

**Antifungal activity OF ethyl acetate extract of four species *Bacillus* isolated from soil**Najwa Mohammed Jameel Ali Abu-Mejdad<sup>1</sup>, Farhan Laythe Aaiz<sup>2</sup>, Outoor Talib Jassim<sup>3</sup>.Biology Department-College of Science-University of Basra<sup>1</sup>  
Basic Medical Department – College of Nursing – University of Basrah<sup>2,3</sup>

**ABSTRACT:** During study isolated and identified four species backed to genus of *Bacillus* from 30 soil samples, in Basrah province (Al-Hartha, Abu-Alkaseeb, Al-Jibasi and Safwan) by using dilution method. Crude metabolites extract from bacterial isolates prepared by ethyl acetate extract for bacterial isolates. The crude metabolite extracted from *Bacillus stearothermophilus* showed antifungal activity against both filamentous fungi and yeasts while extract from *B.licheniformis* Don't exhibited any antifungal activity against all tested.

[Najwa Mohammed Jameel Ali Abu-Mejdad, Farhan Laythe Aaiz, Outoor Talib Jassim. **Antifungal activity OF ethyl acetate extract of four species *Bacillus* isolated from soil.** *J Am Sci* 2013;9(10):172-176]. (ISSN: 1545-1003). <http://www.jofamericanscience.org>. 22

**Keywords:** *Bacillus*, antifungal activity

**I. Introduction**

Antibacterial substances are widely produced by a large number of microorganisms. Currently microorganisms of genera *Bacillus*, *Penicillium* and *Micromonospora* are known to produce more than 5000 different antibiotics [1]

Many species of *Bacillus* are capable of producing biologically active substances disintegrating fungal cell walls (e.g., polymyxines, bacitracin, butirosin, thuricins and cereins). *Bacillus*, widespread naturally as non-pathogenic bacteria, can produce many different antimicrobial substances. The metabolites from *Bacillus* showed the activity against some kinds of fungi [2,3,4]

The object of this study was to characterize *Bacillus* strains GB-017 and GB-0356, which produce antifungal substances. We also investigated the culture conditions required for the production of antifungal and the properties of the antifungal substance produced by *Bacillus* spp. isolated from soil of Korea. The researcher were found to be bacillus-shaped, gram-positive and motile, and to inhibit *Botrytis cineria*, *Fusarium* sp., *Pythium* sp., and *Rhizoctonia solani*. Antagonistic activity was maintained up to pH 9.0, and the antifungal activity was stable to heat at 80°C for 1 h. [5] During this study evaluated the potential antimicrobial activity of a bioactive compound produced by a *Bacillus* sp. isolated from soil sample in Woods Hole Massachusetts and the result showed high antifungal activity against test fungi [6]

Antagonistic properties of the strain *Bacillus subtilis* IB-54 with respect to dermatophytes fungi *Trichophyton rubrum*, *T. mentagrophytes* var. *gypseum*, and *Microsporum canis* was studied. The studied strains of *Bacillus* sp. effectively inhibited growth and development of dermatophytes when were

cultivated on the media containing different carbon sources. [7] (Lukmanova *et al.*, 2008).

Among different bacterial isolates, a potent *Bacillus subtilis* MTCC 8114 was isolated from garden soil samples which showed antifungal activity against test fungi i.e. *Microsporum flavum* and *Trichophyton* sp. [8]

Researcher to search for new and potent antimicrobial compound. Therefore the aim of our research is to the screening of potent antibiotic producing microorganism from the natural sources such as soil. The bacterial species such as *Bacillus stearothermophilus*, *B. polymyxa*, *B. subtilis* and *B. licheniformis* well studied for the production of antibiotic.

**II Materials and methods****Sample collection**

30 Soil samples from different places of the Basrah Basrah province (Al-Hartha, Abu-Alkaseeb, Al-Jibasi and Safwan) were collected in the sterile small reagent bottles from the surface and labeled properly and stored at 4°C until examination.

**Isolation of pure colonies**

Approximately 1 gram of soil sample was suspended in 9 ml sterile distilled water and vortexed vigorously to make uniform suspension. After that successive serial dilutions were made by transferring 1 ml of aliquots to 2nd test tube containing 9 ml of sterile water and in this way dilution up to 10<sup>-10</sup> were made. An aliquot of 0.1 ml of each dilution was taken and spread evenly over surface of nutrient agar plates and incubated overnight at 37°C.

After 24 hours plates were examined and few colonies nearly same in appearance showed [9]

**Identification of *Bacillus* isolates**

Isolate was characterized by morphological and biochemical analysis using the tests prescribed in [9].

- 1- Gram staining
- 2- Spore formatting
- 3- starch analysis(Amylase production)
- 4- Vogus proskauer test(VP)
- 5- Citrate
- 6- Growth at 55
- 7- Oxidase production
- 8- Catalase production
- 9- Motility

#### Extraction of crude antibiotics from four species of *Bacillus*

The primarily screened bioactive *Bacillus* isolates were inoculated into 100 ml glucose nutrient broth each only and incubated for 24 hours at 37°C. After incubation, 10 ml of culture broth was transferred to another 100 ml of sterile broth in 250 ml conical flask and incubated at 37°C for 3 days with shaking (120 rpm). Following 3 days incubation the broth culture was centrifuged at 10000 rpm at /5 mint. The culture supernatant was collected and mixed with equal volume of ethyl acetate solvent in a separating funnel and then shake gently for 2 hours. The organic solvent was collected and dried at room temperature. The solvent extraction was then assayed for antifungal activity by well agar diffusion method. The residue was termed as crude antimicrobial metabolite [1]

#### Test organisms

Eight fungal isolates were Isolated from clinical specimens in Al-Zubair hospital (Onychomycosis, Otomycosis, oral candidiasis) from periods 1-1-2013/1-4-2013 this fungal isolates identification depended on [10,11]

#### Antifungal assay

To test antifungal activities of the crude extract of isolates *Bacillus*, the well agar diffusion method[12]

Activated fungal isolates by re cultured on Sabourauds dextrose agar(SDA) (Himedia, India) incubated at 27 C for week to filamentous fungi and 3 days for yeasts. Spreading fungal isolates suspension at concentration  $6 \times 10^8$  by using MacFrland scale. The crude extract was dissolved in respective solvent to get a solution of known\_concentration (100mg/ml). make (5 mm wells) were then Added 100  $\mu$ l from the crude extract of four species to *Bacillus* were placed on SDA medium evenly seeded with the test fungi. and blank well(fill with solvent) were used as a negative control respectively. The plates were then kept in incubators at 27°C for about 3 days for yeasts and 7 days for filamentous fungi.Finally the plates were incubated at this temperature to allow maximum growth of the fungi. The test materials having antifungal activity inhibited the growth of the fungi and a clear, distinct zone of inhibition was visualized surrounding the well on the medium. The antifungal

activity of the test agent was determined by measuring the diameter of the zone of inhibition in terms of millimeter (mm) with a transparent scale and the experiment was carried out in triplicate.

## II-Results and discussion

Screening for new antibiotics from natural sources is becoming increasingly important for the pharmaceutical industry as pathogenic fungi are increasingly becoming resistant to commonly used therapeutic agents[13]

Scientists are now working to explore alternative drugs from microbial sources to discover new drug as antifungal. Secondary metabolites from microorganisms having a diverse chemical structure and biological activities are produced only by some species of a genus *Bacillus* [14]

During this study The four species of *Bacillus* were identified on basis of morphological and biochemical characteristic (Table 1).

Also this study was performed with an aim of screening and isolation of antimicrobial substances producing bacteria from soil. Four *Bacillus* species were isolated from soil samples as they produced thin clear zone around the colony during well agar diffusion technique and purified by subculture. But one of them *Bacillus stearothermophilus* showed best activity against pathogenic fungi while *B. licheniformis* don't showed any activity against all tested fungi (Table 2, Plate (1,2)) 0. The results showed activity it as antifungal because these substances (crude metabolites) possess an active proteinaceous nature responsible for the activity[4]

The isolated species exhibited inhibitory effect on the growth of different test fungi, the crude secondary metabolites at a dose of 100 mg/ml of *Bacillus stearothermophilus* obtained from ethyl acetate extraction showed strong antifungal activity against Filmentouse fungi and yeast with the zone of inhibition range 6 to 27 mm (Table 2, Plate 1,2).

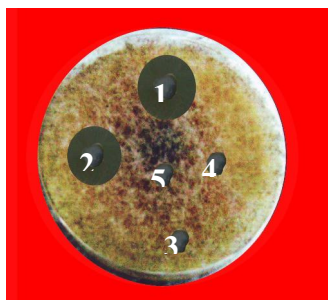
The highest zone of inhibition was 27 found against *Chrysosporium tropicum* followed by *B. polymyxa* may be this activity because this tow isolates can production Bacitracine, polymyxin, colistin, gatavalin, jolipeptid which considered peptide antibiotics cyclic there may be make unusual linkage with component of cell wall for fungi and prevent formation cell wall [15], while *B. subtilis* The highest zone of inhibition was 15 agains *Rhodotorula rubra*. In the case of *B. subtilis* some of these antibiotics have been found to be produced by a great variety of strains, e.g. subtilosin A, sublancin. Others are expressed strain-specifically, for example, ericin is produced by *B. subtilis* A1/3 only. Or may because surface of yeast smaller than filamentous fungi

therefore yeast sensitive while filamentous resistance [16].

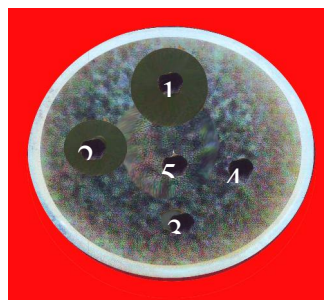
Either *B. licheniformis* didn't showed any activity against test fungal isolates may causes backed to referred [17] he reported that when glucose was added to culture media which seeded *B. licheniformis* results showed inhibited production of bacitracin synthesis which considered antifungal substance during first few hour of growth. he initially concluded that this delay in antibiotic production so may this causes to uncapability to inhibit test fungi.

### Conclusion

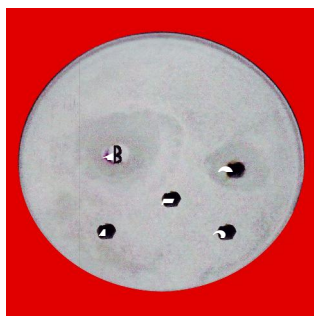
The three bacterial isolates from soil were *Bacillus stearothermophilus*, *B. polymyxa*, and *B. subtilis* possessing appeared antifungal activity. From the above studies we had seen that the crude metabolites extract of these isolates were active against both pathogenic filamentous fungi and yeast therefore This study possible to be encouraging to treat fungal infections and it marks the beginning of extraction and purification of antibiotics produced by a species listed.



*Rhizopus*



*Alternari*



*Chrysosporiu*



*Chaetomiu*

Plate(1):antifungal activity of four species Bacillus against filamentous fungi

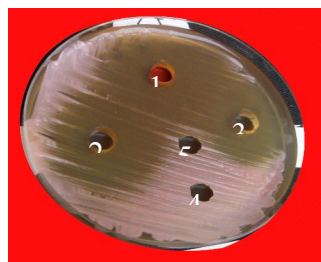
1- *Bacillus stearothermophilus*

2- *B. polymyxa*

3-*B. subtilis*

4-*B. licheniformis*

5-Control

*Rhodotorula rubra**Candida tropicalis**Cryptococcus neoformance**Candida albicans*

Plate(2):antifungal activity of four species Bacillus against yeasts

1- *Bacillus stearothermophilus*

2- *B. polymyxa*

3-*B.subtilis*

4-*B.licheniformis*

5-Control

Table(1): Morphological and Biochemical characteristics of Bacillus

Test Species	Gram staining	Spore formation	amylase	VP	citrate	Growth at 55°C	oxidase	catalase	Motility
<i>Bacillus stearothermophilus</i>	+	+	+	-	-	-	+	+	+
<i>B. polymyxa</i>	+	+	+	+	-	-	+	+	+
<i>B. subtilis</i>	+	+	+	+	+	-	+	+	+
<i>B. licheniformis</i>	+	+	+	+	+	+	+	+	+

Table (2) Antifungal activity of crude metabolites of four species Bacillus Diameter of inhibition zone (in mm)

Series	Fungal isolates	Inhibition zone (mm)				
		<i>Bacillus stearothermophilus</i>	<i>B. polymyxa</i>	<i>B. subtilis</i>	<i>B. licheniformis</i>	control
1	<i>Rhodoturla rubra</i>	25	19	15	0	0
2	<i>Candida albicans</i>	13	12	12	0	0
3	<i>Candida tropicalis</i>	15	6	0	0	0
4	<i>Cryptococcus neoformans</i>	6	0	0	0	0
5	<i>Rhizopus stolonifer</i>	12	10	0	0	0
6	<i>Chrysosporiumtropicum</i>	27	20	0	0	0
7	<i>Alternaria alternate</i>	15	13	0	0	0
8	<i>Chaetomium globosum</i>	20	15	6	0	0

## References

[1] Dev Sharma, S.C; Shovon, M, S, Sarowar Jahan M. G.; Asaduzzaman, A.K.M.; Ajijur Rahman, M.D.; Biswas, K.K.; Roy, N.A.(2013) A ntibacteriak and cytotoxic activity of Bacillus

methylotherophilus -SCS2012 isolated from soil. JMBFS. 2 (4) 2293-2307 .

[2] Awais, M; Perverz, A.; Yaqub, A. and Shah, M.M. (2010). Production of antimicrobial metabolites by *Bacillus subtilis* immobilized in

- polyacrylamide Gel. Pakistan. J.Zool.42(3):267-275.
- [3] Wan,H.K.; Xiao, R.F. and Qi, W.(2012). Antifungal activity of *Bacillus coagulans* TQ 33, isolated from skimmed milk powder against *Botrytis cinerea*. original scientific paper.1-17.
- [4] Oliveira, V.F.; Abreu, Y.J.L.; Fleming, L.R. and Nascimento, J.S.(2012). Anti-Staphylococcal and antifungal substances produced by Endospore-Forming Bacilli. J. of Appl. Pharm. Sci. 2(4):154-157.
- [5] Kim, H.S.; Park, J.; Choi, S.; Choi,K.; Lee, G.P.; Ban, S.J.; Lee, C.H. and Kim, C.S.(2003). Isolation and characterization of *Bacillus* strain for biological control. J. of Microbiol.3(41): 196-201.
- [6] Okulate, M.A. (2009). Antimicrobial activity of bioactive compounds produced by *Bacillus* species. final report for microbial diversity course.1-14.
- [7] Lukmanova K. A., Gizatullina S.V., Magazov R.Sh., Galimzianova N.F. & Aktuganov G.E. (2008). Antagonistic activity of bacteria from *Bacillus* genus against dermatophyte fungi. Zh Mikrobiol Epidemiol Immunobiol 4: 21-
- [8] Kumar A., Saini P. & Shrivastava J. N. (2009). Production of peptide antifungal antibiotic and biocontrol activity of *Bacillus subtilis*. Indian Journal of Experimental Biology 47: 57-62.
- [9] Cowan, S. T. and Steel, K.J. (1975). Manual for the Identification of Medical Bacteria. 2<sup>nd</sup> ed. Cambridge University Press. Cambridge, London. 238 pp.
- [10] Ellis, D.H. (1994).Clinical mycology: The human opportunistic mycosis. Gillingham. Printers Pty. Ltd. Australia. 166p.
- [11] Hoog de, G.S. & Guarro, J. (1995). Atlas of clinical fungi. center albureau Voor Schimmelcultures and universitat Rovirai Virgili. Spainb. 720p.
- [12] Collee, J.; Fraser, A.; Marmion, B. & Simon, A. (1996). Makie and McCartney practical medical microbiology. 14<sup>th</sup> ed. Churchill Liverstone. New York.978p.
- [13] Khokhar, N.; Mullas, S.; Shah,L. and Vaghela, G.(2013). Characterization of clinical isolates like bacteria and fungi from ocular infection. J. infection Disease Letters.1(2):12-15
- [14] Cho MK, Math RK, Hong SY, Islam S Md A, Mandanna DK, Cho JJ, Yun MG, Kim JM, Yun HD (2009). Iturin produced by *Bacillus pumilus* HY1 from Korean soybean sauce (Kanjang) inhibits growth of aflatoxin producing fungi. Food Control, 20: 402–406.
- [15] Awias. M; Shah, A. A and Hameed, F. (2007) Isolation, identification and optimization of bacitracin produced by *Bacillus sp.* park, 39 (4):1303-1312
- [16] Stein T, Borchert S, Conrad B, Feesche J, Hofemeister B, Hofemeister J, Entian KD (2002) Two different antibiotic-likepeptides originate from the ericin gene cluster of *Bacillus subtilis* A1/3. *J Bacteriol* 184: 1703-1711.
- [17] Haavik, H.I. (1974). Studies on the fermentation of bacitracin by *Bacillus licheniformis*: effect of glucose. J. Gen. Micro.

9/10/2013