

## Detection of Community Acquired Methicillin Resistance *Staphylococcus aureus* among *Staphylococcus aureus* isolates.

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**Abstract:** The rates of Methicillin-resistant *Staphylococcus aureus* (MRSA) infections in the hospital, as well as the disease in the community, have continued to rise. Staphylococcal cassette chromosome *mec* (*SCCmec*) is a variable genetic element that contains the methicillin resistance determinant, *mecA*. *SCCmec* typing is one of the most important molecular tools available for distinction between community-acquired MRSA (CA-MRSA) and hospital-associated MRSA (HA-MRSA) occurring on a worldwide basis. CA-MRSA has been reported to carry the loci for Panton Valentin leukocidin (PVL) in high frequency in association with the type IV *SCCmec*. The present study aimed to differentiate between HA-MRSA and CA-MRSA by detection of *SCCmec* and determination the prevalence of PVL gene among MRSA isolates. A total of 34 *Staphylococcus aureus* isolates were included in this study. Susceptibility of Staphylococci was determined by, Disc diffusion method including methicillin, oxacillin and cefoxitin discs. Penicillin Binding Protein (PBP<sub>2a</sub>) Latex Agglutination test was done to detect the presence of PBP<sub>2a</sub> responsible for methicillin resistance. In addition genotypic identification of MRSA was carried out by detecting *mec* gene by real time PCR. Conventional PCR was carried using different set of primers for the amplification of *SCC mec* for differentiating the HA-MRSA and CA-MRSA; moreover detection of *PVL* as virulence factor was also done. The antibiotic sensitivity of CA-MRSA ranged from 11.76% for ceftazidime to 47.06% for Imipenem, Erythromycin and Gentamycin; while the sensitivity of HA-MRSA ranged from 2.94% for Amoxicillin and Ampicillin/sulbactam to 29.41% for Amikin. Out of 34 *S. aureus* strains; 26(76.47%) isolates were found to be resistant to oxacillin disc, 30(88.24%) isolates were resistant to methicillin; and all strains were resistant to cefoxitin disc. All MRSA strains were confirmed to be methicillin resistant by detection of *mecA* gene using real time PCR. Out of 34 MRSA strains 32 (94.12%) were PBP<sub>2a</sub> producer. In the present study, though, the majority (25 out of 34) of our strains were not *SCC mec* typable, yet among the nine typable strains the six hospital strains belonged to type II and III as reported in the literature and the three CA-MRSA belonged to the novel type V reported by other workers to be associated with CA-MRSA and the only PVL positive CA MRSA strain was untypable.

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### 1. Introduction:

Methicillin-resistant *Staphylococcus aureus* (MRSA) is a serious threat to hospitalized patients globally and now represents a challenge for public health, as community-acquired infections appear to be on the increase in various regions and countries<sup>(1,2)</sup> including North America<sup>(3)</sup>, Australia<sup>(1)</sup>, Saudi Arabia<sup>(4)</sup> and Finland<sup>(5)</sup>. Rising colonization rates lead to increased infection rates in the community and in hospitals. The consequence to the health care system is longer hospital stays and greater costs<sup>(6)</sup>. Patient risks include significantly higher mortality and morbidity rates with invasive MRSA infection<sup>(7)</sup>.

MRSA strains produce penicillin binding protein<sub>2a</sub>, which possesses reduced affinities for binding to lactam antibiotics resulting in lactam resistance. The *mecA* gene, encoding PBP<sub>2a</sub>, is carried on a peculiar type of mobile genetic element

inserted into the staphylococcal chromosome, designated staphylococcal cassette chromosome *mec* (*SCCmec*) elements. *SCCmec* elements typically share four characteristics: first, they carry the *mec* gene complex (*mec*) consisting of the methicillin resistance determinant *mecA* and its regulatory genes and insertion sequences; second, they carry the *ccr* gene complex (*ccr*) consisting of *ccr* genes that are responsible for the mobility of the element and its surrounding sequences; third, they have characteristic directly repeated nucleotide sequences and inverted complementary sequences at both ends; and last, they integrate into the 3' end of an open reading frame (ORF), *orfX*<sup>(8)</sup>.

In *S. aureus*, three major *mec* complex classes have been described, and concerning the *ccrAB* locus, three major allotypes (*ccrAB1* to *ccrAB3*)<sup>(9)</sup> and one sporadic allotype (*ccrAB4*)<sup>(10)</sup>

have been identified. Recently, a new type of *ccr* gene complex, which consists of only one gene (*ccrC*) not closely related to the *ccrA* or *ccrB* gene, was reported<sup>(11)</sup>. *SCCmec* carries other sequences that define the overall genetic organization of the resistance cassette. These regions may be used as targets for typing strategies<sup>(12)</sup>, and polymorphisms within these regions, particularly in the region downstream of the *ccrAB* genes (the J1 region), define *SCCmec* subtypes or variants<sup>(13)</sup>.

Based on the class of *mecA* gene complex and the type of *ccr* gene complex present, *SCCmec* cassettes are classified into seven major types<sup>(14)</sup>. The hospital-associated strains of MRSA (HA-MRSA) strains contain the larger type I, II, or III cassettes<sup>(13)</sup>, while the two smallest *SCCmec* types, *SCCmec* IV and *SCCmec* V, have been associated with community-associated MRSA (CA-MRSA)<sup>(2,15)</sup>.

Clinically, CA-MRSA usually causes skin and soft tissue infection. However, it can cause serious life-threatening conditions, which in addition to necrotizing pneumonia include necrotizing fasciitis, bloodstream infection, and septic shock<sup>(16)</sup>. Reports have suggested that certain strains of CA-MRSA may be more virulent than HA-MRSA<sup>(17, 18)</sup>. The expression of Pantone-Valentine leukocidin (PVL), a two-component, pore-forming, cytolytic toxin that targets mononuclear and polymorphonuclear cells and causes cell death by necrosis or apoptosis, has been strongly associated with CA-MRSA<sup>(17)</sup>. The PVL toxin consists of two synergistic proteins, LukS-PVL and LukF-PVL, encoded by the *pvl* genes *lukF* and *lukS*, which are carried on a temperate bacteriophage.<sup>(2, 19)</sup>

Clinicians are now faced with emergence of (CA-MRSA) strains that are genetically different from MRSA strains originating in the hospital. Moreover, with the recent trend and shift in epidemiology, CA-MRSA is now being found in hospitals and in some instances displacing classic (HA-MRSA)<sup>(20)</sup>.

The present study aimed to differentiate between hospital acquired MRSA and community acquired MRSA by detecting different Staphylococcal Chromosomal Cassette *mec* types (*SCCmec*), and detecting the prevalence of Pantone Valentine Leukocidin as virulence factor for CA-MRSA and determining whether its carriage could be used as a surrogate marker for CAMRSA

## 2. Material and Methods

### I-Bacterial Isolates:

Thirty nine Staphylococcal isolates were included in this study. Data recording previous hospital admission during the 6 months ago were collected.

### I-Identification of staphylococcal isolates:

The staphylococcal isolates were identified morphologically and biochemically by standard laboratory procedures<sup>(21)</sup>. The coagulase plasma test was performed on organisms exhibiting typical staphylococcal colony morphology to allow for discrimination of *S. aureus* from coagulase-negative staphylococci.

### II-Antibiotic susceptibility testing:

Methicillin resistance (using Methicillin, Oxacillin and Cefoxitin) and susceptibility to different antibiotics were determined by the agar disk diffusion method<sup>(22)</sup>.

### III-Detection of Penicillin Binding Protein 2a (PBP2a)

Latex Agglutination Test (Oxoid®) based on the agglutination of latex particles sensitized with monoclonal antibodies against PBP2a, was used according to the manufacturer's instructions<sup>(23)</sup>. Agglutination was visualized and was scored as positive, negative, or weakly positive.

### IV- Polymerase Chain Reaction (PCR)

#### Staphylococcal DNA extraction:

Staphylococcal colonies were emulsified in 200 µl sterile distilled water to produce a heavy suspension, and heated at 100°C for 15 min. then centrifuged at 14,000 rpm for 5 min.

PCR for *mecA* gene detection was performed using Real time (TaqMan) PCR.<sup>(24)</sup>

The primers and probe used were as follows:

Forward primer,  
5'TGCTAAAGTTCAAAGAGTATTTATAACAA  
CA 3'; Reverse primer,  
5'TGTGCTTACAAGTGCTAATAATTCACC 3';  
and

Probe, 5' FAM-  
ATTATGGCTCAGGTACTGCTATCCACCCTCAA  
A-TAMRA 3'. The PCR mixture was prepared using TaqMan® Universal PCR Master Mix (2X) with final PCR mixture volume of 25 µl. Five µl of template DNA and 30 pmole of each primer and 7.5 pmol of probe were added to each test. A negative control was prepared by the addition of the same contents to the tube with water instead of the extract. Amplification was performed using MX3000P™ (Stratagene) Real Time PCR System programmed to hold at 95°C for 10 min for AmpliTaq gold activation and 30 cycles of denaturation at 95°C for 15 sec and annealing and extension at 60°C for 1min with end point fluorescence detection.

PCR for detection of *SCCmec* gene and *PVL*: Polymerase chain reaction for detection of *SCCmec* and *PVL* gene was performed using genomic DNA from each MRSA isolate as template. Primers used to amplify the different *SCCmec* and *PVL* are listed in table (1). For *SCCmec*, 2 sets of primers were used. The first set (Oliveira primers) is designed to type *SCCmec* (I-IV)<sup>(25)</sup> based on selected loci (A through F) upstream and downstream the *mecA* gene. Another set of primers (Zhang)<sup>(26)</sup> was used for detecting *SCCmec* type II, III and the newly described *SCCmec* type V.

#### PCR Conditions:

Eight µl of DNA extract were amplified by PCR in a final volume of 25 µl using 2x PCR master mix (Fermentas life sciences®) containing 0.05 units/µl of Taq DNA polymerase, 50 picomol of each primer, PCR buffer, 2 mM MgCl<sub>2</sub>, 0.2mM of each dNTP. A negative control was prepared by the

addition of the same contents to the tube with water instead of the extract. Amplification was performed in a Perkin-Elmer 9600 thermocycler. For Oliveira primers the cycle program was performed with an initial denaturation for 5 min at 94°C, then 35 cycles of denaturation for 30 sec at 94°C, annealing for 30 sec at 53°C and extension for 1min at 72°C and final extension for 5 min. While for Zhang primers: the cycles begin with an initial denaturation step at 94°C for 5 min followed by 10 cycles of 94°C for 45 seconds, 65°C for 45 seconds, and 72°C for 1.5 min and then another 25 cycles of 94°C for 45 seconds, 55°C for 45 seconds, and 72°C for 1.5 min, and end with a final extension step at 72°C for 10 min. For amplification of *PVL* gene<sup>(27)</sup>: the cycle program was performed with an initial denaturation for 5 min at 94°C, then 35 cycles of denaturation for 40 sec at 94°C, annealing for 40 sec at 53°C and extension for 1min at 72°C and final extension for 10 min.

**Table (1): Primers used for amplification of *SCCmec* and *PVL***

Locus	Oliveira's Primers <sup>(25)</sup>	Oligonucleotide sequence (5'-3')	Amplicon size (bp)	Specificity ( <i>SCCmec</i> type)
A	CIF2 F2 CIF2 R2	TTTCGAGTTGCTGATGAAGAAGG ATTTACCACAAGGACTACCAGC	495	I
B	KDP F1 KDP R1	AATCATCTGCCATTGGTGATGC CGAATGAAGTGAAAGAAAGTGG	284	II
C	MECI P2 MECI P3	ATCAAGACTTGCATTCAGGC GCGGTTTCAATTCACCTTGTC	209	II,III
D	DCS F2 DCS R1	CATCCTATGATAGCTTGGTGTC CTAAATCATAGCCATGACCG	342	I,II,IV
E	RIF4 F3 RIF4 R9	GTGATTGTTTCGAGATATGTGG CGCTTTATCTGTATCTATCGC	243	III
F	RIF5F10 RIF5R13	TTCTTAAGTACACGCTGAATCG GTCACAGTAATTCATCAATGC	414	III
<b>Zhang's Primers:</b> <sup>(26)</sup>		Oligonucleotide sequence (5'-3')	Amplicon size (bp)	Specificity
Type II-F Type II-R		CGTTGAAGATGATGAAGCG CGAAATCAATGGTTAATGGACC	398	<i>SCCmec</i> II
Type III-F Type III-R		CCATATTGTGTACGATGCG CCTTAGTTGTCGTAACAGATCG	280	<i>SCCmec</i> III
Type V-F Type V-R		GAACATTGTTACTTAAATGAGCG TGAAAGTTGTACCCCTTGACACC	325	<i>SCCmec</i> V
<b>PVL primers:</b> <sup>(27)</sup>		Oligonucleotide sequence (5'-3')	Amplicon size (bp)	Specificity
PVL-1 PVL -2		ATCATTAGGTAATAATGTCTGGACATGATCCA GCATCAASTGTATTGGATAGCAAAAAGC	433bp	PVL

Detection of PCR products was done using 2% agarose gel stained with ethidium bromide and using of the molecular weight markers (50 bp DNA ladder (Fermentas Life Sciences)).

### 3. Results

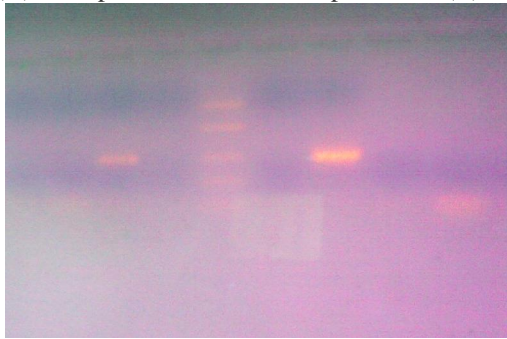
1-Bacterial isolates and identification: Among the 39 *Staphylococcus* isolates, 34 (87.18%) were free coagulase positive by tube coagulase test.

2-Detection of methicillin resistance by Disc diffusion method: out of 34 *S. aureus* strains; 26(76.47%) isolates were resistant to oxacillin disc, 30(88.24%) to methicillin and all strains were resistant to ceftaxime disc (100%).

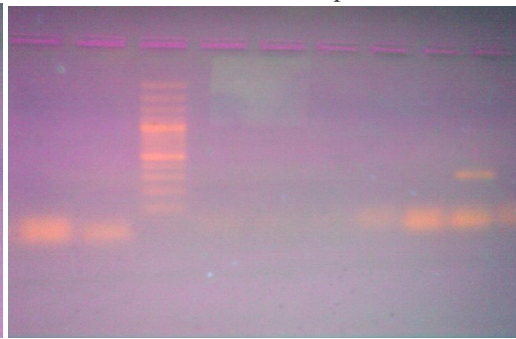
3-Antibiotic susceptibility: According to data regarding previous hospital admission, the 34 collected strains were divided into 18 CA-MRSA & 16 HA-MRSA strain. CA-MRSA was more sensitive, the sensitivity in CA-MRSA ranged from (22.22% for ceftazidime) to (88.89% for Imipenem, Erythromycin and Gentamycin); while the sensitivity in HA MRSA ranged from (6.25% for Amoxicillin and Ampicillin/sulbactam) to (62.5% for Amikacin). All MRSA were sensitive to vancomycin.

4-Detection of *mec A* gene: All the 34 strains were *mecA* gene positive. The presence of *mecA* gene

(A) 284bp DNA ladder 284bp (B)

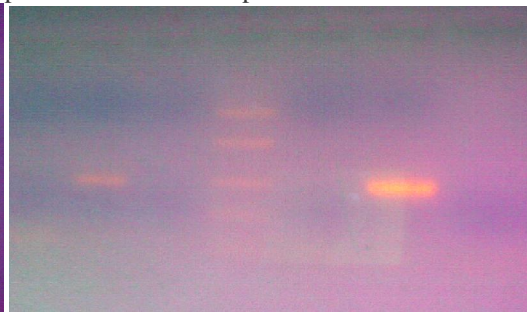
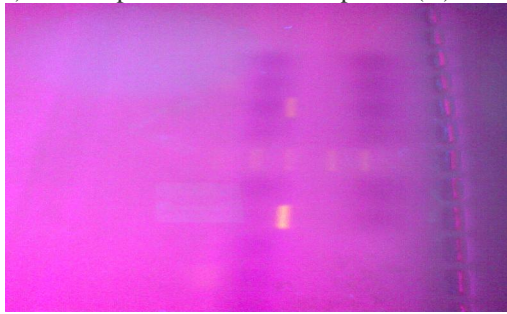


DNA ladder 243bp



**Fig (1A):** Ethidium bromide stained agarose gel showing a single band of 284 bp specific for SCC *mec* type II at lanes 3 and 7. Lane 5 shows 50 bp DNA ladder. **Fig (1B):** show a single band of 243 bp specific for SCC *mec* type III at lanes 11. Lane 5 shows 100 bp DNA ladder.

(C) 398 bp DNA ladder 398 bp (D) 325bp DNA ladder 325bp



**Fig (1C):** Ethidium bromide stained agarose gel showing a single band of 398 bp specific for SCC *mec* type II at lanes 6 and 10. Lane 8 shows 50 bp DNA ladder. **Fig (1D):** show a single band of 325bp specific for SCC *mec* type IV at lanes 3 and 3. Lane 12 shows 50 bp DNA ladder.

detected by real time PCR was considered to be the gold standard in evaluating the 3 disc diffusion methods to identify MRSA.

5- Gene expression (PBP<sub>2a</sub> Production) among the 34 *mecA* positive: out of 34 MRSA strains 32 (94.12%) were PBP<sub>2a</sub> producer while 2 (5.88%) were PBP<sub>2a</sub> non producer. The 2 PBP<sub>2a</sub> non producers isolates were CA MRSA.

6- *SCC mec* Typing:

Out Of the 16 HA-MRSA: 5 isolates were *SCCmec* type II (Two by Oliveira primers and three by zhang primers) and only one was detected as type III. While out of the 18 CA-MRSA Three isolates belonged to the newly described Type V reported to be associated with CA MRSA. The remaining 25 MRSA strains could not be typed.(Fig 1(A-E))

7-Detection of PVL: only one strain (CA-MRSA) out of the 34 MRSA was positive for Pantone Valentine leukocidin (PVL) by PCR. (Fig 2)

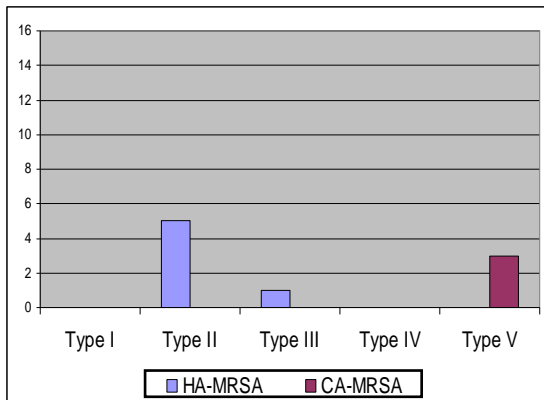


Figure 1E: SCC mec typing of the 34 MRSA

#### 4. Discussion:

Methicillin resistance of *S. aureus* remains to be a significant problem. Rapid and accurate determination of methicillin resistance is important for initiation of appropriate antimicrobial therapy. Misdiagnosing this resistance leads to treatment failures and spread of infections with these resistant strains.

Phenotypic techniques as disk diffusion and microdilution methods are employed in routine laboratories for the detection of methicillin resistance. However, these methods are often not entirely reliable at detecting some strains that harbor the *mecA* gene<sup>(28)</sup>. Identification of the *mecA* gene remains the most reliable method of detecting MRSA isolates, however not all laboratories can include molecular biology techniques in their routine clinical practice.

In this study cefoxitin disk diffusion tests was 100% sensitive for MRSA detection. Alternatively, only 76.47% and 88.24% isolates were resistant to Oxacillin and methicillin respectively. These results were in accordance with those of several studies<sup>(28-30)</sup>. This means that disc diffusion testing using cefoxitin disc is far superior to most of the currently recommended phenotypic methods and is now an accepted method for the detection of MRSA by many reference groups including CLSI<sup>(31)</sup>.

Identification of MRSA, is more accurate by either directly detect the gene encoding the methicillin resistance determinant (*mecA*) or its product (PBP2a)<sup>(32)</sup>. MRSA-Screen test is a rapid and simple to perform method. Many studies reported that the accuracy of MRSA screen latex agglutination test for detection of PBP2a approaches the accuracy of PCR and more accurate than susceptibility testing methods with sensitivity of 97%-100% and a

Fig 2: 433bp of PVL DNA ladder

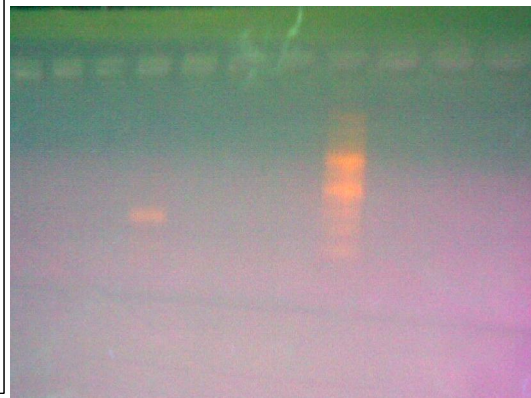


Fig 2: Ethidium bromide stained agarose gel showing a single band of 433 bp specific for PVL at lane 5. Lane 9 shows 100 bp DNA ladder

only two (5.88%) PBP2a non producer isolates (false negative) were identified. Other authors<sup>(35, 36)</sup> reported that false-negative results may occur with MRSA isolates with low oxacillin MICs (4 or 8 µg/ml) due to production of smaller amounts of PBP2a or the failure to express the gene phenotypically.

MRSA infection can be categorized into 2 distinct groups: HA-MRSA and CA-MRSA. The community isolates are distinctly different from the hospital strains both epidemiologically and microbiologically<sup>(37)</sup>. Both CA-MRSA and HA-MRSA are resistant to traditional anti-staphylococcal -lactam antibiotics. However, CA-MRSA isolates tend to be more susceptible to other antibiotics (including to sulfa drugs, tetracyclines) than are HA-MRSA<sup>(38)</sup>, and their narrow spectrum of resistance is solely due to determinants harbored on genetic elements present on the SCC<sup>(39)</sup>. In this study, most of the 18 CA-MRSA isolates were susceptible to tetracyclines (83.3%) Erythromycin (88.9%), Gentamycin (88.9%), co-trimoxazole (77.8%), Ofloxacin (83.3%). On the other hand, the 16 HA-MRSA showed a wide spectrum of drug resistance ranging from 81.25% (ofloxacin, gentamycin and tetracyclines) to 68.7 % (erythromycin) and 56.2% (co-trimoxazole). Comparable results were obtained by various authors<sup>(40, 41)</sup>. Kaplan<sup>(40)</sup> noted that most of the CA-MRSA isolates are susceptible to most antibiotics, while Anbumani *et al.*,<sup>(41)</sup> reported that 250 MRSA isolated from different clinical specimens were multi-drug resistant.

SCCmec typing is one of the most important molecular tools available for understanding the epidemiology and strain relatedness of MRSA<sup>(26)</sup>. In the present study, 2 sets of primers were used in an

attempt to type our MRSA strains. The first set was chosen according to Oliviera typing scheme designed to type *Sccmec* (I-IV)<sup>(25)</sup>, and using six primer pairs only 3 of the 16 HA MRSA were typable by Oliviera primers. Two belonged SCC *mec* type II (amplified locus B) and one SCC *mec* type III. (amplified locus F), and none of the 18 CA MRSA could be typed by this scheme. As Oliveira's assay has limitations in detecting the newly described SCC*mec* type V, misclassifying them as type III, while failing to discriminate type IV into subtypes a, b, c, and d<sup>(26)</sup>. Also, because of difficulties in assay optimization another trial of typing the SCC *mec* was carried using Zhang set of primers unique and specific for SCC*mec* types. So, another 3 out of 16 HA MRSA were classified as SCC*mec* type II, moreover another three belonged to SCC*mec* type V among 18 CA MRSA. So using both typing methods only 9 out of the 34 MRSA were typable. Similar data were noticed by others<sup>(25,42)</sup>. Oliviera *et al.*,<sup>(25)</sup> reported that, 8% of their isolates were non-typeable for SCC*mec* by their primers used in this study. Also, Shore *et al.*,<sup>(42)</sup> used also two typing methods the first for amplification of the *ccr* and *mec* genes, and the second method of Oliveira and de Lencastre, used in the present study and (50%) out of their 172 isolates harbored two apparently different SCC*mec* elements when tested by both typing methods. They suggested that PCR amplification and sequencing of the entire SCC*mec* element was necessary for the complete characterization of the SCC*mec* elements harbored by isolates with ambiguous multiplex patterns.

The differentiation between the typical HA-MRSA and CA-MRSA strains based on epidemiologic definitions becomes difficult, along with molecular distinction based on the SCC*mec* type is beginning to blur.<sup>(43)</sup> In the present study, though, the majority (25 out of 34) of our strains were not SCC*mec* typable, yet among the nine typable strains the six hospital strains belonged to type II and III as reported in the literature and the three CA-MRSA belonged to the novel type V reported by other workers to be associated with CA-MRSA

CA-MRSA has been reported to carry the loci for PVL in high frequency, and to be associated with the type IV (SCC*mec*).<sup>(44)</sup> In the present study, the only PVL positive CA MRSA was untypable. This was contrary to that noted by Berglund *et al.*<sup>(45)</sup> who detected PVL genes in 66% of the CA-MRSA isolates. However, Holms *et al.*<sup>(46)</sup> revealed that the PVL genes are carried by a relatively low number (1.6%) of *S. aureus* isolates from their clinical laboratories, indicating an unequal distribution of the genes encoding PVL among their strains. In addition, the overall proportion of MRSA isolates carrying *pvl* was 1.8% among the 1,389 MRSA isolated from

Ireland. 7.5% of these isolates were CA-MRSA strains, of which only 6.7% carried *pvl* genes and the carriage of *pvl* was not restricted to CA-MRSA<sup>(47)</sup>. Similarly, 78% of CA-MRSA isolates referred to a central reference facility were *pvl* negative and 25% of *pvl* positive isolates in this group were HA. Additionally, Ko *et al.*<sup>(48)</sup> was not able to detect *pvl* gene, in any of their MRSA isolates. Moreover, Kim *et al.*<sup>(49)</sup> reported that none of their Korean CA-MRSA isolates contained the PVL genes. These findings agree with reports that carriage of *pvl* cannot be used as a sole marker for CA-MRSA<sup>(47)</sup>.

In the present study, the combination of SCC*mec* typing, in addition to the detection of PVL was not sufficient to discriminate between HA and CA MRSA due to the near absence of PVL among the CA MRSA and the limited capacity of the SCC*mec* typing methods among our strains.

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