

Dietary supplementation with polyunsaturated fatty acids source and its effects on the native turkey's liver important beneficial fatty acids

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Abstract: This experiment was conducted to evaluate canola oil on some of the beneficial fatty acids of Iranian native turkey liver. Nine male turkey chicks randomly divided into three experimental treatments (Three levels of canola oil; 0, 2.5 and 5 percent) with three replicates were arranged in a completely randomized design. Experimental diets consisted of: Basal diet with 0, 2.5 and 5 percent of canola oil. arachidonic acid, docosapentaenoic acid and α -linolenic acid significantly affected canola oil and this status shows that canola oil could improve liver fatty acids profile.

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1. Introduction

Essential fatty acids such as omega-3 fatty acids are molecules that cannot be synthesized by the human body but are vital for normal metabolism. Deposition of EPA and DHA in animal products such as meat and eggs provides a valuable source in the human diet. Feeding the long chain n-3 fatty acids EPA and DHA leads to their deposition in intramuscular fat and in eggs, particularly as phospholipids. There is some production by the animal of EPA and DHA from 18 carbon n-3 fatty acids (linolenic synthesis) which occurs in some vegetable oils such as canola seed or oil and linseed (flax). We know chicken meat due to relatively low prices could play an important role in human diets. The poultry has a peculiar capability of incorporating omega-3 fatty acids in the internal fat of meat and other tissues (Saricicek et al., 1997, Crespo and Esteve-Garcia, 2001, Zollitsch et al., 1997 and Osek et al., 2001). Due to increased knowledge of people, consumers have become health conscious and want to consume only healthy foods. Therefore in many countries manufacturers have started producing health-promoting foods. However, animal tissues enriched with n-3 fatty acids could effectively help human health (Kinsella et al., 1990). Canola oil is marketed as oil very low in saturated fatty acids and high in monounsaturated fatty acids, and one of good sources of omega-3 fatty acids (Mehta, 2000^{a,b}), and could directly usage in human foods or indirectly meets from enriched animal tissue such as meat or giblet. Poultry liver prices is low and poor people application that for food cooking. Iranian native turkey keeping in the

north of Azerbaijan village as range breeding, in this experiment deference level of canola oil usage in native turkey's diets and their effects on the liver fatty acids profile were studied.

2. Materials and methods

Nine male native turkey chickens were distributed in a completely randomized design (three level of canola oil 0, 2.5, 5 percent) with three experimental units each (ten chicks/pen). The experimental diets formulated isonitrogenous and isoenergetic, accordance with the 1994 recommendations of the National Research Council (table 1). The birds were given access to water and diets ad-libitum. The composition and calculated nutrient composition of the treatment diet is shown in Table 1. At the end of the growing period the number of two pieces from each pen randomly selected and slaughtered with cutting the neck vessels and experimental samples from each liver tissue samples prepared and sent to the laboratory at temperature -20°C below zero were stored and the composition of fatty acids present in the samples (Table 2) was determined by gas liquid chromatography, according to Folch et al (1957).

Statistical Analysis

The performance and analytical data obtained were analyzed by variance analysis using the procedure described by the SAS version 8.2. The Duncan mean separation test was used to determine significant differences between mean values.

3. Results

Least square means for EPA and DHA fatty acids of turkey liver shown in table 2. Eicosapentaenoic acid (EPA) or (5, 8, 11, 14, 17-eicosapentaenoic acid) not significant and experimental treatment numerically decreased compared control group. Docosahexaenoic acid (DHA) or (4, 7, 10, 13, 16, 19-docosahexaenoic acid) contents have a same rate and between different treatment have not significant deferent. C22: 4n-6, adrenic acid or (7,10,13,16-docosatetraenoic acid) Control group with 7.7425 percent and treatment with 2.5 percent canola oil with 8.0057 percent not significant, but experiment with percent canola oil with 9.0928 percent compared other group significantly is higher. C22:5 n-3, docosapentaenoic acid (DPA) or (7,10,13,16,19-docosapentaenoic acid) significantly affected levels of canola oil and from 3.3204 percent in control group respectively reached to 7.2176 and 8.0682 percent for 2,5 and percent of canola oil. C18:3 n-3, -linolenic acid or (9,12,15-

octadecatrienoic acid) in control group is 4.2234 percent ad usage canola oil in experimental treatment significantly increased content this fatty acid and respectively reached to 7.5523 and 7.9896 percent. DPA and DHA are chemically derived from linolenic acid; the conversion of linolenic acid into these products is not very efficient in mammals. The direct addition of these longer chain omega-3 fatty acids to the diet can significantly increase the amounts of EPA, DPA, and DHA in the tissues (Leskanich et al., 1997, Wood and Enser, 1997). Dietary fatty acids to modify tissue fatty acids profile (Azain, 2004). The fatty acid concentrations in meat, adipose and tissues reflect the animal's own metabolic processes as well as the diet it has consumed. Previous work has shown how the type of oil or fat consumed by animals can change the fatty acid concentrations in body tissues proportionately to the concentrations found in the diet (Skelley et al., 1975, Busboom et al., 1991, Romans et al., 1995, and Bee et al., 2002). This result suggests that the animals were able to process and incorporate the PUFAs into body tissues.

Table 2: Least square means for EPA and DHA fatty acids of turkey liver

| | Canola oil levels | | | P value | SEM |
|-------------------------------|---------------------|---------------------|---------------------|---------|--------|
| | control | 2.5 | 5 | | |
| C20:5n-3 ¹ | 2.842 ^a | 2.321 ^a | 2.500 ^a | 0.8729 | 0.7111 |
| C22:6 n-3 ² | 3.030 ^a | 2.637 ^a | 3.425 ^a | 0.4450 | 0.4089 |
| C22: 4n-6 ³ | 7.7425 ^b | 8.0057 ^b | 9.0928 ^a | 0.0290 | 0.2756 |
| C22:5 n-3 ⁴ | 3.3204 ^b | 7.2176 ^a | 8.0682 ^a | 0.0004 | 0.4136 |
| C18:3 n-3 ⁵ | 4.2234 ^b | 7.5523 ^a | 7.9896 ^a | 0.0008 | 0.3843 |

1- eicosapentaenoic acid (EPA) or (5,8,11,14,17-eicosapentaenoic acid) 2- docosahexaenoic acid (DHA) or (4,7,10,13,16,19-docosahexaenoic acid) 3- adrenic acid or (7,10,13,16-docosatetraenoic acid) 4- docosapentaenoic acid (DPA) or (7,10,13,16,19-docosapentaenoic acid) 5- -linolenic acid or (9,12,15-octadecatrienoic acid)

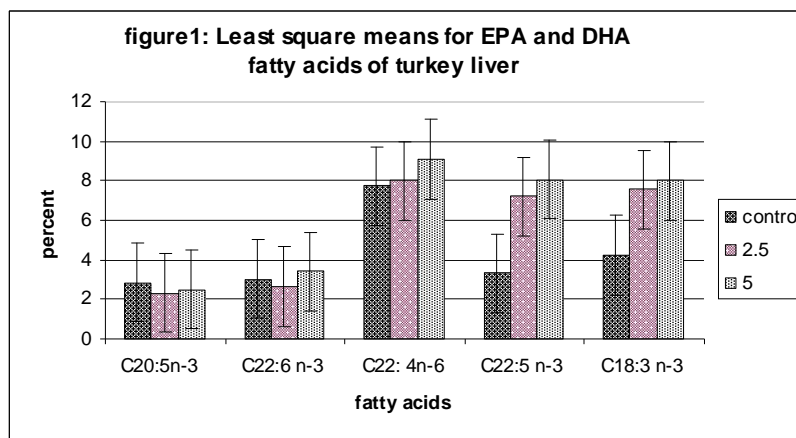


TABLE 1. Percentage composition of experimental diets in four period

| Ingredients' | 4 -8 week | | | 8 - 12 week | | | 12 - 16 week | | | 16 - 20 week | | |
|-----------------------------|-----------|--------|--------|-------------|--------|--------|--------------|--------|--------|--------------|--------|--------|
| | T1 | T2 | T3 | T1 | T2 | T3 | T1 | T2 | T3 | T1 | T2 | T3 |
| Corn | 42.50 | 38.00 | 36.00 | 45.60 | 43.00 | 35.00 | 56.64 | 48.50 | 40.00 | 64.41 | 58.00 | 48.00 |
| SBM | 34.40 | 36.00 | 31.15 | 28.25 | 27.30 | 28.24 | 26.00 | 27.00 | 27.50 | 21.00 | 21.00 | 21.00 |
| Oi | 0.00 | 1.25 | 2.50 | 0.00 | 2.50 | 5.00 | 0.00 | 2.50 | 5.00 | 0.00 | 2.50 | 5.00 |
| Fish | 4.80 | 3.70 | 6.60 | 8.00 | 8.00 | 8.00 | 2.64 | 1.82 | 1.50 | 0.65 | 0.70 | 0.67 |
| Starch | 3.10 | 3.22 | 1.56 | 7.46 | 3.32 | 3.37 | 6.57 | 6.51 | 6.50 | 7.10 | 5.56 | 6.71 |
| Alfalfa | 3.47 | 5.00 | 6.00 | 3.00 | 5.00 | 6.00 | 1.50 | 4.00 | 6.00 | 1.00 | 3.80 | 6.00 |
| DCP | 1.38 | 1.52 | 1.11 | 0.63 | 0.61 | 0.62 | 1.03 | 1.15 | 1.18 | 1.17 | 1.15 | 1.15 |
| Met | 1.50 | 1.50 | 1.50 | 1.50 | 1.50 | 1.50 | 1.50 | 1.50 | 1.50 | 1.50 | 1.50 | 1.50 |
| Lys | 1.50 | 1.50 | 1.50 | 1.50 | 1.50 | 1.50 | 1.40 | 1.50 | 1.50 | 1.50 | 1.50 | 1.50 |
| Oyster | 1.02 | 1.02 | 0.86 | 0.73 | 0.67 | 0.62 | 0.92 | 0.87 | 0.82 | 0.90 | 0.81 | 0.73 |
| wheat bran | 2.00 | 3.00 | 6.00 | 2.50 | 5.00 | 6.00 | 1.00 | 3.00 | 6.00 | 0.00 | 1.70 | 5.00 |
| Vit supp ¹ | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 |
| Min supp ² | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 |
| Salt | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 |
| Sand | 3.58 | 3.54 | 4.47 | 0.08 | 0.85 | 3.40 | 0.05 | 0.90 | 1.75 | 0.02 | 1.03 | 1.99 |
| | 100.00 | 100.00 | 100.00 | 100.00 | 100.00 | 100.00 | 100.00 | 100.00 | 100.00 | 100.00 | 100.00 | 100.00 |
| Calculated nutrient content | | | | | | | | | | | | |
| ME kcal/kg | 2755 | 2755 | 2755 | 2850 | 2850 | 2850 | 2945 | 2945 | 2945 | 3040 | 3040 | 3040 |
| Crude protein (%) | 24.7 | 24.7 | 24.7 | 20.9 | 20.9 | 20.9 | 18.1 | 18.2 | 18.1 | 15.7 | 15.7 | 15.7 |
| Calcium (%) | 0.95 | 0.95 | 0.95 | 0.81 | 0.81 | 0.81 | 0.71 | 0.71 | 0.71 | 0.62 | 0.62 | 0.62 |
| Available P (%) | 0.48 | 0.48 | 0.48 | 0.40 | 0.40 | 0.40 | 0.36 | 0.36 | 0.36 | 0.31 | 0.31 | 0.31 |
| ME/CP | 112 | 112 | 112 | 136 | 136 | 136 | 163 | 162 | 163 | 194 | 194 | 194 |
| Ca/P | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |

¹Vitamin content of diets provided per kilogram of diet: vitamin A,D, E and K.

² Composition of mineral premix provided as follows per kilogram of premix: Mn, 120,000mg; Zn, 80,000 mg; Fe, 90,000 mg; Cu, 15,000 mg; I, 1,600 mg; Se, 500 mg; Co, 600 mg

Conclusion

In conclusion, this study has demonstrated that the use of canola oil could significantly increased adrenic acid, docosapentaenoic acid and -linolenic acid content in the liver of native Iranian turkey and improved that quality.

Reference

- Saricicek BZ, Ocak N, Garipoglu AV. A study on utilizing fish oil in broiler diets. *Ziraat Fakultesi Dergisi, Poult. Sci* 1997; 12 : 33 – 42.
- Crespo N, Esteve-Garcia E. Dietary fatty acid profile modifies abdominal fat deposition in broiler chickens. *Poult. Sci* 2001; 80 : 71-78.
- Zollitsch W, Knaus W, Aichinger F, Lettner F. Effects of different dietary fat sources on performance and carcass characteristics of broilers. *Anim. Feed Sci. Tech* 1997; 66: 63-73.
- Osek M, Janocha A, Klocek B, Wasilowski Z. Influence of feed mixtures containing different fats on production coefficients and meat quality of slaughter chicken. *Poult. Sci* 2001; 22: 153-164.
- Kinsella JE, Lokesh B, Stone RA. Dietary n-3 polyunsaturated fatty acids and amelioration of cardiovascular disease: Possible mechanisms. *Am.J.Cli. Nutr* 1990; 52: 1-28.

Mehta BV, Mogal YM, Lawande VW. Rapeseed - Mustard oil: Fatty acid composition - India. In: *Sea Millennium Handbook on Indian Vegetable Oil Industry and Trade, The Solvent Extractors' Association of India*, 7th ed 2000a; pp. 962.

Mehta BV, Mogal YM, Lawande VW. Seed composition, fatty acid composition and characteristics of oil of a few varieties of linseed grown in India. In: *Sea Millennium Handbook on Indian Vegetable Oil Industry and Trade, The Solvent Extractors' Association of India*, 7th ed 2000b; pp. 987.

Folch J, Lees M, Sloane-Stanley GH. A simple method for the isolation and purification of total lipids from animal tissues, *J. Biol. Chem* 1957; 226: 497-509.

SAS Institute. SAS User's Guide: Statistics. SAS Institute Inc., Cary, NC, 1988.

Leskanich CO, Matthews KR, Warkup CC, Noble RC, Hazzledine M. The effect of dietary oil containing (n-3) fatty acids on the fatty acid, physicochemical, and organoleptic characteristics of pig meat and fat. *J. Anim. Sci* 1997; 75:673-683.

Wood JD, Enser M. Factors influencing fatty acids in meat and the role of antioxidants in

improving meat quality. *Br. J. Nutr* 1997; 78:s49-s60.

Azain MJ. Role of fatty acids in adipocyte growth and development. *J. Anim. Sci* 2004; 82:916-924.

Skelley GC, Borgman RF, Handlin DL, Acton JC, McConnell JC, Wardlaw FB, Evans EJ. Influence of diet on quality, fatty acids and acceptability of pork. *J. Anim Sci* 1975; 41:1298-1304.

Busboom JR, Rule DC, Colin D, Heald T, Mazhar A. Growth, carcass characteristics, and lipid composition of adipose tissue and muscle

of pigs fed canola. *J. Anim. Sci* 1991; 69:1101-1108.

Romans JR, Johnson RC, Wulf DM, Libal GW, Costello WJ. Effects of ground flaxseed in swine diets on pig performance and on physical and sensory characteristics and omega-3 fatty acid content of pork: I. Dietary level of flaxseed. *J Anim Sci* 1995; 73:1982-1986.

Bee G, Gebert S, Messikommer R. Effect of dietary energy supply and fat source on the fatty acid pattern of adipose and lean tissues and lipogenesis in the pig. *J. Anim. Sci* 2002; 80:1564-1574.

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