

Surface, Biological and Antitumor Activity of some thio- based cationic surfactants

Sharbat A. Bakr

Chemistry Department, Faculty of Science, Al-Azhar University, Nasr City, Cairo, Egypt, B.O. Box, 11754.
dr.sharbatbakr@yahoo.com

Abstract: Three thio-based cationic surfactants were synthesized depend on reaction of anthranilic and sulphanic acids with fatty isothiocyanate having hydrocarbon chain length of C₁₂, C₁₆ and C₁₈ followed by quaternization with ethyl iodide. Elemental analysis, FTIR spectra, mass spectra and ¹H NMR spectra were performed to insure the structure of the prepared compounds. Their anticancer activity was tested by using Ehrlich Ascites Carcinoma (EAC) as a model system of mice cell tumor. The compounds were also tested in vitro on three human tumor cell lines: MCF 7 (breast carcinoma), HEPG 2 (liver carcinoma) and HCT 116 (colon carcinoma), antimicrobial activity against different species of bacteria and fungi using agar well diffusion methods. The surface properties of these surfactants were investigated. The surface properties studied included Critical Micelle Concentration (CMC), maximum surface excess (Γ_{max}), and minimum surface area (A_{min}). Free energy of micellization (ΔG_{mic}) and adsorption (ΔG_{ads}) were calculated.

[Sharbat A. Bakr. **Surface, Biological and Antitumor Activity of some thio- based cationic surfactants.** *J Am Sci* 2017;13(2):106-120]. ISSN 1545-1003 (print); ISSN 2375-7264 (online). <http://www.jofamericanscience.org>. 14. doi:[10.7537/marsjas130217.14](https://doi.org/10.7537/marsjas130217.14).

Key words: Surface activity, Biological activity, Antitumor activity, thio-based cationic surfactants.

1- Introduction

Malignancy is a diverse class of diseases in which a group of cells display uncontrolled growth (division beyond the normal limits), invasion (intrusion on/and destruction of adjacent tissues), and sometimes metastasis (spread to other locations in the body via lymph or blood).

Cancers are caused by abnormalities in the genetic material of the transformed cells [1]. These abnormalities may be due to the effects of carcinogens, such as tobacco smoke, radiation, chemicals, or infectious agents. Other cancer-promoting genetic abnormalities may randomly occur through errors in DNA replication, or are inherited, and thus present in all cells from birth. The heritability of cancers is usually affected by complex interactions between carcinogens and the host's genome. In vivo studies have revealed that some of anionic, cationic, non-ionic, and zwitter surfactants have antitumor activity [2] and inhibit the development of tumor metastases [3,4]. The action of one halogenic quaternary ammonium compound on the in vitro proliferation of different lines of human cancer cells, indicate that halogenic quaternary ammonium present a potent growth inhibitory activity of different cancer cells lines. The presence of quaternary ammonium group, responsible for some alkylating effect [5] Cationic surfactants (CSs) have been used as antibacterial [6-8] and antifungal [9-10] agents as well as denaturants of protein molecules [11,12] in biomedical and biochemical studies [13-15] and in the design of new functional materials [16,17], drug delivery systems [18-21], and as corrosion inhibitors [22].

Quaternary ammonium salt compounds show germicidal, antibacterial, antifungal and anticancer activities. A series of quaternary ammonium salts with general formula R₃N⁺X⁻, which include hetero nitrogen atom was the hetero atom, enhance the biological action [23].

Among the various classes of surfactants the cationic and more especially the quaternary ammonium salts are the most effective antimicrobial agent which kill or inhibit the growth of both Gram-positive and Gram-negative bacteria. In clinical and industrial environments quaternary ammonium compounds are widely used for controlling bacterial growth. The mode of antimicrobial action of quaternary ammonium compounds involves distortion of outer membrane lipid bilayers through association of the positively charged quaternary ammonium with the polar head groups of acidic phospholipids. The hydrophobic tail part interacts with the hydrophobic membrane core [24]. The studied surface activity relationship showed that in addition to the positively charged site, a significant hydrophilic component of the surfactant is used for controlling the biological activity of the cationic surfactant [25].

Several works deal with the synthesis of different cationic surfactant compounds and studied the relationship between surface activity and antimicrobial activity against wide strain of pathogenic bacteria, fungi and yeast [26-28]. Some quaternary ammonium compounds with different counterions, such as chloride or sulphate, have been used widely in clinical, food production health care, and domestic environments as the antiseptics preservatives to

eliminate bacterial infections and contaminations. However, since the irritant and cytotoxic effects of these compounds on human cells and tissues, such as keratinocytes, fibroblasts, cornea, and respiratory mucosa, have previously been shown,^[29-32] improvement of Quats is necessary not only for antimicrobial activity but also as far as safety with human cells.

The aim purpose of this work is to report on a relatively simple and practical synthesis of new environmentally friendly thio-based cationic surfactants derived from condensation of anthranilic acid and sulphanic acids with a mixture of fatty acid chlorides and ammonium thiocyanate (structure and abbreviations shown in Schem 1), as well as to evaluate their Surface, biological and antitumor activities.

II - Materials & Methods

Fatty acids (lauric, palmitic and stearic), thionyl chloride, ammonium thiocyanate, anthranilic acid, sulphanic acids and ethyl iodide were obtained from Merck and were purified and acetone and diethyl ether were all obtained from Merck and used without further purification.

FTIR spectra were recorded on a Perkin Elmer spectrometer in 4,000- 400 cm⁻¹ range using KBr discs. ¹H NMR spectra were recorded on a Bruker model DRX-300 NMR spectrometer with CDCl₃ as solvent and mass spectra were recorded on a Joel JMS-AX 500 (EI and FAB). Elemental analyses (C,H,O,N,I,S) of the synthesized materials were performed using a Varian Elemental instrument and are in satisfactory agreement with the calculated values.

II-1 -Synthesis of Fatty Acid Chlorides

Fatty Acid Chlorides are prepared according to the method^[33].

In a four necked round bottom flask equipped with stirrer and thermo pocket charge 0.1mole of fatty Acid at room temperature and heat to 40°C., then add 0.15mole of Thionyl Chloride under Nitrogen atmosphere over a period of 1 hour and heat the reaction mixture to reflux for 1 ½ hour to obtain crude Acid Chloride.

II-2 - Synthesis of 2 -Carboxy-N- (alkanoyl (Ethyl)Ammonio) Carbonothioyl)-N-Ethylbenzenaminium Iodide and 4 -Sulphonate -N-(Alkanoyl (Ethyl)Ammonio) Carbonothioyl) - Ethyl benzenaminium Iodide (IIa-c and IIIa-c).

Ammonium thiocyanate (0.0063 mol) was dissolved in 50 mL dry acetone and introduced into a

250-mL round-bottom flask after adding 1.5 mL fatty acid Chloride according to^[34]. After heating the solution at 90°C for about 60 min, (0.01mol) anthranilic acid and sulphanic acid were added and the solution was continuously stirred for about 8–10 h. After stirring, ethyl Iodide was added to the solution and placed aside on reflux for about 6 h. Finally, the product collected on ice and washed with doubly distilled water for the removal of impurities. The crude product was crystallized by diethyl ether. The pure products (IIa-c), (IIIa-c) were prepared and their physicochemical properties investigated as shown in table (1) such as molecular formula, molecular weight, colour, melting point, yield % and elemental analysis (Table 1), (Table 6). The Structure of the prepared compounds were analyzed by IR, ¹HNMR, Mass Spectra, The reactions were illustrate in scheme 1:

Step (1)



Step (2)

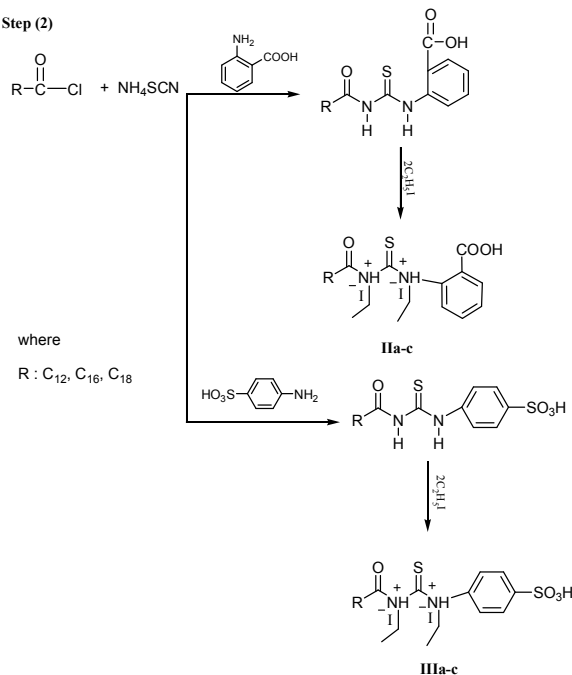


Table 1: *Physicochemical properties of 2 -Carboxy-N-(Dodecanoyl (Ethyl)Ammonio)Carbonothioyl)- N-Ethylbenzenaminium Iodide*

R	R	Abbreviations	Molecular formula	Mol.Wt	Colour	M.P	Yield %	Elemental analysis					
								C %	H %	N %	S %	I %	O %
Dodecyl	Dodecyl	2a	C ₂₅ H ₄₂ N ₂ S O ₃ I ₂	704.11	Yellowish brown	285-290	85	42.60 42.63	6.01 6.03	3.98 3.10	4.54 4.56	36.07 36.09	6.81 6.83
Hexadecyl	Hexadecyl	2b	C ₂₉ H ₅₀ N ₂ S O ₃ I ₂	760.17	Yellowish red	300-305	87	45.78 45.80	6.63 6.66	3.68 3.70	4.20 4.22	33.39 33.42	6.31 6.33
Octadecyl	Octadecyl	2c	C ₃₁ H ₅₄ N ₂ S O ₃ I ₂	788.20	Brown	300-305	90	47.20 47.23	6.90 6.94	3.55 3.57	4.06 4.08	32.19 32.22	6.09 6.12

II-3 Biological Activity

The synthesized cationic amphiphiles were screened for their antimicrobial activity against bacteria and fungi using agar well diffusion methods [35]. Bacteria species used in this study were *Pseudomonas aeruginosa* (RCMB 010043), *Escherichia coli* (RCMB 010052) as gram negative bacteria; *Streptococcus pneumoniae* (RCMB 010010), *Bacillus subtilis* (RCMB 010067) as gram positive bacteria and *Aspergillus fumigatus* (RCMB 02568), *Syncephalastrum racemosum* (RCMB 05922), *Geotrichum candidum* (RCMB 05097) and *Candida albicans* (RCMB 05036) as fungi.

II- 4 Antitumor or Activity Ehrlich ascites carcinoma (EAC)

The antitumor activity for the compounds under investigation was carried out at pharmacology unit, National Center Institute, Cairo, University.

The choice of Ehrlich ascites carcinoma (EAC) as a model system of mice cell tumor and it is a suitable tool for studying the biological behavior of malignant tumor and drug action within cells. A set of sterile test tubes used, where 2.5 × 10⁵ tumor cells per ml were suspended in phosphate buffer saline. Then 25, 50, 100 µg/ml from drug were added to the suspension, kept at 37°C for 2 hrs. Typan blue exclusion test was then carried out to calculate the percentage of nonviable cells [36].

Total number of NVC of cell

%NVC = number of NVC / total number of cell × 100 (7)

II-5 Measurements of human potential cytotoxicity by SRB assay

Human Potential cytotoxicity of the compound was tested using the method [37]

The cells were plated in 96-multiwell plate (104 cells/well) for 24 hrs before treatment of cells to the wall of the plate. Different concentrations of the compound under test (0, 5, 12.5, 25, 50 µg/ml) were added to the cell monolayer. Triplicate wells were prepared for each individual dose. Monolayer cells were incubated with the compound to be tested for 48 hrs at 37°C and in an atmosphere of 5% CO₂. After 48 hrs, the cells were fixed, washed and stained with Sulfo-Rhodamine -B stain. Excess stain was washed with acetic acid and attached stain was recovered with Tris EDTA buffer. The color intensity is measured in an ELISA reader. The relation between surviving fraction and drug concentration is plotted to get the

survival curve of each tumor cell line for the specified compound.

II-6 Surface Tension Measurements:

Surface tension measurements (ST) were performed for freshly prepared solutions of surfactants in concentration ranges 10⁻¹ to 10⁻⁶ mol / l using Du-Nouy tensiometer at 25°C. These values which remained constant for a period of 30 min. were taken as the surface tension of the solution. The results were accurate within ±0.1 mNm⁻¹. The critical micelle concentration (cmc) and the surface tension at the cmc were determined as the values of the break point of surface tension vs. concentration. Also the hydrophilic substituent's have a considerable role in their cmc values [38-39]. Plots of the free energies of adsorption and micellization vs. carbon numbers were illustrated. The maximum surface excess concentration air water interface (Γ_{max}) The packing densities of surfactants at the air – water interface are very important to interpret the surface activity of the surfactants [40] and can be calculated by the following Gibbs adsorption isotherm equation (1) [41, 42].

$$\Gamma_{\max} = \frac{1}{2.303 R n T} \left(\frac{\delta \gamma}{\delta \log C} \right) T \quad (1)$$

where R = gas constant (8.314 Jmol⁻¹ K⁻¹), T is absolute temperature (298 K), (δγ/δ log C) is the slope of the γ vs Log C plot at 25°C, The surface area occupied by surfactant molecules at the air/ water interface (A_{min}) is calculated using the equation (2):

$$A_{\min} = \frac{10^{14}}{\Gamma_{\max} \times N_A} \quad (2)$$

Where Γ_{max} and N_A are the maximum surface excess and Avogadro's number respectively.

Effectiveness of the synthesised cationic surfactants is determined from the difference between the surface tension of the surfactant solution at the critical micelle concentration (γ_{CMC}) and the surface tension of the distilled water (γ_o).

[43-44] as shown in eqn. (3). Larger π_{CMC} value of aqueous surfactants solution indicates its higher surface activity than aqueous surfactant solution with smaller π_{CMC} value [45].

$$\pi_{\text{CMC}} = \gamma_o - \gamma_{\text{CMC}} \quad (3)$$

Adsorption efficiency, (PC₂₀) of the surfactant concentrations that are capable of suppressing the surface tension of the solution by 20 mN/m and is obtained by using eqn. (4)

$$P_{C_{20}} = -\log C_{20} \quad (4)$$

C₂₀ is the minimum concentration which lead to saturation of the surface adsorption [46]. The surfactant with larger cmc/C₂₀ ratios has greater tendency to adsorb at the interface than to form micelle in the solution [47-48].

The free energies of micellization (ΔG_{mic}) and adsorption (ΔG_{ads}) were calculated using the thermodynamic Eqs. (5,6) [49,50,51] as follow:

$$\Delta G_{mic} = 2.303 RT \log (CMC) \quad (5)$$

$$\Delta G_{ads} = \Delta G_{mic} - (0.006 \pi_{CMC} \times A_{min}) \quad (6)$$

III. Results and discussion:

This work is report one simple practical synthesis of new environmentally friendly thio-based cationic surfactants derived from condensation of anthranilic and sulphanic acids with a mixture of fatty acids chlorides and ammonium thiocyanate) to produce compounds IIa-c & IIIa-c as illustrated in schem 1.

Table 2. Critical micelle Concentrations (CMC), Surface tension at cmc (γ_{cmc}), surface max (τ_{max}), Minimum Area/Molecule (A^2), Standard Free Energy of Micellization (ΔG_{mic}), Standard Free Energy of Micellization per Methylene group ($\Delta G_{mic}/CH_2$), Standard Free Energy Of adsorption (ΔG_{ads}) and Standard Free Energy of adsorption per methylene group ($\Delta G_{ads}/CH_2$) of 2 -Carboxy-N-(Dodecanoyl (Ethyl)Ammonio)Carbonothioyl)- N-Ethylbenzenaminium Iodide at Liquid Air Interface at 25C°.

R	Abbreviation	cmc mol/l	γ_{cmc} mN/m	τ_{max} mole/cm ²	Area mole A ² ×10 ² Nm ²	- ΔG_{mic} KJ/mol	- $\Delta G_{mic}/CH_2$ KJ/mol	- ΔG_{ads} KJ/mol	- $\Delta G_{ads}/CH_2$ KJ/mol
Dodecyl	IIa	7.1 x 10 ⁻³	28.0	1.25	132.8	12.01	0.11	12.29	0.12
Hexadecyl	IIb	6.68 x 10 ⁻³	30.0	1.42	116.9	12.29		12.59	
Octadecyl	IIc	6.3 x 10 ⁻³	32.0	1.6	103.8	12.58		12.90	

Table 3. Surface Tension Reduction (C₂₀), Efficiency (PC₂₀), cmc/C₂₀ Ratios, Effectiveness (π_{cmc}) and Effective Number of Carbon Atom (N_{eff}) of 2 -Carboxy-N-(Dodecanoyl (Ethyl)Ammonio)Carbonothioyl)- N-Ethylbenzenaminium Iodide at the liquid /air interface at 25C°

R	Abbreviation	C ₂₀	PC ₂₀	cmc\C ₂₀	π_{eff}	π_{cmc} mN/m	N _{eff}
Dodecyl	IIa	1.24 x 10 ⁻³	2.91	5.73	26.74	44	15.5
Hexadecyl	IIb	1.12 x 10 ⁻³	2.95	5.96	26.82	42	19.5
Octadecyl	IIc	0.89 x 10 ⁻³	3.25	7.08	28.97	40	21.5

Table 4: Antimicrobial Activity of the Synthesized Thio-based Cationic Amphiphiles

Tested micro organisms Samples	Mean of Zone diameter (mm) different concentrations			
	Gram-positive		Gram-negative	
	Streptococcus pneumoniae (RCMB 010010)	Bacillissubtilis (RCMB 010067)	Pseudomonas aeruginosa (RCMB 010043)	Escherichia coli (RCMB 010052)
IIa	21.2±0.58	24.2± 1.2	16.5± 1.2	20.1 ± 1.2
IIb	21.9±0.72	25.5±0.58	17.0± 1.2	20.4± 0.63
IIc	23.2±0.63	27 ± 1.2	17.2± 1.2	20.6 ± 1.5
IIIa	18.6± 1.2	20± 1.5	14.5± 1.2	20 ± 0.63
IIIb	20.3 ± 1.5	21± 0.63	15.5 ± 1.2	20.3 ± 0.58
IIIc	21.0± 1.2	22 ± 1.2	16 ± 1.2	21 ± 1.2
Control	23.8± 0.72	32.4 ± 0.63	17.3± 0.58	19.9 ± 0.63

Ampiciline is used as reference for Gram positive bacteria and Gentamicine is used as reference For Gram negative bacteria. IIa-c is 2 -Carboxy-N-(Alkanoyl (Ethyl)Ammonio)Carbonothioyl)-N-Ethylbenzenaminium Iodide, III a-c is 4 -Sulphonate -N-(fatty alkanoyl (Ethyl)Ammonio)Carbonothioyl)- N-Ethylbenzenaminium Iodide a= C₁₂, b = C₁₆, C = C₁₈

Confirmation of the structures of the prepared compounds illustrated in figs 1-6 Spectra of 2 -

Carboxy-N- (Dodecyl (Ethyl)Ammonio) Carbonothioyl)-N- Ethylbenzenaminium Iodide (IIa).

IR spectra (cm⁻¹, KBr) for compound IIa showed strong absorption band at 3444.87, 3417.88 cm⁻¹ for (N–H) and 3.037.5 cm⁻¹ for (Ar–H, C–H SP²), 2824.09, 2850.79 cm⁻¹ for (C–H, SP³) cm⁻¹ for 1627.76 cm⁻¹ for (CO) and 1720.79 cm⁻¹ for (COOH) fig(1).

¹HNMR Spectra for compound IIa showed chemical shift (δ, DMSO): 0.87-0.91 (3H, t, CH₃), chemical shift (δ) 1.27-1.35, chemical shift (δ) (24H, (CH₂, m), chemical shift (δ) 1.61-1.67 for (3H, t, CH₃), chemical shift (δ) 2.32-2.34 for (4H, 2CH₂), chemical shift (δ) 7.28 for (2H, d, CH) and chemical shift (δ) 12 (H,S, COOH)n. fig (2).

ESI-MS for compound IIa showed m/z; (704.0, 7.4 %) and base peak m/z 156.2. fig (3).

Spectra of 4 –Sulphonate –N–((Dodecanoyl (Ethyl)Ammonio) Carbonothioyl)-Ethylbenzenaminium Iodide (IIa).

IR spectra (cm⁻¹, KBr) showed strong absorption band at 3472.2, 3370.9 cm⁻¹ for (N–H), 3195.6 cm⁻¹ for (Ar–H, C–H SP²), 2853.9, 2924.7 cm⁻¹ for (C–H SP³), 1627.5 cm⁻¹ for (CO) and 1160.4 cm⁻¹ for (SO₃H). fig (4).

¹HNMR for IIIa: showed chemical shift (δ, DMSO) 0.88-0.91 for (3H, t, (CH₃), (δ, DMSO) 1.25-1.3 for (24H, m, (CH₂)₁₂), (δ, DMSO) 1.61-1.65 for (3H, t, CH₃), (δ, DMSO) 7.32 for (2H, d, CH) and (δ, DMSO) 8 for (NH, S). fig(5).

ESI-MS for compound IIIa showed m/z (740.0, 63.97 %) and base peak 641.4. fig (6).

III.1 Surface tension and Critical Micelle Concentration (CMC)

Surface tension was measured to determine the Critical Micelle Concentration (CMC). The variation for the surface tension vs. log. concentration of the synthesized surfactants at 25°C are shown in figs (7, 9). In all cases surface tension of the solution decrease rapidly with the initial increase in surfactants concentration. At the break point, The surface tension began to decrease slowly, indicating the formation of surfactant aggregations. The concentration at the break point is shown as the CMC of the cationic surfactants series. It was found that with hydrophobic chain length increasing, the cmc value gradually decreased for the enhanced hydrophobic interaction between the longer alkyl chains, Tables 2, 7. Derivatives which contain sulfonate groups (IIIa-c) have a higher depression in cmc values than those containing carboxylate groups (IIa-c) and the maximum value for cmc was observed for (IIIa-c); 9.4×10^{-3} mmol⁻¹ while the lowest value was observed for (IIc) at 6.3×10^{-3} mmol⁻¹ at 25°C.

From surface tension lowering ability γ_{CMC} in Tables (2, 7), it can be seen that γ_{CMC} of compounds (IIa-c), **(IIIa-c) increases with increasing hydrophobic chain length.**

III-2 Maximum surface excess (Γ_{max}) and minimum surface area (A_{min})

The calculated values of the maximum surface excess showed increasing trend from C₁₂ to C₁₈ as represented from the slope of pre-CMC region of surface tension profile; figs 7, 9.

The values of (A_{min}) listed in tables 2,7 Increasing the maximum surface excess values indicates the increasing of adsorbed molecules at the interface, hence the area available for each molecule will decrease. That causes the compacting of surfactant molecules at the interface to form denser layer.

From Tables (2,7), it can be seen the maximum area occupied at the interface was 147.5 nm² which obtained for IIIa surfactants, While the lowest area was 103.8 nm² for IIc surfactants and A_{min} is greatly influenced by the hydrophobic alkyl chain length. Increasing the hydrophobic alkyl chain length decreases A_{min} values at the interface. Also, increasing the maximum surface excess (the accumulation of surfactant molecules at the air/water interface, Γ_{max}) indicates the surfactant/water repulsion in the bulk of the solution which pumps the surfactant to the air/water interface. That can be referred to the change of the hydrophobicity of the molecules under consideration by the type of substituents.

III- 3 Effectiveness (π_{CMC}), Efficiency (PC₂₀) and (CMC/C₂₀) ratio

Other important surface active parameters, such as the adsorption efficiency (P_{C20}) the effectiveness of surface tension reduction (π_{CMC}) and (CMC/C₂₀) ratio can also be obtained from the surface curve as listed in tables 2,7.

By inspected the data in Tables 2,7, the surface tension lowering ability (γ_{CMC}) of cationic amphiphiles increase with increasing hydrophobic chain length. This explained the increase of hydrophobicity of surfactants and compounds with dodecyl group in the alkyl chain for two series that showed the maximum surface lowering ability. The effectiveness (π_{CMC}) values showed gradual decrease by increasing the hydrophobic chain length indicating the increasing of accumulated surfactants molecules at the interface. The largest π_{CMC} was observed for IIa 44 mN/m.

Adsorption efficiency, (P_{C20}) of the surfactant concentrations that are capable of suppressing the surface tension of the solution by 20 mN/m Therefore, C₂₀ can be a measurement for the adsorption efficiency of the surfactant molecule greater the value of PC₂₀ is, the higher adsorption efficiency of surfactant is. The efficiency values PC₂₀ of the targeted surfactants (Tables 3, 7) showed their good tendency to modify the surface activity of their solutions at considerably low concentrations.

Table 5: Antifungal Activity of the Synthesised Thio-based Cationic Amphiphiles:

Tested microorganisms	Mean of Zone diameter (mm) different concentrations			
	Aspergillus fumigatus (RCMB 02568)	Syncephalastrum racemosum (RCMB 05922)	Geotricum candidum (RCMB 05097)	Candida albicans (RCMB 05036)
IIa	22.3 ± 0.72	19.2 ± 0.58	24.2 ± 0.63	NA
IIb	22.9 ± 0.58	19.5 ± 0.63	22.8 ± 0.72	NA
IIc	23.2 ± 0.58	19.6 ± 0.72	20 ± 0.72	NA
IIIa	20.6 ± 0.63	18.4 ± 0.58	23 ± 0.72	NA
IIIb	21.6 ± 0.58	18.7 ± 0.63	21.5 ± 1.2	NA
IIIc	23.4 ± 0.58	19.5 ± 0.72	20.7 ± 0.72	NA
Control	23.7 ± 1.2	19.7 ± 0.72	28.7 ± 0.72	25.4 ± 0.58

Amphotericin B is used as reference for fungi is 2 -Carboxy-N-(Alkanoyl (Ethyl)Ammonio)Carbonothioyl)- N-Ethylbenzenaminium Iodide, III_{a-c} is 4 -Sulphonate -N-((fatty alkanoyl (Ethyl)Ammonio)Carbonothioyl)- N-Ethylbenzenaminium Iodide, IIa-c, a = C₁₂, b = C₁₆, c = C₁₈
 NA: No activity

Table 6: Physicochemical properties of 4 -Sulphonate -N-(fatty alkanoyl (Ethyl)Ammonio)Carbonothioyl)- N-Ethylbenzenaminium Iodide derivatives

R	Abbreviation	Molecular formula	Mol.Wt	Colour	M.P	Yield %	Elemental analysis						
							C %	H %	N %	S %	I %	O %	
Dodecyl	III _a	C ₂₄ H ₄₂ N ₂ S ₂ O ₄ I ₂	740.01	Reddish brown	303-306	77	38.91	5.72	3.78	8.65	34.29	8.65	8.64
							38.93	5.75	3.80	8.68	34.32	8.67	8.66
Hexadecyl	III _b	C ₂₈ H ₅₀ N ₂ S ₂ O ₄ I ₂	796.16	Orange	310-315	80	42.47	6.37	3.54	8.09	32.08	7.45	8.08
							42.49	6.39	3.56	8.13	32.11	7.48	8.11
Octadecyl	III _c	C ₃₀ H ₅₄ N ₂ S ₂ O ₄ I ₂	824.19	Brown	315-320	85	43.68	6.60	3.40	7.77	30.79	7.76	7.76
							43.70	6.63	3.43	7.79	30.82	7.78	7.78

At air/ water interface. So the Increasing the hydrophobic chain length of the studied amphiphiles results a decrease in the surface tension of the surfactant solution indicating the high tendency of the longer hydrophobic molecules to adsorb at the interface. The lowest PC₂₀ value was observed to 3a 2.85. The value of CMC/C₂₀ ratio is to determine structural factors in the adsorption and micellization process. From Tables 2, 7, it can be seen that The CMC/C₂₀ ratios increase with the increase of hydrophobic chain length for series IIa-c, IIIa-c. This result suggests that cationic surfactants with long hydrocarbon chains adsorb at the air/ water interface in preference to forming micelles, whereas cationic surfactants with short hydrocarbon chains do not adsorb efficiently at the interface. The affinity of a particular surfactant to reduce surface tension of solvent depend upon CMC/C₂₀ ratio, greater the observed value, greater is the tendency of amphiphile to reduce surface tension of the system. It can be found that the CMC/C₂₀ values of cationic surfactants containing sulphonate group and longer chain length are higher than those of cationic surfactants containing carboxylate group of the same chain length. the CMC/C₂₀ values (7.08) for IIc (carboxylate derivatives) and (8.43) for IIIc (sulphonate derivatives).

III-4 The interfacial activity (I_{act}.)

The interfacial activity (I_{act}.) which expressed physicochemical parameter.

$\Delta G_{ads}/A_{min}$ where (ΔG_{ads}) is a standard free energy of adsorption of the surfactant at the air/ solution interface and (A_{min}) is the minimum surface area of the surfactant as shown in Tables 2,6. It is clear that I_{act} decreased by increasing alkyl chain length and high depression were observed for compound IIa.

3.2 Thermodynamic Parameters; Free energy of micellization and adsorption (ΔG_{mic} and ΔG_{ads}).

Thermodynamic functions describe the energetic parameters of adsorption and micellization process of surfactants molecules either at the interface or in bulk of their solutions.

Micellization and adsorption free energies are in negative values, Which indicate their spontaneous occurrence in the solution, While large negative value indicate the majority of one process than the other. The standard free energies for adsorption and micellization of the synthesised surfactants are negative. The value of adsorption free energies are more negative than those of micellization process. That indicate the high tendency of the synthesised surfactants towards adsorption at the interface than micellization and the tendency towards adsorption referred to the interaction between the aqueous phases and the hydrophobic chains which pumps the surfactants molecules to the interface. The presence of these surfactants at the interface decrease the difference in phases interactions. The maximum depression in ΔG_{ads} was observed at -12.9 KJ / mol for IIc. This result showed the

applicability of these surfactants in several applications including: emulsification, detergency and corrosion inhibition application^[52].

IV- Biological Activity

Quaternary nitrogen compounds act as amphiphilic cations in aqueous solution so many cationic surfactants are used as pesticides and antimicrobial against wide spectrum of organisms as well as for killing sulfate reducing bacteria which can exist in petroleum field and cause many problems such as block out pipe line, produce H₂S toxic gas, form iron sulfide hard scale and cause microbial corrosion^[53-55].

Tables (4, 5) show the antibacteria and anti fungal activities of the prepared compounds IIa-c, IIIa-c. Ampicillin and gentamicin were taken as the reference drugs for antibacterial activity and amphotericin B as antifungi. By analyzing the data in table 4,5, it is clear that the difference in activity depend on the prepared cationic surfactants. Increasing the hydrophobic chain length increase the antibacterial activity, these results are in consistence with the adsorption tendency of these amphiphiles at the interface. The octadecyl derivatives (C18) in the two series showed the maximum antimicrobial activities

against the tested bacterial and fungi strains. Overall, it was observed that the synthesized cationic surfactants exhibited higher antibacterial activity against Gram – positive bacteria than Gram negative bacteria. This can be explained by different cell membrane structures of the two bacterial types. On the other hand, the most effective factor is the carbon chain length; it was found that, as the carbon chain length (hydrophobic part) increases, the compound efficiency also increases. That can be related to their ability of adsorption at the interface. Increasing the absorbability^[56-62] increases their action on the cell membrane. The results of the antifungal activity of the synthesized quaternary ammonium salts showed that negative effect against the most pathogenic fungal strain,. That may be ascribed to the resistivity of the fungi strain to the aggressive environmental components due to the rigidity of their cellular membranes, so that these fungi have high resistance against synthesized cationic surfactants and best results for compounds (IIIa-c) for all fungi species except *Candida albicans* (RCMB 05036) as illustrated in Table 5.

V- Antitumor Activity using EAC:

Table 7: Critical micelle Concentrations (CMC), Surface tension at cmc (γ_{cmc}), surface max (τ_{max}), Minimum Area\Molecule (A^2), Standard Free Energy of Micellizaion (ΔG_{mic}), Standard Free Energy of Micellizaion per methylene group ($\Delta G_{mic}/CH_2$), Standard Free Energy Of adsorption (ΔG_{ads}) and Standard Free Energy Of adsorption per methylene group $\Delta G_{ads}/CH_2$ of 4 –Sulphonate –N-(fatty alkanoyl (Ethyl)Ammonio)Carbonothioyl)- N-Ethylbenzenaminium Iodide at Liquid Air Interface at 25C°

R	Abbreviations	CMC mol/l	γ_{cmc} mN/m	$\tau_{max} \times 10^{-10}$ mole/cm ²	Area mole A ² × 10 ² nm ²	- ΔG_{mic} KJ/mol	- $\Delta G_{mic}/CH_2$ KJ/mol	- ΔG_{ads} KJ/mol	- $\Delta G_{ads}/CH_2$ KJ/mol
Dodecyl	IIIa	9.40 x 10 ⁻³	29.00	1.13	147.5	11.56	0.124	11.97	0.111
Hexadecyl	IIIb	8.90 x 10 ⁻³	32.00	1.14	145.6	11.69		12.26	
Octadecyl	IIIc	7.50 x 10 ⁻³	34.00	1.17	141.3	12.12		12.56	

From Table 9, it is clear that the cytotoxic effects of these compounds were dose dependent, i.e. by increasing the concentration on these compounds in the culture media; the percentage of non-viable cells (EAC) is increased. Also from data of Table 9 one observes that upon testing the compounds which contains more than one benzene ring, the antitumor activity dramatically increases from 23 % of non-viable EAC cells at 25 mg/ml to 95% of these cells at 100 mg/ml and also NVC% increases by increasing the chain length of the hydrophobic radical. To achieve the maximum therapeutic damage of tumor cells using the minimum concentration of drugs, the tumor cells lines used in this study for 3a which causes the death of 95% of EAC cell are liver carcinoma (HEPG2), breast carcinoma (MCF 7) and colon carcinoma (HCT 116), Tables 10 and 11. The 50% growth inhibitory concentration (IC 50) values for 3a-c at different concentration ranged from 5 to 50 mg/ml

is recorded (18.7,16.4,14.2) whill (IC 50) for 2a-c (12.5, 10.6, 8.7). Data showed that 3a was found to exhibit high activity in vitro system on the tumor cell lines investigated and the highest cytotoxic effect on HEPG2, HCT116 and MCF7, respectively.

In our research, we found that 2a surfactant affects tumor tissue at very low concentrations at values lower than their cmc values which means that there is a strong relationship between very small values of cmc of this compound and its ability to reach IC50 values under very low concentration. This is due to the fact that increasing the concentration of surfactants causes an increase in the adsorption process on cells membranes till the cmc is reached; after this adsorption slowly decreases and then stops due to the formation of micelles, which prevent mobility and suppresses antitumor activity. increasing their doses. A significant number of studies have demonstrated successful control of the particle size,

morphology and surface functionalization of the surfactants for diverse applications [64–68].

Antitumor activity of these surfactants was evaluated against three human tumor cells such as HEPG2 (liver), HCF7 (breast) and HCT116 (colon). The cytotoxic effects of these compounds were dose dependent, i.e. by increasing the concentration on

these compounds in the culture media; the percentage of non-viable cells (EAC) was increased. Also, data showed that (IIIa) was found to exhibit higher activity than (IIa) in vitro system on the tumor cell lines investigated and the highest cytotoxic effect on HEPG2, HCT116 and MCF7, respectively.

Table 8: Surface Tension Reduction (C_{20}), Efficiency(PC_{20}), cmc/C_{20} Ratios, π_{eff} Effectiveness (π_{cmc}) and Effective Number of Can atom (N_{eff}) of 4 – (Ethyl)Ammonio)CarbonothioylSulphonate -N-(fatty alkanoyl)- N-Ethylbenzenaminium Iodide derivatives at liquid air interface at 25°C.

R	Abbreviations	C_{20}	PC_{20}	CMC/C_{20}	π_{eff}	Π_{CMC} mN/m	N_{eff}
Dodecyl	IIIa	1.4×10^{-3}	2.85	6.71	25.2	43	18.0
Hexadecyl	IIIb	1.12×10^{-3}	2.95	7.95	25.8	40	22.0
Octadecyl	IIIc	0.89×10^{-3}	3.05	8.43	26.1	38	24.0

Table 9: Antitumor activities of thio based cationic surfactants Derivatives Using (E.A.C.)

Samples name	Sulphanilic IIIa-c				Anthranilic IIa-c				
	% Inhibition of cell viability								
			Ug/ml						
Concentrations	Abbreviations	25%	50%	100%	Concetrations	abbreviations	25%	50%	100%
Dodecyl	IIIa	23%	47%	95%	IIa		13%	32%	90%
Hexadecyl	IIIb	22%	45%	90%	IIb		15%	35%	88%
Octadecyl	IIIc	20%	40%	87%	IIc		18%	37%	80%

Table 10: Inhibitory effect of 2- Carboxy N-(lauroyl (Ethyl)Ammonio)Carbonothioyl)- N-Ethylbenzenaminium Iodide

Compound(IIa)	Cell lines		
	HEPG2	MCF7	HCT116
2- CarboxyN-(lauroyl (Ethyl)Ammonio)Carbonothioyl)- N-Ethylbenzenaminium Iodide	IC_{50} 12.5 μ g	IC_{50} 8.7 μ g	IC_{50} 10.6 μ g

Table 11: Inhibitory effect of 4- Sulphonate N- (lauroyl (Ethyl)Ammonio)Carbonothioyl)- Ethylbenzenaminium Iodide

Compound(IIIa)	Cell lines		
	HEPG2	MCF7	HCT116
4- Sulphonate N- (lauroyl (Ethyl)Ammonio)Carbonothioyl)- N-Ethylbenzenaminium Iodide	IC_{50} 18.7 μ g	IC_{50} 14.2 μ g	IC_{50} 16..4 μ g

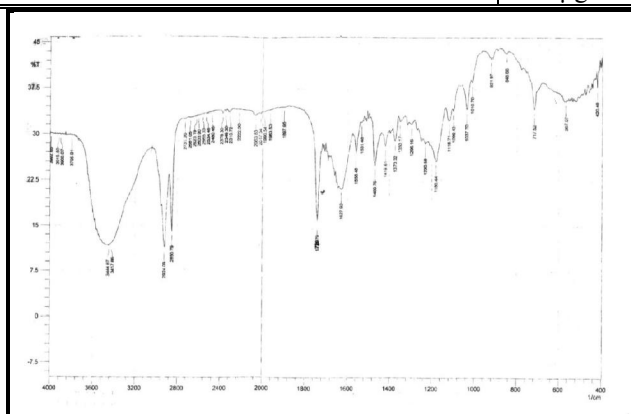


Figure (1) FTIR spectra of 2-carboxy- (Dodecanoyl (Ethyl ammonio) carbonothioyl)-N-Ethyl benzenaminium Iodide

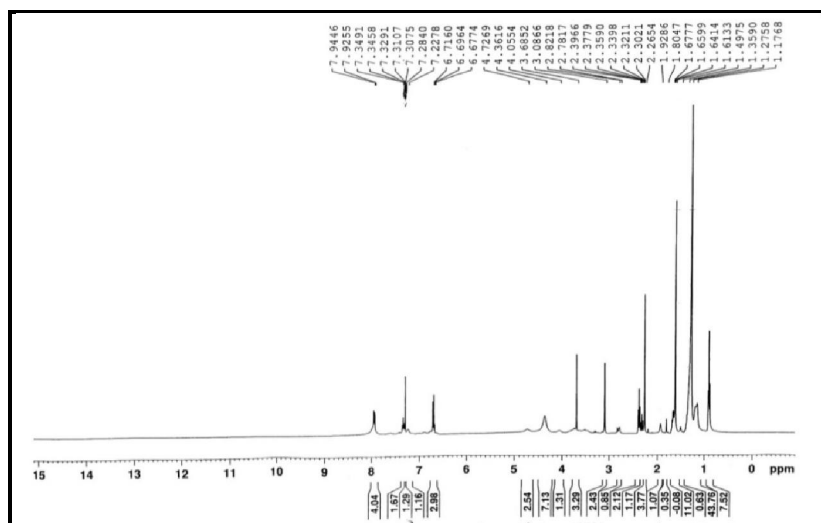


Figure. (2) ^1H NMR spectra of 2-carboxy-(Dodecanoyl(Ethyl ammonio) carbonothioyl-N-Ethyl benzenaminium Iodide

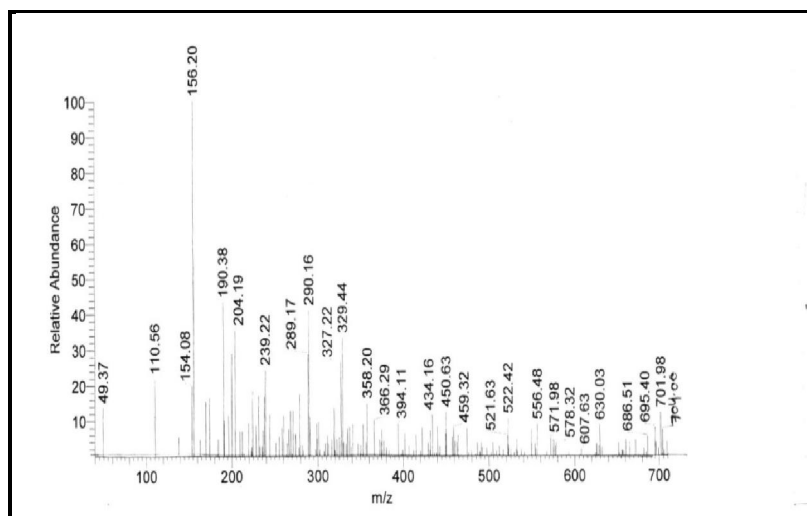


Figure. (3) Mass spectra of 2-carboxy-(Dodecanoyl(Ethyl ammonio) carbonothioyl-N-Ethyl benzenaminium Iodide

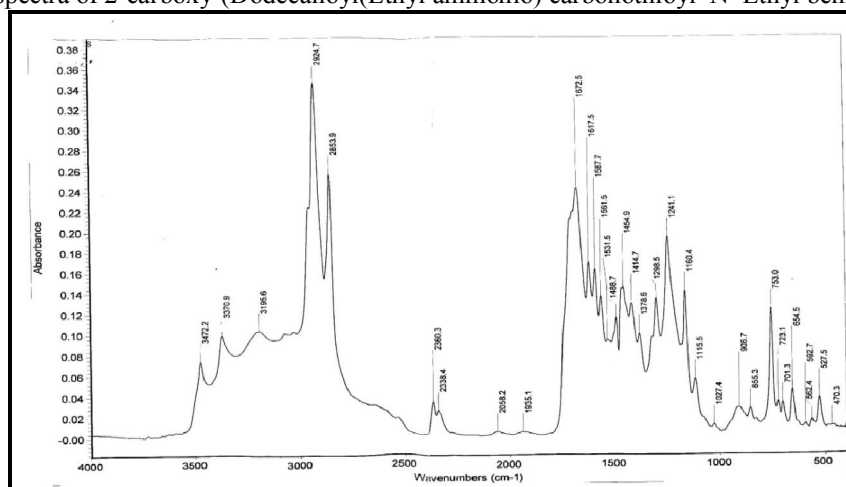


Figure. (4) FTIR spectra of 4-sulphonate-N-(Dodecanoyl ethyl ammonio)carbonothioyl-N-Ethyl benzenaminium Iodide

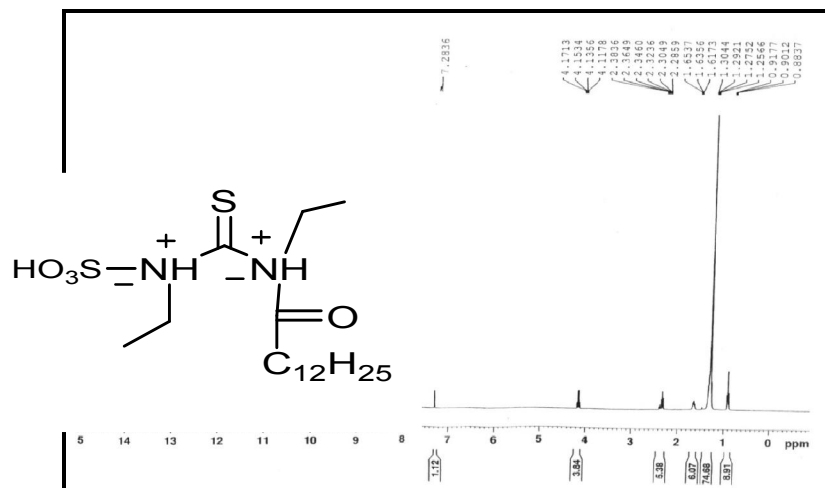


Figure (5): ^1H NMR Spectra of 4-sulphonate-N-(Dodecanoyl ethyl ammonio) carbonothioyl-N-Ethyl benzenaminium Iodide

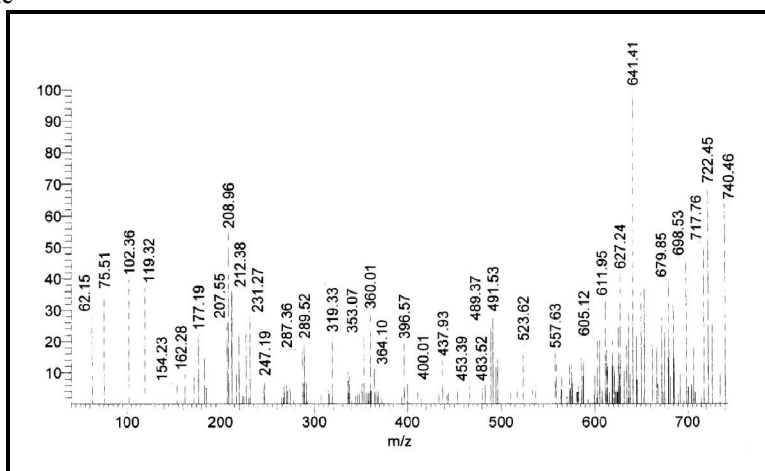


Figure (6) Mass spectra of 4-sulphonate-N-(Dodecanoyl ethyl ammonio) carbonothioyl-N-Ethyl benzenaminium Iodide

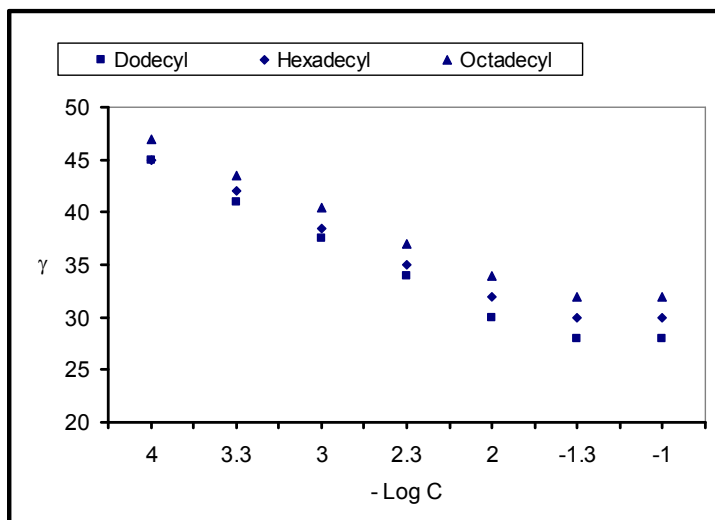


Figure (7): Represents surface tension (γ) vs-log C of 2-Carboxy-N((Dodecanoyl)(Ethyl)Ammonio)Carbonothioyl)-N-Ethylbenzenaminium Iodide in distilled water

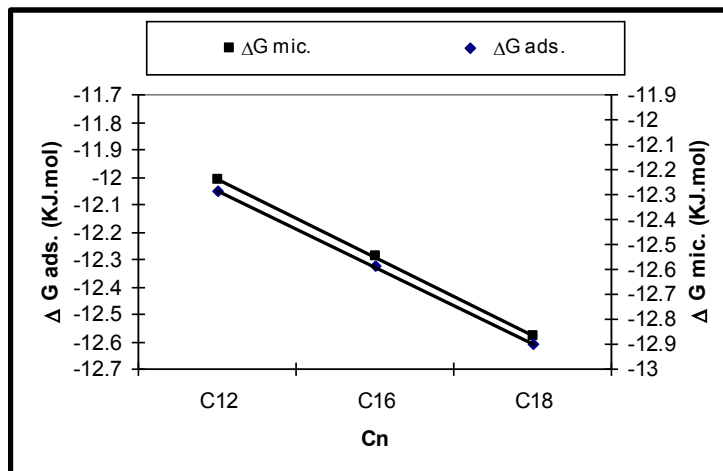


Figure. (8) Represents Standard Free Energy of Micellization (ΔG_{mic}) and Standard Free Energy of Adsorption (ΔG_{ads}) vs Carb. no. of 2-Carboxy-N-(Dodecanoyl) (Ethyl)Ammonio) Carbonothioyl)- N-Ethylbenzenaminium Iodide in distilled water

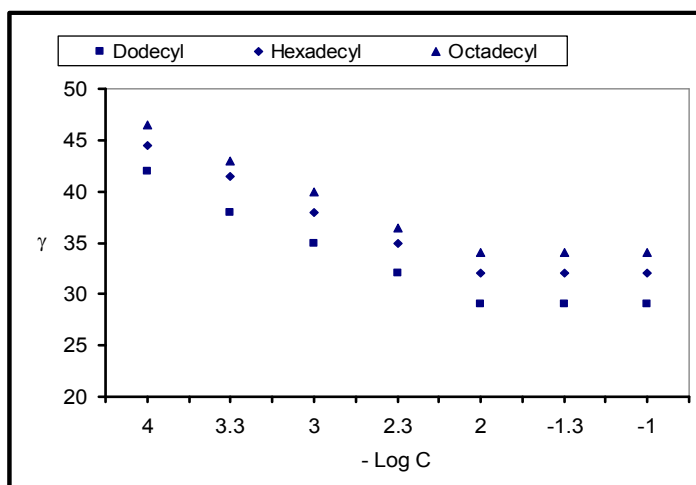


Figure. (9): Represents surface tension (γ) vs-log C of 4-Sulphonate-N-(fatty alkanoyl) (Ethyl)Ammonio) Carbonothioyl)- N-Ethylbenzenaminium Iodide in distilled water

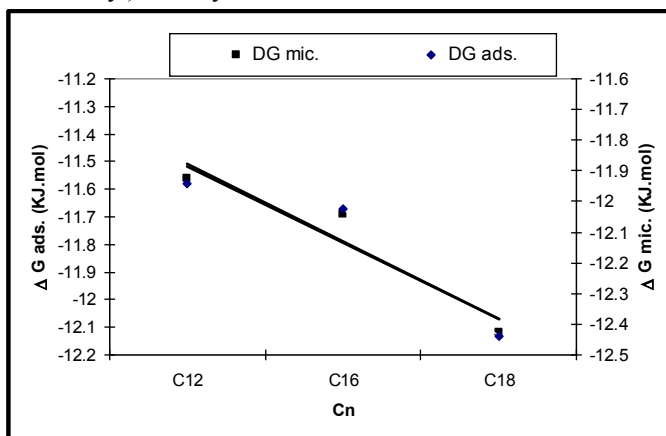


Figure. (10) Represents Standard Free Energy of Micellization (ΔG_{mic}) and Standard Free Energy of Adsorption (ΔG_{ads}) vs Carb. no. of 4-Sulphonate-N-(fatty alkanoyl) (Ethyl)Ammonio) Carbonothioyl)- N-Ethylbenzenaminium Iodide in distilled water

6 Conclusion:

The synthesized surfactants showed higher surface activity and their activity was influenced by the chemical structures and hydrophobic chain length. The temperature of the solution causes a significant effect on the thermodynamic parameters of adsorption and micellization. Also, the antimicrobial activity of quaternary salts toward bacteria and fungi were high and depended on the chemical structures of the biocide these surfactants in their solutions were also calculated. Biocidal activity of prepared amphiphiles was investigated against bacterial and fungi species which showed that our surfactants are in general capable of inhibiting the growth of bacteria and fungi to a moderate extent. Also, it is clear that the biological activities of the synthesized surfactants against the tested microorganism increase by increasing their doses. Antitumor activity of these surfactants was evaluated against Ehrlich ascites carcinoma (EAC) and against three human tumor cells such as HEPG2 (liver), HCF7 (breast) and HCT116 (colon). The cytotoxic effects of these compounds were dose dependent, i.e. by increasing the concentration on these compounds in the culture media; the percentage of non-viable cells (EAC) was increased. Also, data showed that surfactant compound (IIIa) was found to exhibit higher activity than surfactant compound (IIa) in vitro system on the tumor cell lines investigated and the highest cytotoxic effect on HEPG2, HCT116 and MCF7, respectively.

Acknowledgments:

We would like to thank the chemistry department, Faculty of Science Al-Azhar University for supporting this work, Cancer National Institute, Cairo, Egypt for Antitumor activities measurements.

References

1. Kinzler, Kenneth W., Vogelstein and Bert V.; "The genetic bases of humane cancer" 2nd illustrated, revised ed.) New York Mc Graw-Hill, Medical Pub. Division.5. (2002) DOI: 10.1056/NEJM199810013391418.
2. Nishikawa Y., Yoshimoto K., Okabe M., and Fukuoka F.; Chemical and biochemical studies on carbohydrate esters III. Antitumor activity of unsaturated fatty acids and their ester derivatives against Ehrlich ascites carcinoma. *Journal of Chem. Pharm. Bull. (Tokyo)* 24:756-762 (1976) DOI.org/10.1248/cpb.24.756.
3. Franchi G., Morasca L., Reyers-Degli-Innocenti I. and Garattini S. T.; WR 1339 (TWR), an inhibitor of cancer dissemination and metastases. *European Journal of Cancer*, 7:533 - 544. (1971), DOI: 10.1016/0014-2964(71)90059-4.
4. Silk M. and Sigman E.; Effect of pluronic-F68 on the development of tumor metastasis. *Journal of Cancer Phila*, 29:171-172 (1972), DOI: 10.1002/1097-0142(197201)29.
5. Abdelftah M. Badawi, Nadia I. Zakhary, Salwa M. I. Morsy, Gilane M. Sabry, Mervat M. Fouad, and Ahmed M. Mousa; Biochemical study on the effect of Metallo-Surfactant and its loaded nano-analogue as anticancer drug; *Journal of American Science*; 8(3) 2012, Doi.org/10.7537/marsjas080512.36.
6. Martin V.I., de la Haba R.R., Ventosa A., Congiu E., Ortega-Calvo J.J., Moya M.L., Colloidal and biological properties of cationic single-chain and dimeric surfactants, *Colloid. Surf. B Biointerfaces* 114 (2014) 247–254 - Doi.org/10.1016/j.colsurfb.2013.10.017.
7. Tavano, L. A. Pinazo, M. Abo-Riya, M.R. Infante, M.A. Manresa, R. Muzzalupo, L. Perez, Cationic vesicles based on biocompatible diacyl glycerol-arginine surfactants: physicochemical properties, antimicrobial activity, encapsulation efficiency and drug release, *Colloid. Surf. B Biointerfaces* 120 (2014)160–167. DOI.10.1016/j.colsurfb.2014.04.009
8. Greber, K.E. M. Dawgul, W. Kamysz, W. Sawicki, J. Lukasiak, Biological and surface-active properties of double-chain cationic amino acid-based surfactants, *Amino Acids* 46 (2014) 1893–1898, DOI: 10.1007/s00726-014-1744-9.
9. Oblak E., A. Piecuch, A. Krasowska, J. Luczynski, Antifungal activity of geminiquaternary ammonium salts, *Microbiol. Res.* 168 (2013) 630–638. doi.org/10.1016/j.micres.2013.06.001.
10. Schlecht U., R.P. St Onge, T. Walther, J.M. Francois, R.W. Davis, Cationic amphiphilic drugs are potent inhibitors of yeast sporulation, *PLoS One* 7 (2012)e42853 DOI..org/10.1371/journal.pone.0042853.
11. Pandey N.K., Ghosh S. and Dasgupta S., Effect of surfactants on preformed fibrils of human serum albumin, *Int. J. Biol. Macromol.* 59 (2013) 39–45 DOI. 10.1016/j.ijbiomac.2013.04.014.
12. Li P., Nielsen H.M., Fano M., Mullertz A., Preparation and characterization of insulin-surfactant complexes for loading into lipid-based drug delivery systems, *J. Pharm. Sci.* 102 (2013) 2689–2698 DOI: 10.1002/jps.23640.
13. Mintzer M.A., Simanek E.E., Nonviral vectors for gene delivery, *Chem. Rev.* 109(2009) 259–302 DOI: 10.1021/cr800409e.
14. Parvizi, Jubeli P. E., L. Raju, N.A. Khalique, A. Almeer, Allam H., Manaa, M.A. H. Larsen, D. Nicholson, Pungente M.D., Fyles, T.M. Aspects

- of nonviral gene therapy: correlation of molecular parameters with lipoplex structure and transfection efficacy in pyridinium-based cationic lipids, *Int. J. Pharm.* 461 (2014) 145–156 DOI 10.1016/j.ijpharm.2013.11.045
15. Fabregas A. Sanchez-, N. Hernandez, J.R. Tico, E. Garcia-Montoya, P. Perez-Lozano, J.M. Sune-Negre, C. Hernandez-Munain, C. Sune, M. Minarro, A new optimized formulation of cationic solid lipid nanoparticles intended for gene delivery: Development, characterization and DNA binding efficiency of TCERG1 expression plasmid, *Int. J. Pharm.* 473 (2014) 270–279. DOI: 10.1016/j.ijpharm.2014.06.022 .
 16. Yin J., Shi S., Hu J and Liu S., Construction of polyelectrolyte-responsive micro-gels, and polyelectrolyte concentration and chain length-dependent adsorption kinetics, *Langmuir* 30 (2014) 9551–9559, DOI: 10.1021/la501918s.
 17. Yamada H., Urata C., Higashitani S., Aoyama Y., Yamauchi Y. and Kuroda K, Critical roles of cationic surfactants in the preparation of colloidal mesostructured silica nanoparticles: control of mesostructure, particle size, and dispersion, *ACS Appl. Mater. Interfaces* 6 (2014) 3491–3500., DOI: 10.1021/am405633r.
 18. Fanguiero J.F., Andreani T., Egea M.A., Garcia M.L., Souto S.B., Silva A.M. and Souto E.B., Design of cationic lipid nanoparticles for ocular delivery: development, characterization and cytotoxicity, *Int. J. Pharm.* 461 (2014) 64–73, DOI: 10.1016/j.ijpharm.2013.11.025.
 19. Chou T.H., Liang C.H., Lee Y.C. and Yeh L.H., Effects of lipid composition on physico-chemical characteristics and cytotoxicity of vesicles composed of cationic and anionic dialkyl lipids, *Phys. Chem. Chem. Phys.* 16 (2014) 1545–1553, DOI: 10.1039/C3CP54176B.
 20. Doktorovova S, Santos D.L., Costa I, Andreani T., Souto E.B., Silva A.M., Cationic solid lipid nanoparticles interfere with the activity of antioxidant enzymes in hepatocellular carcinoma cells, *Int. J. Pharm.* 471 (2014) 18–27, DOI: 10.1016/j.ijpharm.2014.05.011.
 21. Duangjit S., Pamornpathomkul B., Opanasopit P., Rojanarata T., Obata Y., Takayama K. and Ngawhirunpat T., Role of the charge, carbon chain length, and content of surfactant on the skin penetration of meloxicam-loaded liposomes, *Int. J. Nanomed.* 9 (2014) 2005–2017, DOI.org/10.2147/IJN.S60674.
 22. Muthukumar N., S. Maruthamuthu and N. Palaniswamy, Role of cationic and non-ionic surfactants on biocidal efficiency in diesel-water interface, *Colloids Surf. B Biointerfaces* 57 (2007) 152–160, Doi:10.1016/j.colsurfb.2007.01.019.
 23. Tawfik S. M., Simple one step synthesis of gemini cationic surfactant-based ionic liquids: Physicochemical, surface properties and biological activity *J. Mol. Liq.* 209 (2015) 320–326, Doi.org/10.1016/j.molliq.2015.05.054.
 24. Andrew J., Ruth B. Mc, G.L., Louiss E.M., Carl C. and Peter G.t., Effects of Quaternary-Ammonium-Based Formulations on Bacterial Community Dynamics and Antimicrobial Susceptibility *Appl. Environ. Microbiol.* 70 (6) (2004) 3449–3456, DOI 10.1128/AEM.03141.
 25. Negm N.A., Elkholy Y.M., Ghuiba F.M., Zahran M.K, Mahmoud S.A. and Tawfik S.M., 'Benzothiazol-3-ium Cationic Schiff Base Surfactants: Synthesis, Surface Activity and Antimicrobial Applications against Pathogenic and Sulfur Reducing Bacteria in Oil Fields', *J. Dispersion Sci. Technol.* 32 (2011) 512–518, DOI: 10.1080/01932691003756902.
 26. Shaban S.M., Sayed A., Tawfik S.M., Abd-Elal A., Aiad I., Corrosion inhibition and Biocidal effect of some cationic surfactants based on Schiff base, *J. Ind. Eng. Chem.* 19 (2013) 2004–2009, DOI.org/10.1016/j.jiec.2013.03.013.
 27. Negm N.A., Aiad I., Tawfik S.M., Screening for Potential Antimicrobial Activities of Some Cationic Uracil Biocides Against Wide-Spreading Bacterial Strains, *J. Surfactants Deterg.* 13 (2010) 503–511, DOI: 10.1007/s11743-010-1229-0.
 28. Aiad I., Tawfik S.M., Shaban S.M., Abd-Elal A., El-Shafie M., Enhancing of Corrosion Inhibition and the Biocidal Effect of Phosphonium Surfactant Compounds for Oil Field Equipment, *J. Surfactants Deterg.* 17 (2014) 391–40, Doi:10.1007/s11743-013-1512-y.
 29. Augustin, C. and Damour, O. Pharmacotoxicological applications of an equivalent dermis: three measurements of cytotoxicity, *J. Cell Biology and Toxicology*, 11: 167–171 (1995), DOI: 10.1007/BF00756519.
 30. Damour, O., Hua, S.Z., Lasne, F., Villain, M., Rousselle, P., and Collombel, C., Cytotoxicity evaluation of antiseptics and antibiotics on cultured human fibroblasts and keratinocytes. *Journal of the International Society for Burn Injuries* 18: 479–485, (1992), DOI: 10.1016/0305-4179(92)90180-3 .
 31. Steinsvag, S.K., Bjercknes, R. and Berg, O.H. Effects of Topical Nasal Steroids on Human Respiratory Mucosa and Human Granulocytes in Vitro *J. Acta Oto- Laryngologica*, 116: 868–875 (1996), DOI.org/10.3109/00016489609137943.

32. Tripathi, B.J. and Tripathi, R.C., Cytotoxic effects of benzalkonium chloride and chlorobutanol on human corneal epithelial cells in vitro. *Lens and Eye Toxicity Research*, 6: 395–403(1989), Doi: 10.1111/j.1442-9071.2008.01803.x.
33. Youngs, C.G., Epp, A., Craig, B.M. et al., Preparation of long-chain fatty acid chlorides, *J Am Oil Chem Soc* (1957) 34: 107, DOI:10.1007/BF02640442.
34. ImdadUllah, Khurshid Ahmad, Afzal Shah, Amin Badshah Usman Ali Rana, Imran Shakir, Zia-ur-Rehman and Shahan Zeb Khan, Synthesis, Characterization and Effect of a Solvent Mixture on the CMC of a Thio-Based Novel Cationic Surfactant Using a UV-Visible Spectroscopic Technique *J Surfact Deterg* (2013), DOI 10.1007/s11743-013-1554-1.
35. Kiehlbauch, JA., Hannett, GE, Salfinger, M., Archinal, W., Monserrria, C AND Clin, J.: Use of the national committee for clinical laboratory standards guidelines for disk diffusion susceptibility testing in New York state laboratories, *Microbial* 38 (2000) 3341- 3348, DOI: 0095-1137 / 00 / 04.0010.
36. McIlmains, W.F.; Davis, E. V.; Glover, F.L. and Rake, G. W.; The submerged culture of mammalian cells; the spinner culture, *J. Immunol.*, 79,428, (1957), DOI: 10.4172/2155-9538.1000204.
37. Skehan P., Storeng R., et al., Newcoloremtric cytotoxicity assay for anti-cancer drug screening. *J Natl Cancer Ins..* (1990) 82: 1107-1112 DOI: 10.1093 / jnci/82.13.1107.
38. Oda R, Hucb K, and Sauveur J, Gemini surfactants, the effect hydrophobic chain length and dissymmetry. *Chem. commun.* (1997) 56: 2105-2112, DOI: 10.1039/A704069E.
39. NagarajanR, and Ruckern S.E Molecular theory of microemulsion. *Langmuir* (2000) 16: 6400-6408, DOI: 10.1021/la991578t.
40. Zhang, Q., Gao, Z.N., Xu, F., Tai, S.X., Liu, X.G., Mo, S.B., Niu, F.: Surface tension and aggregation properties of novel cationic Gemini surfactants with diethyl ammonium head groups and diamido spacer, *Langmuir* 28 (2012) 11979-11987, DOI: 10.1021 / la3011212.
41. Kamboj, R., Singh, S., Bhadani, A., Kataria, H. AND Kau,G.: Gemini imidazolium surfactants: synthesis and their biophysicochemical study, *Langmuir* 28 (2012) 11969-11978, DOI: 10.1021 / la300920p.
42. Kamboj, R., Singh, S., Chauhan, V.: Synthesis, Characterization and surface properties of N- (2-hydroxy alkyl) –N- (2-hydroxy alkyl) – N-(2-hydroxy ethyl) imidazolium surfactants, *Colloids Surf. A. physicochem.Eng.Asp.* 441 (2014) 233-241, DOI: 10.1016 /j.colsurfa.2013.08.063.
43. Ren, C., Wang, F., Zhange, Z., Nie, H., Li, N., Cui, M.: Synthesis, Surface activity and aggregation behavior of Gemini imidazolium surfactants 1,3- bis (3-alkylimidazolium-l-yl) propane bromide, *Colloids and Surfaces A: Physicochem. Eng. Aspects* 467 (2015) 1-8, DOI: 10.1016 /j.colsurfa. 2014.11.031.
44. Nabel, A.N. AND Salah, M..T.: Characterization, Surface properties and biological activity of some synthesized anionic surfactants, *Journalof industrial and Engineering Chemistry* 20 (2014) 4463-4472, DOI: 10.1016 /j.jiec. 2014. 02.018.
45. Rongqiang I., Fengmeiy., Junli Z., Chefeng X. and Jiben W.: The self –assembly properties of aseries of Polymerizable cationic gemini surfactants: Effect of the acryloxy group, *Colloids and Surfaces A: Physicochem. Eng. Aspectt* 444: 276-282, DOI: 10.1016 / j.colsurfa. 2013. 12.079.
46. Azzam EMS, Negm NA, and Gad EAM: Surface and solubilizatoin activities of 1-amino-2-alkyloxy-naphthalene-4-sodium sulfonates. *J. of Ads. Sci.* (2004) DOI: 10.1260/0263617042844184.
47. Viscardi et al., Viscardi G, Quagliotto P, Barolo C, Savarino B.E, and Fisicazo E, Synthesis, surface and antimicrobial properties of novel cationic surfactants. *J. Org. Chem.* (2000) 65: 8197- 8203, DOI: 10.1021/jo0006425.
48. Patial, P., Shaheen, A. Ahmada, I.: Synthesis, Surface active and thermal properties of novel imidazolium cationic monomeric surfactants, *J. of Industrial and Engineering Chemistry* 20 (2014) 4267-4275, Dio: 10.1016 / J.Jiec.2014.01.032.
49. Ismail, A.A., Abdelfatah, M. B., Mohammed, M., Abdallah, A.E., Ahmed, I.A.: Synthesis and Biocidal Activity Of Some Naphthalene- Based Cationic Surfactants, *J Surfact Deterg.* 15 (2012) 223-234, Doi: 10.100/ % 2Fs11743-01-286-z.
50. Cronin, M.T., Aptula, A.O., Dearden, J.C., Duffy, J.C., Ntezeva, T.I., Patel, H.P, Philip, H.R., Wayne, S.T., Andrew P. W., Kon-Stantinos, V., Gerrit, S.: Structure –based classification of antibacterial activity, *J. Chem.Inf. Comput.Sci.*42 (2002) 869-878, DOI: 10.1021 / ci 025501d.
51. Stephen, WM, Donald, JV: The relationship between the interfacial properties of surfactants and their toxicity to aquatic organisms. *Environ Sci Technol* 35 (2001) 954-959, DOI: 10.1021 es0015141.
52. Aiad, I., El-Sukkary, M.M., El-Deeb, A. et al., Surface Properties, Thermodynamic Aspects and

- Antimicrobial Activity of Some Novel Iminium Surfactants, *J Surfact Deterg* (2012) 15: 359, DOI:10.1007/s11743-011-1317-9\.
53. Augusta S., H.F. Gruber, F. Streichsbier, Synthesis and antibacterial activity of immobilized quaternary ammonium salts *J. Appl. Polym. Sci.* 53 (9) 1149–1163(1994), DOI: 10.1002/app.1994.070530903.
 54. Negm N, S.M. Tawfik, Isoxazolium cationic schiff base surfactants surface and antimicrobial properties, *Chimica Oggi/Chemistry Today* 30 (6) 5–8(2012), DOI.org/10.1007/s11164-013-1492-6.
 55. Badawi A.M., M.A. Mekawi, A.S. Mohamed, M.Z. Mohamed, M.M. Khowdairy, Surface and Biological Activity of Some Novel Cationic Surfactants, *J. Surfactants Deterg.* 10 (4) 243–255(2007), DOI: 10.1007/s11743-007-1040-8.
 56. A. Cukurovali, I. Yilmaz, S. Gur, C. Kazaz, *Eur. J. Medic Chem.* 41 (2006) 201–207, DOI: 10.1016/j.ejmech.2005.01.013.
 57. Koch A., *Clin. Microbiol. Rev.* 16 (4) (2003) doi: 10.1128/CMR.16.4.673-687.2003, 673–68, DOI: 10.1128/CMR.16.4.673-687.2003
 58. Negm N.A., Ghuiba F.M., Mahmoud S.A., Tawfik S.M., Biocidal and anti-corrosive activities of benzoimidazol-3-ium cationic Schiff base surfactants, *Eng. Life Sci.* 11 (2011) 496–510, DOI: 10.1002/elsc.201000106.
 59. Salah M. Tawfik, Synthesis, surface, biological activity and mixed micellar phase properties of some biodegradable gemini cationic surfactants containing oxycarbonyl groups in the lipophilic part, *J. Ind. Eng. Chem.* 28 (2015) 171–183, DOI.org/10.1016/j.jiec.2015.02.011.
 60. Zaki M.F. and Tawfik S.M, Synthesis, surface properties and antimicrobial activity of some germanium nonionic surfactants, *J. Oleo Sci.* 63 (2014) 921–931, DOI: 10.5650/jos.ess14052.
 61. Zaki M.F., Aiad I.A. and Tawfik S.M., Synthesis, characterization, surface and biocidal effect of some germinate nonionic surfactants, *J. Ind. Eng. Chem.* 21 (2015), 1174–1182, DOI: 10.1016/j.jiec.2014.05.031 .
 62. Tawfik S.M, Sayed A., Aiad I., Corrosion Inhibition by Some Cationic Surfactants in Oil Fields, *J. Surfactants Deterg.* 15 (2012), pp. 577–558 DOI:10.1007/s11743-012-1339-y.
 63. Nihal O. Shaker; Fatma H. Abd El-Salam; Bahyia M.El-Sadek; Eman M. Kandeel and Sharbat A. Baker Anionic Schiff Base Amphiphiles: Synthesis, Surface, Biocidal and Antitumor Activities, *Journal of American Science*; 7(5) (2011) DOI:10.7537/marsjas070511.57.
 64. Slowing I I, Vivero-Escoto J L, Trewyn B G and Lin V S Y, Mesoporous silica nanoparticles: structural design and applications *J. Mater. Chem.* 20 7924–37, (2010), DOI: 10.1039/C0JM00554A.
 65. Ambrogio M W, Thomas C R, Zhao Y-L, Zink J I and Stoddart J F Mechanized silica nanoparticles: a new frontier in theranostic nanomedicine *Acc. Chem. Res.* 44 903–13, (2011), DOI: 10.1021/ar200018x.
 66. Ariga K, Vinu A, Yamauchi Y, Ji Q and Hill J P, Nanoarchitectonics for mesoporous materials *Bull. Chem. Soc. Japan* 85 1–32, (2012), DOI:10.1246/bcsj.20110162. Li Z, Barnes J C, Bosoy A, Stoddart J F and Zink J I Mesoporous silica nanoparticles in biomedical applications *Chem. Soc. Rev.* 41 2590–605, (2012) Doi: 10.1246/bcsj.20110162.
 67. Tarn D, Ashley C E, Xue M, Carnes E C, Zink J I and Brinker C J, Mesoporous silica nanoparticle nanocarriers: biofunctionality and biocompatibility, *Acc. Chem. Res.* 46 792–801, (2013), DOI: 10.1021/ar3000986.