compared to the control, TC 45.61 ± 3.21 mg % reached a decrease to 18.27 ± 0.28 mg % compared to the control 25.24 ± 0.42 mg % , HDL-Ch recorded 17.91 ± 1.50 mg % increase as compared to the control 12.23 ± 1.15 mg % under the same laboratory conditions, as total blood cholesterol below 200 mg/dL indicating

the relatively low risk of coronary heart disease , even with a low risk this comes in agreement with , Edlin *et. al.* (2009).

Serum levels of total glycerides and free glycerol are important indices of lipid metabolism and cardiovascular disease risk. Khan *et. al.*(2005). Triglycerides(TG) are a type of fat in the bloodstream and fat tissue. Too much of this type of fat, as table 5 indicated this value reached 48.23 ± 2.11 mg % can contribute to the hardening and narrowing of arteries. This puts risk of having a heart attack or stroke; John *et. al.*(2008) .Diseases such as diabetes, obesity, kidney failure or alcoholism can cause high triglycerides. Often, high triglycerides occur along with high levels of [cholesterol](#), which is an important component for the

[manufacture](#) of fat-soluble vitamins including [vitamin A](#), [vitamin D](#), [vitamin E](#), and [vitamin K](#). HDL particles are able to remove cholesterol from [atheroma](#) within [arteries](#) and transport it back to the liver for excretion or re-utilization, which is the main reason why the cholesterol carried within HDL particles, termed HDL-C, is sometimes called "good cholesterol". higher levels of HDL-C recorded 17.91 ± 1.50 mg % as data presented in table 5 seem to indicate fewer problems with [cardiovascular diseases](#), while low HDL-cholesterol levels (less than 40 mg/dl or about 1 mmol/L) have indicated rates for heart disease .

There are several types of cholesterol, each made up of lipoproteins and fats. Each type of lipoprotein contains a mixture of cholesterol, protein and a type of fat (triglyceride), but in varying amounts. Nayaka *et al.*(1988). Of the lipoprotein types, VLDL contains the highest amount of triglyceride. Because it contains a high level of triglyceride, having a high VLDL level means the increased risk of coronary artery disease, which can lead to a heart attack or stroke. A normal VLDL cholesterol level is between 5 and 40 milligrams per deciliter. Since higher levels of LDL particles promote health problems and [cardiovascular disease](#), they are often called the bad cholesterol particles, (as

opposed to [HDL](#) particles, which are frequently referred to as good cholesterol or healthy cholesterol particles). VLDL-cholesterol is a minor lipid component of very low-density lipoprotein (VLDL) particles of VLDL particle; Ren *et al.* (2010). A study involving geriatric patients determined that *Spirulina* helped to significantly reduce the [LDL-to-HDL](#) ratio after four months of supplementation; Park *et al.* (2008). Treatment with over a six week period, exhibited significant changes in [cholesterol](#) and [blood pressure](#) as it lowered total cholesterol, increase [HDL cholesterol](#), lower [triglycerides](#); and lower [systolic](#) ; Torres-Duran *et al.* (2007).

Table 5 showed that AI 0.71 ± 0.13 for the control and 0.27 ± 0.11 for the treatment .i.e. not the least evidence for atherosclerosis. Arteriosclerosis is hardening of the arteries. This condition not only thickens the wall of arteries, but also causes stiffness and a loss of elasticity. Over time, the arteries become harder and harder as they are slowly damaged by high blood pressure. Atherosclerosis is the most common type of arteriosclerosis, or hardening of the arteries, and caused by plaque building up in the vessel. Over time the plaque causes thickening of the walls of the artery, when AI increased than 2 lead to atherosclerosis.

Table (5).Effect of *S. platensis* tablets(100mg/Kg) in lipid profile of the rabbits \pm SE.

Period	group	TGmg%	TC mg%	HDL-ch mg%	AI
1	control	48.93 \pm 5.84	20.63 \pm 3.06	14.43 \pm 2.45	0.726 \pm 0.124
	test	58.8 \pm 3.60	26.5 \pm 2.82	11.22 \pm 1.42	1.74 \pm 0.266**
2	control	46.8 \pm 5.96	14.93 \pm 1.15	9.22 \pm 0.99	0.93 \pm 0.223
	test	49.58 \pm 3.83	19.48 \pm 0.43	18.08 \pm 1.74**	0.37 \pm 0.23*
3	control	45.61 \pm 3.21	25.24 \pm 0.42	12.23 \pm 1.15	0.71 \pm 0.13
	test	48.23 \pm 2.11	18.27 \pm 0.28	17.91 \pm 1.50	0.27 \pm 0.11*

*p<0.1 corresponding to control

**p<0/001 corresponding to control.

**p<0/001 corresponding to control

Effect of *S.platensis* on serum calcium level:

Serum calcium is a test that measures how much calcium is in blood. The presence of free calcium was a necessary condition of co- agulation as certain salts of citrate and oxalates electrolytes have low solubility product, so it can be used as anti-co- agulation. However, in combination with prothrombin, calcium acts not as free element but as a complex that could interfere in blood clotting; Levelock and Porterfield (1952). Fig .1 shows the increase in the serum calcium level of the treated than the control sample in the three periods, which is prominent in the 2nd period, as it amounts to 11.3mg/dl compared to 5.43mg/dl of the control test. Also, the parameter is still high in the 3rdperiod as it measures 9.6 mg/dl compared to 7.1 mg/dl of the control. This decrease may be due to the use of this calcium by the body. the body uses vitamin D to help transport calcium to the bones When blood calcium levels drop too low, the vital mineral is "borrowed" from the bones. It is returned to the bones from calcium supplied through the diet. . Calcium is the most abundant mineral in the body; the bones and teeth accounting for about 99% of the total body stores.

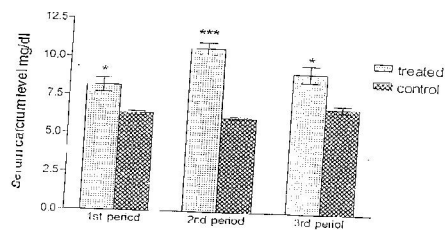


Figure (1) Serum calcium level of control and treated rabbits with daily 100mg/kg tablet of *Spirulina platensis* for one month.
 * significant at $p < 0.05$
 ** highly significant
 *** extremely significant

Fig. (1) *S. platensis* effect on serum calcium.**Effect of *S.platensis* on serum iron and serum ferritin:**

Free iron is [toxic](#) to [cells](#) as it acts as a catalyst in the formation of free radicals from [reactive oxygen species](#) via the [Fenton Reaction](#). [Orino et. al.\(2001\)](#). Hence body uses an elaborate set of protective mechanisms to bind iron in various [tissue](#) compartments. Within cells, iron is stored in a protein complex as ferritin. The amount of ferritin in blood (serum ferritin level) is directly related to the amount of iron stored in the body, i.e. under [steady state](#)

conditions, the [serum](#) ferritin level correlates with total body iron stores; thus, the serum ferritin is the most convenient laboratory test to estimate iron stores as it serves to store iron in a non-toxic form, to deposit it in a safe form, and to transport it to areas where it is required, [Seckback \(1982\)](#). The serum ferritin concentration is a clinical parameter measured widely for the differential diagnosis of anemia. Ferritin is a ubiquitous intracellular protein that stores iron and releases it in a controlled fashion. In humans, it acts as a buffer against iron deficiency and iron overload [Seckback \(1982\)](#). The function and structure of the expressed ferritin protein varies in different cell types. Fig (2,3) illustrates the effect of 100 mg *Spirulina*/ kg daily administration to the rabbits . Control and treated samples showed a significant decrease in serum ferritin in the third period due to the use of its iron in hemoglobin formation for example as high ferritin is correlated to iron in excess. Serum iron increase significantly in the 3rd period as it reached 5.5mg/dl compared to 2mg/dl of the control. Ferritin decreased in the 3rd period 12 mg /dl because of iron consumption in hemoglobin but it still in a higher concentration than control. 9.4 mg/dl.

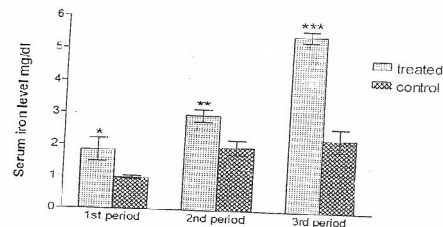


Figure (2) Serum iron level of control and treated rabbits with daily 100mg/kg tablet of *Spirulina platensis* for one month.

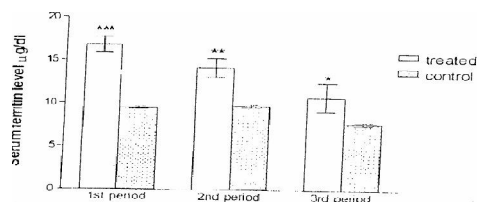
Fig. (2) *S. platensis* effect on serum iron.

Figure (3) Serum ferritin level of control and treated rabbits with daily 100mg/kg tablet of *Spirulina platensis* for one month.
 * significant $p < 0.05$
 ** highly significant $p < 0.01$
 *** extremely significant $p < 0.001$.

Fig. (3) *S. platensis* effect on serum ferritin.

Corresponding Author*

Amber .S. Gad

Chemistry of Natural and Microbial products
Dept.,NRC.,Dokki, Egypt.amber2gad@yahoo.com**References:**

- Aly, M.S. (2000). Ecological studies on phytoplankton in closed lakes at Wadi El Natroun. Egypt.J.Phyc.,1:2000.
- Ali,M.S.,and Amber S. Gad.(2010). Chemical composition and potential application of *S. platensis*. J. Am. Sci.6(10): 1283-1291.
- A.O.A.C.(1995):Association of official analytical chemists 15th ed. ,Washington ,DC.
- Babadzhanov,A.S.;Abdusamatova.N.; Yusupova,FM; Fayzullavea,N; Mezhlumyan, LG.,and Mailkova,MK. (2004). Chemical Composition of *S. platensis* Cultivated in Uzbekistan. Chem.of Nat.Comp.. 40, 3:276-279.
- Barmejo-Bescós, P.; Piñero-Estrada, E.;and Villar del Fresno, A. (2008). Neuroprotection by *S. platensis* protean extract and phycocyanin against iron-induced toxicity in SH-SY5Y neuroblastoma cells. Toxi. in Vitro 22 (6): 1496–1502.
- Ciferri, O. (1983) .*Spirulina*, the Edible Microorganism. Microbiol. Rev. 47, PMID: PMC283708.
- Cingi, C.; Conk-Dalay, M.; Cakli, H., and Bal, C. (2008). The effects of *Spirulina* on allergic rhinitis. Eur. Arch.of Oto-Rhino-Laryngol. 265 (10): 1219–1223.
- Colla, L.M.; Muccillo-Baisch, A.M. ; and Costa, J.A.V. (2008). *S. platensis* effects on the levels of total cholesterol, HDL and triacylglycerols in rabbits fed with a hypercholesterolemic diet. Braz. Arch. Biol. Technol.51 (2) .405-411.
- Diraman,H.;Koru,E. and Dibeklioglu,H. (2009). Fatty acid profile of *S. platensis*. Isr. J.of aqua.-Bamidgeh,61(2):134-142.
- Dreux,C.(1977).Ann. Biol.Clin.35:275.
- Duester, G. (2008). Retinoic acid synthesis and signaling during early organogenesis. Cell 134: 921-931.
- Edlin,G .,and Golanty, E. (2009).Understanding and preventing diseases. P: 332.Health and Fitness.Part IV-804.pages.
- Elizabeth A. Cook, and Lillian N. Ellis (1968).Variations in the Growth Response of Four Different Vitamin B₁₂ assay Microorganisms to the Same Tissue and Standard Preparations.. Appl Microbiol.:16(12): 1831–1840.
- Folsh, J.; lees, M., and Sloane- Stanley, G.H. (1975). A simple method for isolation and purification of total lipids from animals tissue.J.Biol.Chem.226-497.
- García-Martínez, D.; Rupérez ,FJ.; Ugarte, P. *et. al.* (2007). Tocopherol fate in plasma and liver of streptozotocin-treated rats that orally received antioxidants and *Spirulina* extracts. Int J Vitam. Nutr. Res.;77:263-71.
- Gemma, C.; Mesches M. H.; Sepesi B. ; Choo K.; Holmes D. B. , and Bickford ,P. C. (2002). Diets enriched in foods with high antioxidant activity reverse age-induced decreases in cerebellar beta-adrenergic function and increases in pro-inflammatory cytokines. The J. of Neurosci., 22(14):6114-6120.
- Gindler,E. M.,and king,J.D. (1972). Rapid colorimetric determination of calcium in biologic fluids with methylthymol blue, Am.J. Clin. Path 58:376-382.
- Ishikawa,T.;Fujiyama,Y.;;Iganashi,C. *et.al.* (1989).Clinical familial Hypercholesterolemia. Athersclorsis ,75:95-103.
- Isik,O.;Hizarci,L.;Sayin,S.; Gokpinar,S.;Durmaz, Y. and Goksan, T. (2006). The effect of environmental factors on the vitamin C (ascorbic acid) E (alphatopherol), carotene and the fatty composition of *S. platensis*.E.U. J. of Fisher. and Aqu. Sc :23,(3-4) :357-361.
- IUPAC (1982). Standard method for analysis of oil fats and drivitives.7th ed. IUPAC 1982 2 – 301 IUPAC method 2 -3012-303.Blackwel Sci. Pub.Oxford.
- John D. Brunzell, MD, FACP¹; Michael Davidson, MD, FACC², Curt D. Furberg, MD, PhD³; Ronald B. Goldberg, MD⁴; Barbara V. Howard, PhD⁵; James H. Stein, MD, FACC, FACP⁶ and Joseph L. Witztum, MD⁷ (2008).Lipoprotein Management in Patients With cardiometabolic Risk, J. Am. Coll. Cardiol.; 51:1512-1524.
- Khan, M.; Chandra ,J. S. ; Mohan ,I. K.;; Umamaheswara R. Naidu M; .Sundaram ,C.; Singh, S.; Kuppusamy, P.; Kutala ,V.K. (2005). Protective effect of *Spirulina* against doxorubicin-induced cardiotoxicity. Phytoth. Res.; 19(12):1030-7.
- Kumar, N.; Kumar, P., and Singh, S. (2010).Immunomodulatory effect of dietary *S.*

- platensis* type II collagen induced arthritits in rats .RJPBCS,1(4):877-885.
24. Lopez- Virella ,M; Stone,P.;Ellis, S.; Colwell, I.(1977). Cholesterol determination in high density lipoprotein separated by three different methods . Clin. Chem. 23,882-884.
 25. Lovelock J.E., and Porterfield,B.M.(1952). Blood clotting the function of electrolytes and calcium. Biochem. J. 50: 416-420.
 26. Manz, U., and Vuilleumier, J.P.(1988). Determination of added canthaxanthin in complete feeds and premixes with HPLC .P.68-71. In: Analytical methods for vitamins and carotinoids in feed. H.E. Keller, ed., F.Hoffmann. La Roch Ltd.Basel, Switherland.
 27. Mao,T.K.; Van de Water, J.,and Gershwin, M.E. (2005). Effect of *Spirulina* based dietary supplement on cytokine production from allergic rhinitis patients .J.of Medicinal Food 8(1)27-30.
 28. Misbahuddin,M.;AZM Maidul Islam; Salamat Khandker; Ifthaker-Al-Mahmud, Nazrul Islam and Anjumanara (2006). Efficacy of *Spirulina* extract plus zinc in patients of chronic arsenic poisoning: a randomized placebo-controlled study. (Risk factors). J. of Toxicol. Clinical Toxicol.. 44.2 ,p135(7).
 29. Muhling, M.; Belay,A.,and Whitton,A.B. (2005). Variation in fatty acids composition of *Anthrospira (S.) platensis* strain.J. Appl. Phycol.17:137-146.
 30. Nakaya, N. : Homma, Y. ,and Goto, Y(1988). Cholesterol lowering effect of *Spirulina*. Nutr-Rep-Int. Stoneham, Mass.: Butterworth Publishers.. 37 (6) : 1329-1337.
 31. Noaman,N.H. ; Khaleafa, A.F., and.Zaki, S.H. (2004). Factors affecting antimicrobial activity of *Synechococcus leopoliensis*.Microbiol. Res.195:395-402.
 32. Orino, K.; Lehman, L.; Tsuji, Y.; Ayaki, H.; Torti, S. V.,and Torti, F. M. (2001)..Ferritin and the response to oxidative stress. Biochem.J.357 (Pt 1): 241.
 33. [Otle S](#),and [Pire R](#). (2001) .Fatty acid composition of *Chlorella* and *Spirulina* microalgae species. [J AOAC Int.](#),84(6):1708-14.
 34. Olivera,MA.C.;Monterio,M.P.C.;Robbs,P.G.,and Leite,S.G.F. (1999).Growth and chemical composition of *S. maxima* and *S. platensis* at different temp.Aquaculture Int. ,7:261-275.
 35. Park, H.; Lee, Y.; Ryu, H.; Kim, M.; Chung, H.,and Kim, W. (2008). A randomized double-blind, placebo-controlled study to establish the effects of *Spirulina* in elderly Koreans. Annals of nutr. and met. ,52 (4): 322-328.
 36. Quereshi *et. al.* (1994). Poultry Science Assoc. Dept. of Poultry Sci, Nnorth Carolina States ,NC.Pub in :J. of Poultry Science, 73 SI. P . 46 USA . Immune enhancement potential of *Spirulina* in chicken.
 37. Quoc,K.P., and J.P. Dubacq(1997).Effect of growth temp on the biosynthesis of eukaryotic lipid molecular species by the cyanobacterium *S. platensis* .Biochemica et Biophysica Acta. 1346:237-246 .Elsevier.
 38. Qian, H., and Sheng, M. (1998). Simultaneous determination of fat soluble vitamins A, D and E and pro-vitamins D₂ in animals feed by one step extraction and high performance liquid chromatography analysis. Chromtog.A.825:127-133.
 39. Ren ,J. ; Grundy, S.M., Liu, J.; Wang ,W, Wang M, Sun J, Liu J, Li Y, Wu Z, Zhao D. (2010). Long term coronary heart disease risk associated with very-low-density lipoprotein cholesterol in Chinese: The result of a 15-year Chinese multi-provincial cohort study (CMCS). Atherosclerosis. ;211(1):327-32.
 40. Richmond, W. (1973).Preparation and properties of cholesterol oxidase from *Nocardia sp.*and its application to the enzymatic assay to total cholesterol in serum., Clin Chem. ,19:1350.
 41. Raj,M.; Abd Elmageed, Z.; Zhou, J. *et. al.* (2008). Synergistic action of dietary phyto-antioxidants on survival and proliferation of ovarian cancer cells. Gynecol. Oncol.;110:432-438.
 42. Romano,I.; Belitti,M.;Nicolaus B.;Lama,I.; Manca,C. M.; Pagnotta C,and Ganbact,C.(2000).A lipid profile : a useful chemotaxonomic marker for clear classification of a new cyanobacteria in *Spirulina* genus .Phytochem.:54:289-294.
 43. Seckback, J. (1982). Ferriting out the secrets of plant ferritin - A review. J. of Plant Nutrition 5: 369-394.
 44. Trinder ,P.(1969). Determination of glucose in blood using oxidase with an alternative glucose acceptor .Ann.Clin. Chem.,6:24-27.
 45. Tomaselli,L.;Giovannetti, L.;Sacchi, ,A.,and Bocci F,(1988).Effect of temp. on growth and biochemical composition of *S. platensis* strain M2. In: Algal Biotechnology T. Stadler ;J.Mellion; Mc,Verdus;Y.Kaeamanos;H. Morvan; D. Christiaen(eds)Elseiver Appl.Sci. London. pp: 303-314.
 46. [Torres-Duran; Ferreira-Hermosillo, and Juarez-Oropeza. \(2007\). Antihyperlipemic and antihypertensive effects of *S. maxima* in an open sample of mexican population: A preliminary report. Lipids in Health and Disease. 6, 33.](#)
 47. Watanabe, F.; Takenaka ,S.; Kittaka-Katsura ,H.; Ebara, S. ,and Miyamoto, E. (2002).

- Characterization and bioavailability of vitamin B₁₂-compounds from edible algae. J. of Nutrit.Sci. and Vitamin., 48 (5):325-331.
48. Wang, Y; Chang CF; Chou J; Deng, X; Harvey, BK; Cadet JL, and Bickford, PC. (2005). Dietary supplementation with blueberries, spinach, or *Spirulina* reduces ischemic brain damage. Exp. Neur;193(1):75-84.
49. White, D; kramaer, JE; Johnson,G; Dick.F, and Hamilton ,H.(1986).:Am.J.Clin. path., 72:346.
50. Yin, J; Zuberi, A; Gao, Z; Liu, D; Liu, Z; Cefalu, WT., and Ye, J . (2008) . [Effect of *Shilianhua* extract and its fractions on body weight of obese mice](#)". *Metabolism: Clin.and Exp.*57 (7 Suppl 1): S47-51.
51. Zingg, JM.; Azzi, A (2004) .Non-antioxidant activities of vitamin E". *Current medicinal chemistry* 11 (9): 1113-33.

12/2/2010