

Improving gladiolus growth, flower keeping quality by using some vitamins application

¹Bedour, A. Abo Leila and ^{2*}Rawia, A. Eid

Water Relations and Field Irrigation Dept., Ornamental Plant and Woody Trees Dept., National Research Centre, Dokki, Cairo, Egypt. *Corresponding author, emil,rawiaabdelhady@yahoo.com.

Abstract: Response of growth, flowering quality and active chemical constituents of gladiolus plants by using some vitamins such as, thiamin, ascorbic acid and their combination during two seasons were studied. Plant which received the combined treatments of both vitamins recorded the highest growth, flowers quality and cornelets induction. Thiamine treatments had the lowest effect on photosynthetic pigments, while 200 ppm, thiamin+ 200ppm ascorbic acid, improved growth, delayed flowering opening of vase life, stimulated accumulation of carbohydrate and increased photosynthetic pigments and macronutrients status. Photosynthetic pigments and macronutrients. [Bedour, A. Abo Leila and Rawia, A. Eid. **Improving gladiolus growth, flower keeping quality by using some vitamins application.** Journal of American Science 2011;7(3):169-174]. (ISSN: 1545-1003). <http://www.americanscience.org>.

Keywords: gladiolus plant flower quality, vase life and chemical constituents

1. Introduction

Gladiolus flowers is considered a main exportable ornamental plants in Egypt, and the flower can be available the year around, the foreign markets demand Egyptian gladiolus with higher quality. Ascorbic acid (vitamins C) is a product of D-glucose metabolism in higher plants which affect on plant growth and development, and play a role in electron transport system (El-Kobisy et al., (1). Also, Smirnoff et al., (2) proposed a biosynthetic pathway and identified novel some enzymes. They also reported that ascorbate is synthesized from L-galactose via GDP-mannose and GDP- L galactose. Ascorbic acid also has been associated with several types of biological activities in plants such as in enzyme co factors, antioxidant, and as a donor / acceptor in electron transport at the plasma membrane or in the chloroplast (Conklin (3). A high level of endogenous ascorbate is essential effectively to maintain the antioxidant system that protects plants from oxidative damage Cheruth (4). Further, Farahat et al., (5) on *Cupressus sempervirn* L. reported that foliar application of ascorbic acid caused pronounced increases in vegetative growth and chemical constituents as well as essential oil percent.

Thiamine (vitamin B1) is a necessary ingredient for the biosynthesis of co-enzyme. Thiamine pyrophosphates, so it plays an important role in carbohydrate metabolism in plant. It synthesis in leaves and it transported to roots where it controls growth. Thiamine is an important factor for the translocation reactions of the pentose phosphate cycle which provides pentose phosphate for nucleotide synthesis and for the reduction of NADP required for various synthetic.

Youssef and Talaat (6) found that pronounced increases in vegetative growth and chemical constituents of rosemary plants by foliar application of thiamine were observed.

The objective of the present study was to investigate the effect of ascorbic acid and thiamine on improving the flower characters and chemical constituents of gladiolus plant.

2. Materials and methods

A two pot experiment were carried out at the greenhouse of National Research Centre, Dokki, Egypt, during the two successive growing seasons of 2007/ 2008 and 2008/2009 to investigate the effect of ascorbic acid and thiamine on improving growth, flower characters and chemical constituents of gladiolus plant. The effect of gladiolus corms were kindly supplied by ornamental plants research, Ministry of Agriculture, Giza, Egypt. Ninform corms were sown on the first week of December in two seasons using plastic pots (30 cm diameter) that were filled with loamy sand soil, physical and chemical properties are illustrated in Table (1) which determined according to Jackson (7). Gladiolus corms were irrigated regularly with tap water to reach 80 % of water holding capacity of the soil by weighting the pots daily and the needed amount of water was added. All the normal culture practices of growing gladiolus corms were applied as usual manner. The experiment including 8 treatments in addition to the control which were two concentrations of thiamine (Th) and ascorbic acid (ASC) 100 and 200 ppm applied each separately and/or in combination at concentrations of (100Th+100 ASC), (100 Th 100 + 200ASC), (200Th + 100 ASC) and (200 Th + 200 Th), to the respectively in addition untreated plants.

Table (1): Some physical and chemical properties of the used soils.

| Properties | Value | Nutrient content | | Value |
|----------------------|------------|------------------|---------------|-------|
| Clay % | 13.1 | P | mg/100 g soil | 13.4 |
| Silt % | 24.1 | K | | 59.2 |
| Sand % | 62.8 | Mg | | 17.4 |
| Texture | Loamy sand | Fe | ppm | 5.8 |
| EC dSm ⁻¹ | 1.98 | Mn | | 8.3 |
| pH | 7.73 | Zn | | 0.84 |
| CaCO ₃ % | 2.15 | Cu | | 0.96 |

Foliar application of thiamine and ascorbic acid were carried out two times of 30 days intervals, starting at the first week of February at both seasons. The treatments were arranged in a complete randomized block design with six replicates each replicate contained four pots and each pot contained four plants.

At the flowering stage sample was taken from representative three replicates randomly for each treatment and other three replicates for vase life, and the following parameters were determined the parameters included the following data , plant height (cm), no. of leaves per plant, as well as fresh and dry weights of leaves (g/plant), no. of florets/spike, No. of cormlets, and fresh and dry weight of cormlets (g/plant).

The flower spikes were cuts in the 7.00 am, wrapped in Kraft paper in groups inside treatments and translocated to the laboratory in cool water for 3 hours. The experiment design was a completely randomized with three replicates with five spikes per treatments.

The spikes were placed in glass bottles containing of 500 ml of water and kept into laboratory at room temperature for 15 days and the following data were recorded

Percentage of flowers wilted (on 6 days) total water uptake by the spikes and vase-life (days)

Water uptake (ml/l) by the cut spikes was estimated by subtracting the amount of water at the end of experiment from the initial volume.

Flower opening percentage after 24 h of placing the spikes in glass bottle congaing of water.

Chemical analysis

Photosynthesis pigments (chlorophyll a , chlorophyll b)and total chlorophyll (a+b) as well as carotenoids content were determined in fresh leaves as mg/g FW according to **Saric et al., (8)**, Total carbohydrates were determined using colorimetric method as described by **Herbert et al., (9)**. Total nitrogen was determined by the methods of **Chapman and Pratt (10)** while phosphorus

determination was carried out calorimetrically according to **King (11)**. Potassium was determined photo metrically by flame photometer method as described by **Brown and Lillard (12)**.

Statistical analysis:

The recorded data (mean of the two seasons) were statistically analyzed on complete randomized design according to the procedure of **Snedcor and Cochran (13)** Means were compared by least significant differences(LSD 5% levels of probability).

3. Results

Effect of thiamine and ascorbic acid and their combination

Growth parameters:

It is evident from data in Table (2) that there are significant differences among the treatments and control in plant height, leaves number, fresh and dry weights (g/plant). The combined treatment of 200ppm thiamine +200 ascorbic acid followed by 200 thiamine + 100 ppm ascorbic acid were superior to the other treatments and control in previous mentioned parameters. The superiority of the combined treatment over the control was by about 63.41% for plant height, 53.98 % for number of leaves and 17.24 and 62.57 % for fresh and dry weights.

Table (2): Growth parameters of gladiolus plants as affected by some vitamins (mean of the two seasons).

| Treatments | Plant height (cm) | Number of leaves /pl. | Leaves fresh weight | Leaves dry weight | FW /plant | DW/ plant |
|-------------------------|-------------------|-----------------------|---------------------|-------------------|-----------|-----------|
| | | | (g/plant) | | | |
| Control | 33.4 | 5.2 | 5.81 | 1.95 | 25.63 | 4.31 |
| 100 ppm Th | 42.2 | 6.5 | 7.52 | 1.54 | 28.35 | 5.08 |
| 200 ppm Th | 51.6 | 6.9 | 8.21 | 1.74 | 34.83 | 6.17 |
| 100 ppm ASC | 47.8 | 8.4 | 10.54 | 2.07 | 36.74 | 6.97 |
| 200 ppm ASC | 58.7 | 10.5 | 13.83 | 2.92 | 53.20 | 8.14 |
| 100 ppm Th+ 100 ppm ASC | 49.4 | 7.7 | 9.35 | 1.84 | 56.21 | 9.16 |
| 100 ppm Th+ 200 ppm ASC | 83.2 | 9.8 | 12.86 | 2.76 | 58.43 | 10.21 |
| 200 ppm Th+ 100 ppm ASC | 88.0 | 10.7 | 14.58 | 3.62 | 59.09 | 11.34 |
| 200 ppm Th+ 200 ppm ASC | 91.3 | 11.3 | 15.85 | 3.81 | 59.61 | 11.52 |
| LSD (0.05) | 2.12 | 0.23 | 0.43 | 0.11 | 1.12 | 0.21 |

Th=Thiamine ASC=ascorbic acid

Florets:

Foliar application of ascorbic acid, thiamine and their combination improved florets characters of gladiolus compared with the control plants. Combined treatment of 200 ppm ascorbic acid + 200

ppm thiamine was more effective than each treatment alone and all combined treatments. However, single treatments of ascorbic acid showed intermediate effect (Table 3).

Table (3): Flower characters of gladiolus plants as affected by some vitamins (mean of the two seasons).

| Treatments | No. of florets/spike. | Spike length | Diameter of florets | Length of florets | FW of florets/spike | DW of florets/spike |
|--------------------------|-----------------------|--------------|---------------------|-------------------|---------------------|---------------------|
| | | | | | | |
| Control | 3.52 | 10.1 | 2.45 | 4.21 | 14.51 | 2.18 |
| 100 ppm Th | 3.84 | 14.8 | 3.83 | 5.83 | 14.83 | 2.37 |
| 200 ppm Th | 5.71 | 18.5 | 4.74 | 6.32 | 19.77 | 3.22 |
| 100 ppm ASC | 6.83 | 17.3 | 5.03 | 6.84 | 20.42 | 3.47 |
| 200 ppm ASC | 8.51 | 20.4 | 6.08 | 7.75 | 33.40 | 6.02 |
| 100 ppm T h+ 100 ppm ASC | 10.3 | 17.5 | 6.24 | 8.00 | 35.21 | 7.08 |
| 100 ppm T h+ 200 ppm ASC | 11.0 | 22.7 | 7.00 | 8.52 | 36.78 | 7.11 |
| 200 ppm T h+ 100 ppm ASC | 12.2 | 23.8 | 6.90 | 9.00 | 37.42 | 8.10 |
| 200 ppm T h+ 200 ppm ASC | 13.8 | 25.7 | 7.42 | 9.21 | 38.93 | 8.23 |
| LSD at 5% | 0.58 | 0.81 | 0.12 | 0.05 | 0.11 | 0.04 |

Th=Thiamine ASC=ascorbic acid

Cormelets:

Table (4) show that application of ascorbic acid, thiamine and their combination had promotive effect on cormelets production of gladiolus compared with the control plants.

The low values were obtained by using 100 and/or 200 ppm thiamine while the combined treatment of 200 ppm thiamine + 100 ppm ascorbic acid induced the highest values for number of cormelets, fresh and dry weights (g/plant). The increments amounted by 77.11 % for number of cormelets, 73.12 and 55.76 % for fresh and dry weights respectively over the control.

Table (4): Induction of gladiolus cormelets as affected by some vitamins (mean of the two seasons).

| Treatments | No. of cormelets | Cormelets FW | Cormelets DW |
|--------------------------|------------------|--------------|--------------|
| | | | |
| Control | 4.43 | 7.22 | 1.02 |
| 100 ppm Th | 5.81 | 9.10 | 1.21 |
| 200 ppm Th | 7.22 | 12.43 | 1.34 |
| 100 ppm ASC | 8.42 | 13.63 | 1.95 |
| 200 ppm ASC | 12.53 | 18.74 | 2.09 |
| 100 ppm T h+ 100 ppm ASC | 14.22 | 22.11 | 2.00 |
| 100 ppm T h+ 200 ppm ASC | 16.71 | 25.42 | 2.14 |
| 200 ppm T h+ 100 ppm ASC | 18.11 | 26.83 | 2.29 |
| 200 ppm T h+ 200 ppm ASC | 15.27 | 25.11 | 2.10 |
| LSD (0.05) | 0.56 | 0.31 | 0.04 |

Th=Thiamine ASC=ascorbic acid

Photosynthetic pigments:

Results in Table (5) indicated that spraying ascorbic acid, thiamine and/or their combination induced gladiolus leaves with intensive Chl a, Chl b and carotenoids compared with control plants. Single treatments of thiamine gave the lowest favorable effect on photosynthetic pigments Chl a, Chl b and carotenoids, while the combined treatment of 200 ppm thiamine + 100 ppm and/or 200 ppm ascorbic acid gave the greatest promoting effect.

Table (5): Photosynthetic pigments and carotenoids content as affected by some vitamins treatments(mean of the two seasons).

| Treatments | Chl (a) | Chl (b) | a+b | Carotenoids |
|--------------------------|---------|---------|-------|-------------|
| | | | | |
| Control | 0.513 | 0.216 | 0.729 | 0.483 |
| 100 ppm Th | 0.587 | 0.312 | 0.899 | 0.593 |
| 200 ppm Th | 0.583 | 0.311 | 0.894 | 0.611 |
| 100 ppm ASC | 0.634 | 0.345 | 0.979 | 0.573 |
| 200 ppm ASC | 0.687 | 0.367 | 1.054 | 0.613 |
| 100 ppm T h+ 100 ppm ASC | 0.693 | 0.389 | 1.082 | 0.634 |
| 100 ppm T h+ 200 ppm ASC | 0.745 | 0.422 | 1.167 | 0.678 |
| 200 ppm T h+ 100 ppm ASC | 0.814 | 0.432 | 1.246 | 0.719 |
| 200 ppm T h+ 200 ppm ASC | 0.810 | 0.430 | 1.240 | 0.713 |
| LSD.5% | 0.002 | 0.001 | 0.005 | 0.040 |

Th=Thiamine ASC=ascorbic acid

Macronutrients:

Table (6) indicated that spraying gladiolus plants with ascorbic acid, thiamine and /or their combination, gradually stimulated the concentration of N, P and K in gladiolus leaves, consequently single application of 200 ppm ascorbic acid increased N, P and K by about 74.16, 61.44 and 56.22 % over the control plants, respectively. While when 200 ppm was combined with 100 and /or 200 ppm thiamine, the highest P and K concentration were observed. However, it also clear that the combination between ascorbic acid and thiamine were more effective than each single treatment on N, P and K concentration.

Total Carbohydrates:

Data in Table (6) reveal also that vitamins treatments are favourable for increasing carbohydrate % whereas 200 ppm ascorbic acid + 200 ppm thiamine gave the greatest favourable effect, the increment over the control reached 68.18 %.

Table (6): Macronutrient status and total carbohydrates content of gladiolus plant as affected of some vitamins (mean of the two seasons).

| Treatments | N | P | K | Total carbohydrates (%) |
|-------------|-------|-------|-------|-------------------------|
| | | | | |
| Control | 1.321 | 0.080 | 12.34 | 13.45 |
| 100 ppm Th | 2.211 | 0.141 | 15.21 | 29.69 |
| 200 ppm Th | 3.341 | 0.146 | 18.32 | 21.23 |
| 100 ppm ASC | 4.423 | 0.152 | 18.79 | 30.45 |

| | | | | |
|-----------------------------|-------|-------|-------|-------|
| 200 ppm ASC | 5.112 | 0.288 | 20.41 | 32.34 |
| 100 ppm T h+ 100 ppm ASC | 5.642 | 0.219 | 22.32 | 31.23 |
| 100 ppm T h+ 200 ppm ASC | 6.812 | 0.238 | 25.43 | 35.26 |
| 200 ppm T h+ 100 ppm ASC | 7.234 | 0.346 | 28.22 | 32.84 |
| 200 ppm T h+ 200 ppm ASC | 7.200 | 0.368 | 28.71 | 35.84 |
| LSD 5% | 0.230 | 0.004 | 0.82 | 1.28 |

Th=Thiamine ASC=ascorbic acid

Vase life:

Table(7) show that spraying gladiolus plants with vitamins induced significant difference in flowers opened percentage, flowers wilted ,water uptake and vase life compared with untreated plants ,the combined treatments of 200ppm thiamine +100 and/or 200ppm ascorbic acid was the most effective treatments for delaying flowers opening% decreasing the wilting flowers and it has also increasing effect on vase life(day).Maximum water uptake by flowers were recorded by 200 ppm thiamine+100 ppm ascorbic acid treatment.

Table (7): Keeping quality of gladiolus cut flowers as affected by some vitamins (mean of the two seasons).

| Treatments | Flowers opened% | Flowers wilted% | Water uptake(ml) | Vase life (day) |
|-----------------------------|-----------------|-----------------|------------------|-----------------|
| Control | 88.34 | 9.21 | 4.60 | 11.41 |
| 100 ppm Th | 82.53 | 88.32 | 5.30 | 11.00 |
| 200 ppm Th | 78.41 | 75.43 | 6.22 | 19.21 |
| 100 ppm ASC | 71.25 | 86.24 | 6.40 | 19.82 |
| 200 ppm ASC | 60.43 | 72.32 | 7.00 | 21.21 |
| 100 ppm T h+ 100 ppm ASC | 57.48 | 66.43 | 7.30 | 20.41 |
| 100 ppm T h+ 200 ppm ASC | 50.23 | 64.25 | 8.10 | 23.11 |
| 200 ppm T h+ 100 ppm ASC | 48.41 | 55.61 | 9.30 | 25.64 |
| 200 ppm T h+ 200 ppm ASC | 45.08 | 55.72 | 9.5 | 25.22 |
| LSD (0.05) | 0.230 | 0.004 | 0.82 | 1.28 |

Th=Thiamine ASC=ascorbic acid

4.Discussion

As expected some vitamins such as ascorbic acid and thiamine used in this experiment led to increase growth parameters of gladiolus plants with regard to plant height (cm), number of leaves, fresh and dry weights(g/plant). The present observation are fully incorporate the finding of **El-Fawakhry and El-Tayeb (14)** on chrysanthemum, **Youssef et al (15)** on datura plants, **Mona and Iman (16)** on rose geranium and **Rawia et al., (17)** on *Jasminum grandiflorum*, **Nahed et al., (18)** on songoniu, **Tarraf et al., (19)** on lemonegrass, **farahate et al., (5)** on *Cypressus sempervirent*.The regulatory effect of thiamine ,on meristem plant growth and development indirectly

through enhancing the endogenous level of various growth factors as cytokinines and gibberellins **Youssef and Talaat (6)**. However, thiamine synthesized in the leaves and transported to root to control growth **Kawasaki (20)**.

Regarding ascorbic acid, **Price (21)** reported that it is the most abundant antioxidant which protect plant cell and currently considered to be regulators on plant growth owing to its effect on cell division and differentiation. Our results also shown that thiamine was less effective and has little influence than ascorbic acid in this respect. This could be evident from the work of **Sakr et al., (22)** on canola and **Abdel Aziz et al., (23)** on gladiolus. The increment is growth parameters due to ascorbic acid are largely due to stabilize member structures **Blockhina (24)**, modulating membrane fluidity in a similar manner to cholesterol and also membrane permeability to small ions and molecules (**Foyer,25**)., implicated in the regulation of cell division by influencing progression from BI to S phase of cell cycle (**Smirnoff (26)**).

Cormelet and flowers characters tended to increase in the presence of ascorbic acid and/or thiamine comparing with the case of complete absence of vitamin (control). Ascorbic acid was rather than thiamine tended to be more effective and showed the most beneficial effect, and the positive response appeared to be raised with rise in the concentration. The present observations are in agreement with the finding of **Sakr et al., (27)** on wheat. In this respect thiamine is an important factor for the translocation reaction of the pentose phosphate cycle, which provides pentose phosphate for nucleotide synthesis and for the role of NADP required for various synthetic pathway **Kawasaki, (20)**, the author also added that, thiamine is a necessary for biosynthesis of Co-enzyme thiamine pro-phosphate, so it plays important role in carbohydrate metabolism which reflected on the increase in cornelets weights and numbers. The stimulatory effect of thiamine on flower characters was demonstrated by **El-Fawakhry and El-Tayeb (14)** on chrysanthemum flowers and **Rawia et al., (17)** on Jasmine, **Abdel Aziz (23)** on gladiolus . Similar results were obtained by **El-Quesni et al., (28)** on *Hibiscus rose sinesis* L. Ascorbic acid has been associated with several types of biological activities in plants, such as enzyme Co-factor, as antioxidant and as a donor/or acceptor in electron transport at plasma membrane on the chloroplast. Such increments in cornelets number in our result s may be due to increase in photosynthetic pigments which reflect on carbohydrate content and cornelets weight **Conktn, (29)**.

Our results demonstrated that, single application of either ascorbic acid and/or thiamine led

to increments in carbohydrates %, photosynthetic pigments and mineral ions contents N, P and K in gladiolus leaves. Apparently these observations may be due to that thiamine is a necessary for the biosynthesis of Co-enzyme thiamine pyrophosphate which plays a role in carbohydrate metabolism (**Kawasaki, 20**). The promotive effect of thiamine on total carbohydrates content may be due to their important role in the biosynthesis of chlorophyll molecule which in turn affected chlorophyll content **Youssef and Talaat, (18)**.

Such observation were reported by **Hassanein (30)** on *Foeniculum vulgare* L. and **Abu Dahab (31)** on *phelodorn erubescens*, plants, they concluded that thiamine increase photosynthetic pigments in plants. The superiority of ascorbic acid in increasing growth parameters was accompanied by high promotive effect on the previous chemical constituents. Such finding show an analogy with those obtained by **Sakr et al., (22)** on Canola plants. In this connection ascorbic acid occurs in chloroplasts, vacuoles, mitochondria and cell wall (**Anderson et al., 32, Ravtenkranz et al., 33**). The concentration in chloroplast can be high and probably related to its control photosynthesis **Foyer, (34)**. Ascorbate acts as antioxidant (**Asoda, 35**). Co-enzyme co-factor **Davies et al.,(36)**. Electrons donor for photosynthetic and mitochondrial electron transport **Asard et al., (37)**. Ascorbate is produced of D-glucose metabolism which affects some nutritional cycle activity in higher plants **El-Kobisy et al., (1)**.

The present data indicating further that supplemental addition of both thiamine + ascorbic acid greatly improved growth and chemical constituents of gladiolus growth, cornelets and flower induction with high quality and long vase life. Such positive increase could be explained by the fact that vitamins considered as a bio-regulators compounds which in little concentration exerted profound influence upon plant growth and production and may be due to the synergetic effect between ascorbic acid and thiamine in increasing plant growth and metabolism. This result hold true with finding of **Rawia et al., (17)** on *jasminum grandiflorum*. Our results showed that the combined treatments between both vitamins ascorbic acid or thiamine sprayed to gladiolus plants induced flowers with high quality and long vase life. This observation are harmony with the finding of **Muhammad et al.,(38)** may be due to the pro motive effect of vitamins on most chemical constituents of plants.

References

1. El-Kobisy, D.S. ; Kady K.A.; Hedani ,R.A.and Agamy,R.A. (2005): Response of pea plant

- (*pisum sativum* L.) to treatment with ascorbic acid. Egypt. J. Appl. Sci., 20: 36-50.
2. Smirnoff N.; Conklin P.I and Loewus F.A. (2001): Biosynthesis of ascorbic acid in plants: renaissance. Annual Review of Plant Molecular Biology and Plant Physiology. 52: 437-467.
 3. Conklin,P.(2001) :Recent advances in the role and biosynthesis of ascorbic acid in plants. Plant cell environment , 24,383-394.
 4. Cheruth, A.J. (2009): Changes in non enzymatic antioxidants and ajmalicine production in *catharanthus roseus* with different soil salinity regimes. Botany Research International 2(1):1-6.
 5. Farahat, M.M.; Soad Ibrahim M.M., Lobna. T. S.and Fatma, El-Quesni, E.M. (2007): Response vegetative growth and some chemical constituents of *cupressus sempervim* L., to foliar application of ascorbic acid and zinc at Nubaria. World J. of Agric Sci., 3(3): 282-288.
 6. Yousef, A.A. and Iman, M. Talaat (2003): Physiological response of rosemary plants to some vitamins. Egypt. Pharm. J. 1, 81-93.
 7. Jackson, M.L. (1967). Soil Chemical Analysis. Printice Hall of India Private Limit. New Delhi.
 8. Saric, M.; Kastrort R.; Cupina T.; Cuplna T. and Geric L.(1967): Chlorophyll determination. Univ. noven Sodu Parktikum is Kizlolgize Biljaka, Beogard, Haucna, Anjiga, 215.
 9. Herbert, D.P.; Phlipps, P.and Strangle R. (1971): Determination of carbohydrates. Method in Microbial, 58, 209-344.
 10. Chapman, H.D. and Pratt P.F. (1961):Methods of Analysis for Soils, Plants and Waters. Univ. California Div. Agric. Sci. Berkely USA, 445.
 11. King, E.J. (1951): Microanalysis in Medical Biochemistry. 4th Ed J. and Ehar Chill. Ltd, London.
 12. Brown, J.D. and Lilland O. (1946): Rapid determination of Ca and Na in plant material and soil extract by flame photometer. Proc. Amer. Hort. Sci., 48:341-346.
 13. Snedecor, G.W. and Cochran W.G. (1980): Statistical Methods. 7th Ed Iowa State Univ., Press Iowa, USA.
 14. El-fawakhry, F.M. and El-Tayeb H.E. (2003): Effect of some amino acids and vitamins on chrnsynthemum production. J. Agric. Res. Alex. Univ., 8(4)755-766.
 15. Youssef, A.A., Mona, H. Mahgoub and Iman, M. Talaat (2004): Physiological and biochemical aspects of *Matthiola Incana* L. plants under the effect of putresine and kinetin treatments. Egypt. J. Appl. Sci., 19(9B) 492-510.
 16. Mona, H.Mahgoub and Iman,M. Talaat (2005):Physiological response of rose geranium (*Pelargonium graveoents* L.) to phenylalanine

- and nicotinic acid. *Annals of Agric. of Sci. Moshtohor*, 43(2):807-822.
17. Rawia, E.Aid; Lobna, S. Taha and Soad, M.M. Ibrahim (2010): Physiological properties studies on essential oil of *Jasminum grandiflorum* L. as affected by some vitamins. *Ozean J. of Appl. Sci.* 3(1): 87-96.
 18. Nahed G., Abd El-Aziz; Fatma E.M. El-Quesni and Farahat M.M. (2007): Response of vegetative growth and some chemical constituents of *Syngonlum podophyllum* L. to foliar application of thiamine, ascorbic acids and kinetin at Nubaria. *World J. Agric. Sci.*, 3(3): 301-305.
 19. Tarraf, S.A. ; El-Din K.G. and Balbaa L.K. (1999): The response of vegetative growth, essential oil of lemongrass (*Symbogom citrtus* Hort.) to foliar application of ascorbic acid, nicotinamid and some micronutrients. *Arab. Univ. J. of Agric. Sci.* 7, 1: 247-259.
 20. Kawasaki, T. (1991): *Modern chromatographic Analysis of Vitamins*, 2nd Ed., vol 60, New York, NY. Marcel Dekker, Inc. 319-354.
 12. Price, C.E. (1966): Ascorbic stimulation of RNA synthesis: *Nature*, 212-1481.
 13. Sakr, M.T., El-Hadidy, M. Abo Elkheer, A.M. and Farouk, S. (2004): Physiological studies of some osmoregulators on Canola. *International Conservation Microbiology and Biotechnology in Africa and Arab Region*, 27th to 29th pp 295-321.
 14. Nahed G. Abdel Aziz and Lobna S.Taha and Soad.M.M Ibrahim (2009): Some studies on the effect of putrescine, ascorbic acid and thiamine on growth, flowering and some chemical constituents of gladiolus plants at Nubaria. *Ozean J. of Appl. Sci.* 2(2): 169-179.
 15. Blokhina, O; Virolanen, E. and Fagersted, M. (2003): Antioxidant, oxidative damage and oxygen deprivation stress A. *Review Ann. Bot.*, 91:179-194.
 16. Foyer, M.J. (1992): The antioxidant effects of thylakoid vitamin E(-tocopherol). *Plant Cell and Environment*, 15, 381-392.
 17. Smirnoff, N. (1996): The function and metabolism of ascorbic acid in plants. *Ann. Bot.*, 78: 661-669.
 18. Sakr, M.T. ; Khatab E.A. and Afifi M.H. (2008): Role of plant antioxidants (ascorbic acid, glutathione, -tocopherol and spermine) in alleviating salinity stress on growth and yield of wheat plants. *Egypt. J. Agron.* Vol. 30, No (2):153-165.
 19. Fatma,El-Quesni, E.M.,Nahed G.Abd El-Aziz, and Magda,M. Kandil M.(2009): Some studies on the effect of ascorbic acid and -tocopherol on the growth and some chemical composition of *Hibiscus rosa sincses* L. *Ozean J. of Appl. Sci.* 2(2): 159-167.
 20. Conkting, P., (2001): Recent and vanes in the role and biosynthesis of ascorbic acid in plants. *Plant Cell Environment* 24, 283-304.
 21. Hassanein, R.A.M. (2003): Effect of some amino acids, trace elements and irradiation on fennel (*Foeniculum vulgare* L.). Ph.D. Thesis. Fac. Agric., Cairo Univ., Egypt.
 22. Abo-Dahab, T.A.M. and Nahed Abdel-Aziz G. (2006): Physiological effect of diphenylamine and tryptophan on the growth and chemical constituents of *philodendron erubescens*. *World J. of Agric. Sci.*, 2(1): 75-81.
 23. Anderson, J.W.; Foyer, L.H. and Walker D.A. (1983): Light dependant reduction of dehydroascorbate and uptake of exogenous ascorbate by spinach chloroplasts. *Plants* 198: 442-450.
 24. Ravtenkranz A.A.F.; Machler ,Li.; Martinoia, F.E. and Oertli, J.J. (1994): Transport of ascorbic and dehydroascorbic acids across protoplast and vacuole membrane isolated from barley (*Hordeum vulgare* L. cv. Gerbel) leaves. *Plant Physiology*. 106, 187-193.
 25. Foyer, C.H. (1993): Ascorbic Acid In: Alscher R.G. Hess, J.L. Eds. *Antioxidants in higher plants*, Boca Raton: CRC Pre7ss, 31-58.
 26. Asoda, K. (1994): Mechanisms for scavenging reactive molecules generated in chloroplasts under light stress In Baker NR, Boyer J.R., eds, *Photo inhibition of photosynthesis from molecular mechanisms to the field*. Oxford: Bios Scientific publishers, 129-142.
 27. Daveis, M.B. Austin J. and Partridge D.A. (1991): *Vitamin C: Its chemistry and biochemistry*. Cambridge: Royal Society of Chemistry.
 28. Asard H., Haremand N., and Caubergs R.J. (1995): Involvement of ascorbic acid and -type cytochrome in plant plasma membrane red ox reactions. *Protoplasm* 184: 36-41.
 29. Muhammad A.A.; Farrukh,N.;Farid S.and Shaziz,A.(2001):Effect of some chemicals on keeping quality and vase life of tuberose (*Polianthes tuberosa* l.) cut flowers. *J. of Res. Sci.*, Bahauddin Zakariya Univ., Multan, Pakistan, 12,No.1.pp.1-7.