

**Characterization of *Escherichia coli* from diarrheic calves with special reference to plasmid profile**El-Shehedi, Mona<sup>1</sup>, A.; Mostafa, M. Eraqi<sup>2</sup> and Aisha, R. Ali<sup>1</sup>

1. Serology Unit, Animal Health Research Institute, Dokki, Giza, Egypt

2. Microbiology and Immunology Department, National Research Center, Dokki, Giza, Egypt  
[meraqrnc@gmail.com](mailto:meraqrnc@gmail.com)

**Abstract:** Out of 120 fecal samples collected from diarrheic calves at different localities in AL-Qalyoubia Governorate. 43 were positive for *Escherichia coli* with an incidence of 35.83%. Serogrouping of *E. coli* isolates recovered 8 belong to different "O" serogroups which were O<sub>55</sub>, O<sub>111</sub>, O<sub>26</sub>, O<sub>153</sub>, O<sub>8</sub>, O<sub>18</sub>, O<sub>86</sub>, O<sub>157</sub>, and "5" isolates were untypable. Antibiogram pattern of isolated *E. coli* serogroups showed that all tested serogroups of *E. coli* were sensitive to enrofloxacin, and neomycin, all groups were resistant to cefadroxil, colistin sulphate, Tobramicin and ampicillin. Meanwhile variable results were recorded with the remaining used chemotherapeutic agents. Plasmid profile analysis of various *E. coli* serogroups revealed that *E. coli* serogroups O<sub>111</sub> revealed 4 plasmids with their super coiled forms with a molecular weight ranging 1-7 Kbp, while *E. coli* serogroup O<sub>153</sub> revealed 3 plasmid with molecular weight 2.5-7 Kbp and O<sub>18</sub> harbored 2 plasmid with molecular weight 1.8-2.5 Kbp, while *E. coli* serogroups O<sub>26</sub>, O<sub>55</sub>, O<sub>8</sub> revealed 1 plasmid with the same molecular weight 7 Kbp. On the other hand *E. coli* serogroups O<sub>86</sub> have no plasmid.

[El-Shehedi Mona, A., Mostafa, M. Eraqi, and Aisha, R. Ali. **Characterization of *Escherichia coli* from diarrheic calves with special reference to plasmid profile.** *J Am Sci* 2013;9(7):54-59]. (ISSN: 1545-1003). <http://www.jofamericanscience.org>. 6

**Keywords:** diarrheic calves, plasmid profile, *Escherichia coli*

**1. Introduction**

*E. coli* is the most frequently encountered Gram negative organism causing diarrhea. Diarrhea represents an increasing and recurrent problem in young calves, from one week up to 12 weeks age, especially in suckling beef calves, **China et al., (1998)** and **William and Zimmer(2013)**. Certain pathogenic strains cause enteric diseases ranging in symptoms from Cholera-like diarrhea to severe dysentery, other *E. coli* may colonize the urinary tract, resulting in cystitis or pyelonephritis **Donneberg and Whittam (2001)**.

**Saridakis et al., (1997)** isolated 12 serotype of *E. coli* from diarrheic calves while O<sub>26</sub>: H<sub>11</sub>, O<sub>119</sub>: H<sub>25</sub> and O<sub>11</sub>: H, being the most prevalent. Also, **Fectrau et al., (2001)** isolated 25 *E. coli* strains from bacteremic calves for the presence of virulence factors. Isolates of pathotypes O<sub>78</sub>, O<sub>119</sub>, were positive but O nontypable were associated with a high mortality rate.

**Ratchtrachenchai et al., (2004)** determined 20629 *E. coli* strains isolated from children with acute diarrhea using 43 monovalent O antisera. The following serogroups were determined O<sub>6</sub>, O<sub>8</sub>, O<sub>15</sub>, O<sub>18</sub>, O<sub>25</sub>, O<sub>86</sub>, O<sub>119</sub>, O<sub>126</sub>, O<sub>127a</sub>, O<sub>128</sub>, O<sub>146</sub>, O<sub>159</sub> and O<sub>166</sub>. They recorded that were only four serogroups that were exclusively associated with a single pathotype: O<sub>20</sub>, O<sub>124</sub>, O<sub>52</sub> and O<sub>164</sub>.

The properties which allow pathogenic *E. coli* to invade infect and damage host cells are conferred by adhesions, toxins and hemolysins (**Beutin 1991**).

Diarrhoeagenic *E. coli* are recognized as five major pathotypes: enterotoxigenic (ETEC), enteropathogenic (EPEC), enteroinvasive (EIEC) Shiga-like toxin-producing (STEC) or enterohaemorrhagic (EHEC) and enteroaggregative (EAEC) *E. coli* **Nataro and Kaper (1998)**. **Griffin and Tauxe(1991)** mentioned that *E. coli* serotype O<sub>157</sub>: H<sub>7</sub> associated with human infection and death in United States, while (**Blanco et al., 2003**) reported that outbreak of disease have been traced to non-O<sub>157</sub>STEC serotypes. They include O<sub>5</sub>: NM, O<sub>6</sub>: H<sub>31</sub>, O<sub>26</sub>: H<sub>11</sub>, O<sub>26</sub>: NM, O<sub>48</sub>: H<sub>21</sub>, O<sub>91</sub>: NM, O<sub>111</sub>: NM, O<sub>113</sub>: H<sub>21</sub>, O<sub>128</sub>: H<sub>2</sub> and O<sub>128</sub>: NM. Two of these serotypes (O<sub>111</sub>: NM and O<sub>26</sub>: H<sub>11</sub>) can cause post weaning diarrhea in calves.

The emergence and dissemination of antimicrobial resistance *E. coli* has been well documented as a serious problem worldwide **Schroeder et al., (2002)**. Antimicrobial therapy is an important tool in reducing both the incidence and symptoms associated with diarrhea in cattle. However resistance to existing antimicrobials is widespread and of concern to veterinarians **Blanco et al., (1998)**. Also, the genes encoding these factors have been found on large conjugative plasmids and the occurrence of antimicrobial resistance genes on these same plasmids. It impossible that use of antimicrobial agent may select for persistence of *E. coli* containing such plasmids **Johnson et al., (2002)** and **O'Brien et al., (1992)** mentioned that

plasmids were the major vector in the dissemination of resistance genes through bacterial population.

The aim of this study was directed mainly to determine the presence and prevalence rate of enteric pathogen of public health importance as *E.coli* isolated from diarrheic calves. Also complete biochemical and serological identification was done and studying the effect of different chemotherapeutic agents on various serogroups of *E.coli* which had been isolated in the present work and determination of plasmid profile.

## 2. Material and Methods

### 2.1. Collection of Samples:

A total of 120 fresh faecal samples collected from diarrheic calves suffering from profuse watery diarrhea tinged with blood, fever and anorexia aged from 1 week to 2 months at different localities in AL-Qalyoubia Governorate.

### 2.2. Isolation and identification:

All fecal samples were inoculated directly into nutrient broth and incubated for 24 hours at 37°C, then sub cultured onto MacConkey agar and EMB agar the inoculated plates were incubated at 37°C for 24-48 hours. The suspected colonies were identified morphologically, culturally and biochemically according to **Koneman et al., (1996)**.

### 2.3. Serological identification:

Antisera of *E.coli* were used for serological identification of somatic "O" using slide agglutination test according to **(Edwards and Ewing 1972)**.

Antisera -O-sera were obtained from DENKA SEIKEN CO LTD Tokyo, Japan.

### 2.4. Antibio gram pattern:

Antibiogram was applied on the different isolated serogroup of *E.coli* using in vitro disc diffusion technique according to **Quinn et al., (2002)** and performed on Mueller Hinton agar plates and 11 Discs of chemotherapeutic agents were used (enrofloxacin, neomycin, amoxycillin, oxytetracyclin, chloramphenicol, streptomycin, cefadroxil, colistin sulphate, gentamicin, tobramycin, and ampicillin).

The results were interpreted according to **National Committee for Clinical Laboratory Standards (2000)**.

### 2.5. Plasmid analysis:

Plasmid DNA was extracted from cultured cells following alkaline lysis method of plasmid preparation according to **Vincent et al., (2010)** using QIAGEN plasmid MINI KITS.

The samples were processed using Gel electrophoresis to identify the number of plasmid copies present in different isolates. For this purpose, an Agarose Gel of 0.9% was used. Staining of DNA fragments was carried out using ethidium bromide and they were visualized U.V. Trans illumination. Standard DNA molecular weight marker were used to estimate of the plasmid size .The standard DNA molecular weight marker used in the present study were 1Kb ladder, 1 Kb plus DNA ladder and Lambda DNA 1 MIU 1 digest. The molecular weight of each plasmid was detected using Hind 111 Lambda phage digest. Plasmid analysis was performed in Biotechnology Center, Faculty of Veterinary Medicine, Cairo University.

## 3. Results

### 3.1. Bacteriological identification:

Out of 120 faecal samples were collected from diarrheic calves at different localities in AL-Qalyoubia Governorate. 43 samples were positive for *Escherichia coli* with an incidence of 35.83%.

### 3.2. Serological identification:

Serogrouping of (43) isolates of *E.coli* recovered from diarrheic calves revealed ( 8) different "O" serogroups and "5" strains were untypable as illustrated in Table(1).

**Table 1.** Certain characteristic of study sites

Serogroups		
	No	%*
O55	7	16.27
O111	6	13.95
O26	5	11.62
O153	5	11.62
O8	4	9.32
O18	4	9.32
O86	4	9.32
O157	3	6.97
Untyped	5	11.62
Total	43	100

\* the percent was calculated according to the No. of total isolates (43)

### 3.3 Antibio gram pattern:

Antibiogram pattern of pathogenic *E.coli* serogroups recovered from diarrheic calves were recorded in Table (2) which shows that all tested serogroups of *E.coli* were sensitive to enrofloxacin, and neomycin. On the other, *E.coli* were resistant to cefadroxil, colistin sulphate, tobramycin and ampicillin, while variable results were recorded with the remaining used chemotherapeutic agents.

**Table 2.** Antibiogram pattern of *E.coli* serogroups recovered from diarrheic calves

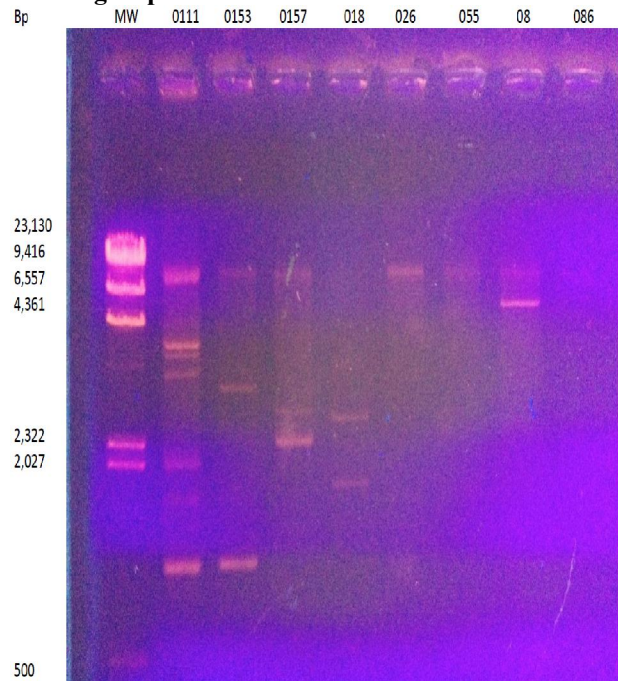
Chemotherapeutic agent	Conc. µg	O55	O111	O26	O153	O8	O18	O86	O157
Enrofloxacin	10	S	S	S	S	S	S	S	S
Neomycin	30	S	S	S	S	S	S	S	S
Amoxycillin	25	R	R	R	R	R	R	M	R
Oxytetracyclin	30	R	R	S	R	R	R	S	S
Chloramphenicol	30	R	R	R	R	R	R	M	R
Streptomycin	10	R	R	S	R	R	R	R	R
Cefadroxil	30	R	R	R	R	R	R	R	R
Colistinsulphate	50	R	R	R	R	R	R	R	R
Gentamicin	10	R	R	R	R	M	R	S	R
Tobramicin	10	R	R	R	R	R	R	R	R
Ampicillin	10	R	R	R	R	R	R	R	R

### 3.4. Plasmid analysis

**Table 3.** Plasmid profile analysis of different *E. coli* serogroups recovered from diarrheic calves

Strain	Plasmid and number		Molecular weight
O11	4 plasmids with their super coiled form		7 Kbp 3.5 Kbp 2 Kbp 1 Kbp
O153	3 plasmids		7 Kbp 3 Kbp 1 Kbp
O157	3 plasmids		7 Kbp 2.5Kbp 2.3Kbp
O18	2 plasmids		2.5 Kbp 1.8 Kbp
O26	1 plasmid		7 Kbp
O55	1 plasmid		7 Kbp
O8	1 plasmid with their super coiling		7 Kbp
O86	No plasmid		-

### Plasmid Profiling Analysis of different *E. coli* serogroups recovered from diarrheic calves:



### 4. Discussions

Pathogenic form of *E. coli* associated with human and animal diseases are remarkably diverse. Diseases that affect young domestic animals during the first few months of life causes great losses for animal industry. The immune system of animals at young age is not well developed and the maternal

immunity would not with stand variable infections **Mahmoud(1993)**. The most important bacterial enteropathogens are enterotoxigenic *E.coli* causing diarrhea in calves the disease is usually referred to as colibacillosis and include either enteric or systemic colibacillosis **Wary et al., (1993)**. Bacteriological examination of 120 faecal samples collected from diarrheic calves revealed that 43 isolates of *E.coli* in percentage of 35.83% .These finding nearly coincides with that of **Mahmoud (1993)** and **khan and khan (1997)** who isolate *E.coli* from diarrheic calves at different percentages. Also, **Vagh and Jani (2010)** isolated different serotypes of *E.coli* from the young cattle and buffalo calves affected with calf scours.

Serogrouping of Bacterial isolates of enteropathogenic *E.coli* collected from diarrheic calves revealed 8 different "O" serogroups and "5" strains were untyped. The most prevalent serogroups were O<sub>55</sub>, O<sub>111</sub>, O<sub>26</sub>, O<sub>153</sub>, O<sub>8</sub>, O<sub>18</sub>, O<sub>157</sub>. These results agree to certain extended those recorded with **Holland (1990)** who also mentioned that the ETEC isolated from calves was limited to a few serogroups especially O<sub>8</sub>, O<sub>9</sub> and O<sub>101</sub>, while **Donkersgoed (1999)** isolated *E.coli* O<sub>157</sub> from faecal samples collected from cattle in Canada, also **Champman et al., (1993)** isolated *E.coli* O<sub>157</sub> from bovine rectal swabs in an incidence of 15% in North America. They suspected that food of bovine origin considered as a vehicle of infection and is food born Zoonoses **Holland et al., (1999)** isolated *E.coli* serogroups O<sub>5</sub>, O<sub>26</sub>, O<sub>111</sub> from both apparently healthy and diarrheic calves, and recorded that *E.coli* strains O<sub>26</sub> are more prevalent in calves with diarrhea. **Thin et al., (2011)** isolated different serogroups of *E.coli* from diarrheic calves in Vietnam and the most prevalent serogroups were O<sub>15</sub>, O<sub>20</sub>, O<sub>103</sub> and O<sub>157</sub>

Antimicrobial therapy is an important tool in reducing both the incidence and symptoms associated with diarrhea in cattle, resistance has become an important problem worldwide. Bacterial resistance to antimicrobial agents has been emerging as rapidly disseminating among many nosocomial and community-acquired pathogens **Tenover (2001)**. The development of antibiotic resistance in *E.coli* has important clinical implications. The development of resistance to older agents such as Ampicillin and Trimethoprim-sulfamethoxazol, as well as the emerging problem of fluoroquinolone resistance may substantially limit our antibiotic choices **Karlowsky et al., (2002)**. In the present study, the results of testing the *E.coli* serogroups against different antimicrobial agents are illustrated in Table (2). All tested serogroups were found to be sensitive to enrofloxacin and Neomycin.

These results coincide with that obtained by **Basoglu et al., (1999)** who recorded that *E.coli*

isolated from diarrheic calves were sensitive to enrofloxacin and cholestyamine. On the contrary, all tested serogroups were resistant to cefadroxil, colistin sulphate, tobramycin and Ampicillin, also variable results were recorded with the remaining tested antibiotic. These finding in agreement with those reported by **Blanco et al., (1996)** who recorded that the highest percentage of antibiotics resistance recorded with Tetracycline, streptomycin, sulphadiazin, ampicillin, kanamycin, chloramphenicol, trimethoprim and cotrimoxazole in treatment of *E.coli* in diarrheic lambs in Spanish, also **Schroeder et al., (2002)** and **Guerra et al., (2003)** reported that emergence and dissemination of antibacterial resistance in *E.coli* O<sub>26</sub> may complicate treatment of certain urinary tract and enteric infection in human and animals. Meanwhile **Mora et al., (2005)** recorded that most strains of *E.coli* showed resistant to five or more antibacterial agent (MDR) multiple drug resistant. The wide spread uses of antibacterial agents promoted the increasing frequency of *E.coli* multidrug resistance isolates in bovine which facilitate the spread of resistant plasmid to other bacteria. In addition **Nyman et al., (2010)** suggest multiple drug resistant in *E.coli* from Swedish dairy calves.

Antimicrobial resistance plasmids have been increasingly associated with both Gram positive and Gram negative bacterial infections. This trend is accelerated by the fact that *E.coli* is a common enteric commensal of mammals and a common cause of human infections. As such *E.coli* strain are routinely exposed to a wide range of antimicrobial agents. *E.coli* also has a very wide natural distribution and a propensity to plasmid carriage **Sherley et al., (2003)** Resistance to various antibiotic is relatively common in clinical pathogens in Turkey and also common *E.coli* strains **Özden et al., (2003)** and it is frequently plasmid-mediated **Neu (1992)**. In this study, plasmids were screened to determine their antibiotic resistance profiles and focusing on the changes on molecular level, could provide valuable insights for its management, these results showed a more than one plasmids in one strains as shown in table (3).

It was observed that there was no correlation between plasmid occurrence and multiple antibiotics resistance for all isolates because with serogroup O<sub>86</sub> which have no plasmid and produce multiple drug resistance with many antibiotics, also some serogroups contain 4 or 3 or 2 plasmids. These results are congruent with a molecular weight ranging 1-7 Kbp to the results reported by **Celebi et al., (2007)** who evaluate the plasmid and antibiotic resistance patterns among 118 uropathogen *E.coli* strains from infected urinary system plasmids were detected

113 strains (97%). Some isolates harbored up to 10 plasmids ranging from 1-19 Kb in size. in the contrary (Vincent, *et al.*, 2010)

Also Nasreen *et al.* (2009) mentioned that some isolates of *E.coli* possess single sized plasmid while other had multiple plasmids with different size ranged from 2.3 Kbp to 26 Kbp, very high antibiotic resistance was detected from isolates possessing high molecular weight plasmids.

Meanwhile Vincent *et al.*, (2010) reported that (62.9%) of clinical isolates of *E- coli* O<sub>157</sub> which have Multidrug resistance harbored plasmids all of which were no less than 2.1 Kbp in size but only 35% contained multiple plasmids. Moreover Nasreen *et al.*, (2009) reported that the clinical isolates of *E- coli* gain resistance to numerous of antibiotics by various mechanisms. This drug resistance increases as function of time and their microorganisms exposure to many factors (antibiotics, chemicals) besides, the bacteria acquire resistance through different route, such as natural or intrinsic resistance (inaccessibility of the target, multidrug inactivation), mutational resistance (drug target site modification reduce permeability or uptake, metabolic by pass and derepression of multidrug efflux). All these mechanisms of antibiotic resistance warrant a detailed investigation of multiple factors, with prioritization of the studies of molecular characterization.

Such multidrug resistance has serious implications for the empiric therapy of infections caused by *E.coli* and for the possible co-selection of antimicrobial resistance mediated by multidrug plasmid. Thus, the study confirm the important role of plasmid numbers that controls the resistance characteristics in *E.coli*.

#### Acknowledgements:

Authors are grateful to the Animal Health Research Institute, National Research Center and Majmaah University for support to carry out this work.

#### Corresponding Author:

Dr. Mostafa, M. Eraqi.

**Permanent address:** Department of Microbiology and Immunology National Research Center, Egypt

**Current address:** Department of Medical Laboratories, College of Science, Majmaah University, KSA

E-mail: [meraqrnc@gmail.com](mailto:meraqrnc@gmail.com)

#### References:

1. Basoglu, A.; Sevinc, M.; Birdane, F.M. and Camkerten, I.(1999): A comparison of the antibiotic enrofloxacin and cholestyramine therapy in neonatal diarrheic calves. Israel, J. Vet Med., 54, 28-29.
2. Beutin, L.(1991): The different haemolysins of *E.coli*. Med. Microbiol. Immunol.(Berlin), 180: 167-182.
3. Blanco, M.; Blanco, J.E.; Mora, A. and Blanco, J. (1998): Distribution and characterization of necrotogenic *E.coli* CNF1+ and CN F 2+ isolated from healthy cows and calves. Vet. Microbiol., 59: 183-192.
4. Çelebi, A.; Duran, N.; Öztürk, F.; Açık, L.; Aslan, G. and Aslantas, Ö.(2007): Identification of clinic uropathogen *Escherichia coli* isolates by antibiotic susceptibility, plasmid and whole cell protein profiles. Advances in Molecular Biology(1): 31-40.
5. Champman, P.A.; Siddons, C.a.; Wright, D.j.; Norman, P.; Fox, j. and Crik, E. (1993): Cattle as a possible source of Verotoxin producing *Escherichia coli* O157 infection in man. Epidemiol. Infect. 111, 437-439.
6. China, B.; Pirson, V. and Mainil, J.(1998): Prevalence and Molecular typing of attaching and effacing *Escherichia coli* among calf population in Belgium. Vet. Microbiol., 63: 249-259.
7. Chowdhury, M. and Das, R.(2002): Evaluation of pathogenicity of *E.coli* strains isolated from diarrheic calves. J. of Intercardemia 6(special): 598-603.
8. Donkergoed, J.; VanGraham, T. and Gannon, V.(1999): The prevalence of Verotoxins, *Escherichia coli* O157: H7 and Salmonella in the faces and rumen of cattle processing. Canadian. vet. J. 40, 332-338.
9. Donnenberg, M.S. and Whittam, T.S.(2001): Pathogenesis and evaluation of virulence in enteropathogenic and enterohemorrhagic *E.coli* J. Clin. Invest., 107: 539-457.
10. Ewing, W.H.(1986): Edward's and Ewing's Identification of Enterobacteriaceae 4th Ed. Elsevier Science New York.
11. Fecteau, G.; Fairbrother, J.M. ; Higgins, R; Metre, D.V.; Pare, J; Smith, B.P. ; Homberg, C.A. and Jang, S.(2001): Virulence factors in *E.coli* isolated from blood of bacteremic neonatal calves. Vet. Microbiol., 78: 241-249.
12. Griffin, P.M. and Tauxe, R.V.(1991): The epidemiology of infection by *E.coli* O157: H7, other enterohemorrhagic *E.coli*, and the associated hemolytic uremic syndrome. Epidemiol. Rev. 13: 60-98.
13. Gross, R.J.; Ward, L.R.; Threlfall, E.J.; Cheasty, T. and Rowe, B.(1998): Drug resistance among *Escherichia coli* strains isolated from cerebrospinal fluid. J.Hyg. Camp. 90: 195-198.
14. Guerra, B.; Junker, E.; Schrorter, A.; Malorny, B.; Lehmann, S. and Helmuth, R.(2003): Phenotypic and genotypic characterization of antimicrobial resistance in German *Escherichia coli* isolated from cattle, swine and poultry. J. Antimicrob. Chemother., 52, 489-492.
15. Hermoso, M.; Hermoso, J. ; Alonso, M.P.; Dahbi, G.; Gonzalez, E.A.; Bernardez, M.I. and Blanco, J.(2003): Serotypes, virulence genes and intimin types of Shiga toxin (Virotoxin) - producing *E.coli* isolates from

- healthy sheep in Spain. *J.Clin. Microbiol.*, 41: 1351-1356.
16. Holland, R.E. (1990): Some infectious causes of diarrhea in young Farm Animals. *Clin. Microbiol. Rev.*, 3, 345-375.
  17. Holland, R.E.; Wilson, R.A.; Holland, M.S.; Yuzbasiyan, G.V.; Mullaney, T.P. and White, D.C. (1999): Characterization of *E.coli* isolated from healthy and diarrhoeic calves. *Vet. Microbiol.* 66, 251-263.
  18. Johnson, J.R.; Stell, A.L.; O'Bryan, T.T.; Kuskowski, M.; Nowiki, B., and Johnson, C.(2002) : Global molecular epidemiology of the O15: K52: H1 extraintestinal pathogenic *Escherichia coli* clonal group: evidence of distribution beyond Europe. *J. Clin. Microbiol.* 40, 1913-23.
  19. Karlowsky, J.A.; Kelly, L.J.; Thornberry, G.; Jones, M.E. and Sahn, D.F.(2002): Trends in antimicrobial resistance among urinary tract infection isolates of *Escherichia coli* from female out patients in the United States. *Antimicrob. Agents Chemother.* 46: 2540-2545.
  20. Khan, A., and Khan, MZ., (1997): A etiology of neonatal calf mortality *J. Islamic Acad., Sci.*, 159-165.
  21. Mahmoud, A.R.(1993): Clinical pathological studies on some diseases of cattle and buffalo calves. ph.D. Thesis (clinical pathology) Fac. Vet. Med. Cairo Univ.
  22. Nasreen, J.; Sudar, U. M. and Archana, K. (2009): Plasmid profile analysis of multidrug resistant *E.coli* isolated from UTI patients of Nagpur City, India. *Romanian Society of Biological Sciences*, Vol.14, No.5, 4635-4640.
  23. National Committee for Clinical Laboratory Standards (2000): Performance standards for antimicrobial disk susceptibility tests. Wayne, Pennsylvania: National Committee for Clinical Laboratory Standards . NCCLS Document, M2-A7.
  24. Nayman, A.; Greko, G.; Bengtsson, B. and De Vendier, K. (2012): Antimicrobial resistance and virulence factors in *E.coli* from Swedish dairy calves. *Acta. Vet. Scand.* 54.(1): 2.
  25. Neu, H.C.(1992): The crisis in antibiotic resistance. *Science.* 257: 1064-1072.
  26. O'Brien, A.D.; Tesh, V.L.; Donohue-Rolfe; Fackson, M.P.; Olsnes, S.; Saundvig, K; Lindberg, A.A. and Keusch, G, T. (1992): Shiga toxin: biochemistry genetics, mode of action and role in pathogenesis *Curr. Topics Microbio. Immunol* 180: 65-94.
  27. Özden, M.; Kalkan, A.; Demirdağ, K.; Kiliç, S, and Özdemirelli. (2003): Aciprofloxacin and cotrimoxazole resistance and extended spectrum beta-lactamsae production in *E.coli* strains isolated from urinary tract infection *Int. Antimicrob. Agents.* 21: 492-493.
  28. Quinn, P.J.; Carter, M.E.; Markey, B. k.; Donnelly, W.J.C. and Leonard, F.C. (2002): "Veterinary Microbiology and Microbial Disease." Great Britain by MPG, Book Ltd, Bodmin, Corn wall, U. K. Ratchtrachenchai, O.; Subpasu, S.; Hayashi, H. and Thein, W.(2004): Prevalence of childhood diarrhea associated *E.coli* in Thailand. *J. Med. Microbiol.*, 53: 237-243.
  29. Saridakis, H.O; EL-Gared, S.A.; Viodotto, M.C. and Guth, B.E.C.(1997): Virulence properties of *E.coli* strains belonging to enteropathogenic (EPEC) serogroups isolated from calves with diarrhea. *Vet., Microbiol.*, 54: 145-153.
  30. Schroder, C.M.; Zhao, C, ; Debroy, C.; Torcolini, J.; Zhae, S. and White, D.G.(2002): Antimicrobial resistance of *E.coli* O157 isolated from human, cattle, swine and food, *Appl. Environ. Microbiol.* 68: 576-581.
  31. Sherley, M.; Gordon, D.M. and Colligon, P.J.(2003): Species differences in plasmid carriage in the *Enterobacteriaceae* plasmid. 49: 79-85.
  32. Tan Duc, Nguyen, Thin Thanh, Vo., and Hung, Vu-Khac (2011): Virulence factors in *Escherichia coli* isolated from calves with diarrhea in Vietnam. *J. Vet Sci.* 2011 June; 12(2): 159-164.
  33. Tenovar, F.C.(2001): Development and spread of bacterial resistance to antimicrobial agents. An overview. *Clin. infect. Dis.* 33: 108-115.
  34. Thin, T.V.; Hung, V.K. and Tn, D.N.(2011): Virulence factors in *Escherichia coli* isolated from calves with diarrhea in Vietnam. *J. Vet. Sci.* 12: 159-164.
  35. Vagh, A.A. and Jani, R.G.(2010): Prevalence and comparative studies of some major serotype of *E.coli* from cattle and buffalo calf scour. *Vet. World*, 3: 458-459.
  36. Vincent, N.CH.; Veronca, G.U.; Stella, I.S.; Etinosa, O.I. and Anthony, I.O. (2010): Multidrug resistance and plasmid patterns of *Escherichia coli* O157 and other *E.coli* isolated from diarrhoeal stools and surface waters from some selected sources in Zaria, Nigeria. *Int. J. Environ. Res. Public Health.* 7, 3831: 3841.
  37. Wary, C. McLaren, I. and Carrall, P.J.(1993): *Escherichia coli* isolated from farm animals in England and Wales between 1986 and 1991. *Vet. Rec.* 13: 439-442.
  38. William, A. and Zimmer, D.V.M. (2013): Management of calves with clinical *Escherichia coli*. [www.Bio-Vet.com](http://www.Bio-Vet.com).

5/23/2013