

## The effect of topical application of Strontium Ranelate on induced bone defect's healing in Guinea pigs

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**Abstract:** This study was done to evaluate histological effect of topical application of Strontium Ranelate on induced bone defect's healing in Guinea pigs. Twelve male guinea pigs were used in this study and randomly divided into three equal groups based on their time of scarification at 2, 4 and 6 weeks post surgically. The animals were subjected to implantation of strontium ranelate (Servier Industries, France) in mandibular surgically induced bony cavity in the right side. Moreover, a second bony cavity of the same size was induced on the left side without application of (SrRan) to serve as control. All the specimens were examined for evaluation of the healing process of the jaw defects. The control histological sections revealed that healing of the bony cavities proceeded in a normal sequence of bone healing starting from 2 weeks to 6 weeks. There was gradual increase in the formation of bone trabeculae and decrease in the amount of granulation tissue. In the study groups, there was slower formation of bone than in the control group. It was proved that healing was slower or delay with SrRan.

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### 1. Introduction

Filling of bone defect with normal bone is representing a challenge in dentistry. Local loss of bone tissue can be derived from several causes such as dental extraction, fractures, infection and bone metastasis. Several factor can affect the filling of bone defect with bone which include presence of concomitant diseases, the individual age, nutrition and status of hormone.<sup>(1)</sup>

Bone defect healing is multiphase process start with phase of bleeding and inflammation then to the second phase which is the formation of the woven bone which undergo remodeling by osteoclast followed by the replacing it by the lamellar bon by osteoblast.<sup>(2)</sup>

Bone Grafting is highly recommended in case of extensive bone loss especially in individual with systemic bone disease as osteoporosis to insure well being and life quality, traditional approaches, such as bone graft and the insertion of scaffolds of osteoinductive biomaterials might have an advantageous alternatives which is the local or systemic administration of anabolic or anticatabolic drugs, growth factors and mesenchymal stem cells.<sup>(3-6)</sup>

Promoting of bone defect healing in osteoporotic patient by the anabolic or anticatabolic drugs administration commonly used to increase bone formation and inhibit bone resorption, Strontium

ranelate (SrRan), has the double benefits so it attract the interest in investigation looking for improvement of bone defect healing In several study on animal model has been reported the dual effect of SR, where it has been approved that SR prevent bone loss by increase bone formation and inhibit bone resorption.<sup>(7-11)</sup>

SrRan has been shown to have dual effect on the bone by reducing bone resorption by inhibition of osteoclasts and increasing bone formation by stimulation of osteoblasts,<sup>(12-14)</sup> SrRan stimulates the proliferation and differentiation of osteoplastic cells and inhibits the activity and differentiation of osteoclasts, thereby enhancing matrix deposition and ultimately new bone formation.<sup>(15-22)</sup>

Strontium ranelate is shown a distinct advantage compared to bisphosphonates as its treatment not associated with osteonecrosis of the jaw (ONJ).<sup>(23)</sup>

Recent studies on healthy animals have reported that systemic oral administration of SrRan improves the bone volume and strength.<sup>(24, 25)</sup> however the systemic treatment with SrRan might show the occurrence of some side effects such as headache, nausea, diarrhea and in rare cases, cutaneous hypersensitivity.<sup>(26)</sup>

Developing of systems for local administration and topical application of strontium to overcome the

adverse systemic effects and achieve acceleration and increasing of bone healing by The positive effects of SrRan on bone homeostasis may be an important indicator of success in relation to decreasing the healing periods associated with bone defect.<sup>(27)</sup>

## 2. Materials and methods

Twelve adult guinea pigs with an average weight of 350-500 gm. were used in this study. All animals were kept under the same nutritional and environmental conditions. Each animal was subjected to implantation of strontium ranelate (SrRan) granules (Servier Industries, France) in surgically induced bony cavity in the right side of the submental area of the mandible. Moreover, a second bony cavity of the same size was induced on the left side without application of (SrRan) granules to serve as control.

After operation the animals were transferred to their cages for daily observation till complete recovery.

At 2, 4, and 6 weeks, the animals of all groups were sacrificed and divided equally among groups A, B, and C respectively.

After sacrificing the animals, their mandibles were separated for preparation of paraffin blocks and histological evaluation.

## 3. Histological results

### The control group (left Cavity):

The histological sections revealed that healing of the bony cavities proceeded in a normal sequence of bone healing starting from 2 weeks and completed at 6 weeks.

At 2 weeks postsurgically the bony cavities were found to be filled with granulation tissue and newly formed bony trabeculae. From 2 to 6 weeks there was gradual increase in number and size of the newly formed bony trabeculae with widening of narrow spaces, and appearance of resting lines in between bone lamellae. The amount of granulation tissue gradually decreased.

### The study group (rightcavity):

The results of the present study at 2 weeks showed a cavity wound with an empty space and a condensation of granulation tissue at the cavity opening. At 4 weeks the experimental sections informed that bridge of bone at wound opening, granulation tissue seen at cavity center. At 6 weeks the experimental sections showed appearance of heavy condensation of granulation tissue with thin bone trabecule could be seen.

## 4. Discussion

Bone tissue regeneration and bone defect healing still represent a significant challenge in the dentistry especially with large cavity that exceeds the critical

size which cannot heal normally by regeneration. Recent studies tried to enhance the bone formation and overcome the limitation of bone graft material by using anabolic agent that accelerate bone healing.<sup>(28)</sup>

Strontium (SR) in human biology and pathology has less attention than calcium and magnesium, and for many years it did not have clinical interest. Although this is still true, there is an growing knowledge of the biological effect of SR after the releasing of the drug strontium ranelate, which has recently been reported to decrease risk of fractures in osteoporotic patients.<sup>(19)</sup>

At the last decade, several studies have revealed that strontium as a stable element shows positive effects on bone formation and resorption and enhances trabecular bone mass when administrated in animals, without inducing deleterious effect on bone mineralization.<sup>(29, 30)</sup>

There are growing evidences that strontium controls bone remodeling by affecting both bone formation and bone resorption. In vitro, strontium reduces bone resorption,<sup>(8)</sup> and induces bone formation,<sup>(31)</sup> In vivo, low concentration strontium administration decreases bone resorption,<sup>(32)</sup> and increases bone formation, as evaluated by bone histomorphometry in<sup>(33, 34)</sup> osteoporotic patients.<sup>(18)</sup>

The great effect of SrRan on bone fracture healing according to several studies made by **Aslam et al**,<sup>(35)</sup> **Neves et al**,<sup>(36)</sup> and **Tan et al**,<sup>(37)</sup> who has approved the benefacial effect of SrRan in promoting the bone healing. Not only promoting effect but SrRan has a dual effect on bone inducing bone formation and reducing of bone resorption.<sup>(38, 39)</sup>

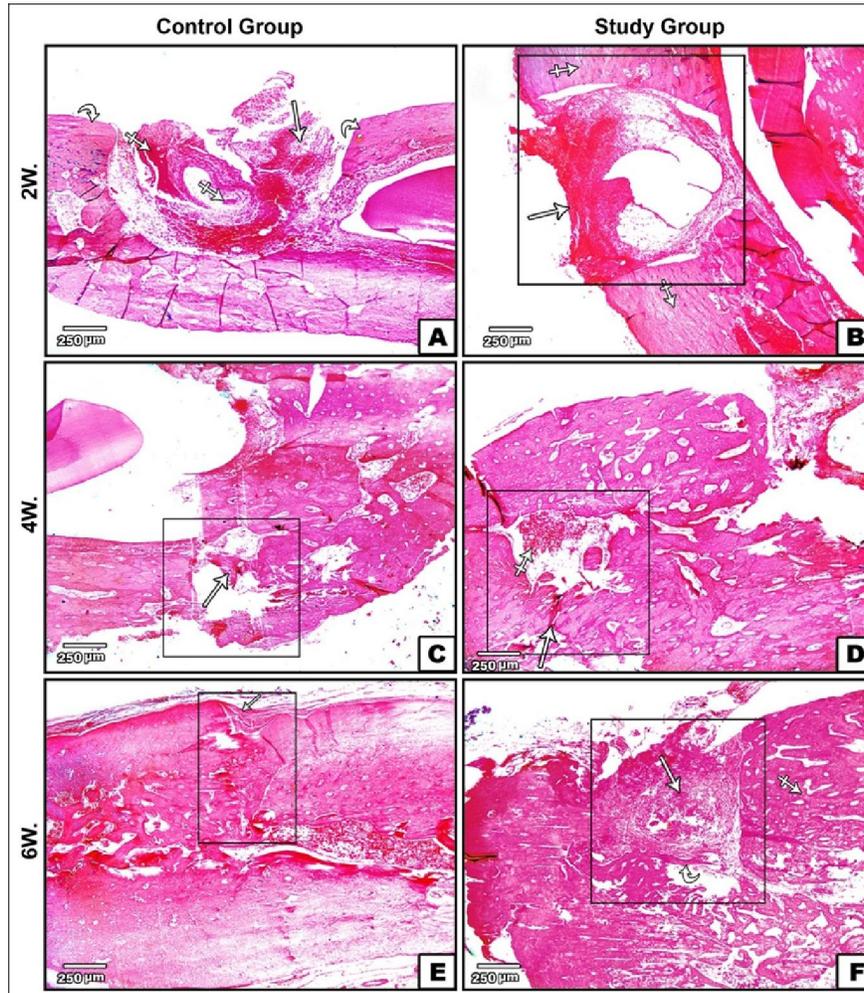
Although, the previously mentioned marvelous effect of SrRan on bone healing but there are rapidly growing evidence about the side effect of systemic administration of the SrRan which include:<sup>(40, 41)</sup> venous thromboembolism<sup>(42)</sup> gastrointestinal disturbance,<sup>(43)</sup> DRESS<sup>(44, 45)</sup> and Rarely memory loss.<sup>(46)</sup>

To overcome the systemic side effect several studies has been demonstrated to use local delivery system in order to achieve maximum benefit of SrRan without its systemic side effect,<sup>(47)</sup> **Tian et al**, studied the effect of titanium implant coated with SR on osseointegration which reported that SR enhances the osseointegration between bone and implant interface.<sup>(48, 49)</sup> also **Baier et al**, conclude that strontium based calcium phosphate cement accelerate the implant osseointegration in osteoprotic patient,<sup>(50)</sup> In present study in order to overcame systemic drawbacks and to make topical application is more easier and simple we used SrRan granules which may be more control in application.

In dentistry, the study of pharmaceutical drugs that flavor bone healing and regeneration is a vital

importance,<sup>(51)</sup> in the present study the dual effect of strontium ranelate on bone healing was evaluated on

bone defect in normal guinea pigs mandible.



**A:** The wound cavity filled with granulation tissue (arrow) interspersed with small fragment of new bone pieces (crossed arrows) border of old bone (curved arrows). **B:** cavity wound with an empty space and a condensation of granulation tissue at cavity opening (arrow) old mature bone (crossed arrow) **C:** Partial closure of the cavity with bone trabeculae (arrow). **D:** Bridge of bone at wound opening (arrow) granulation tissue seen at cavity center (crossed arrow). **E:** complete obliteration of the wound cavity arrow refers to opening of the wound. **F:** heavy condensation of granulation tissue (arrow) with thin bone trabeculae (curved arrow).

In the present study we use the guinea pigs to serve as animal models due to the similarity in the histological and hematological reaction in evaluation of bone formation according to **Skoloudik's** study who chosen guinea pigs to evaluate effect of transplanted mesenchymal stromal cells in the treatment of postoperative temporal bone defect,<sup>(52)</sup> also **Jang et al**, used guinea pigs to assess the role of bioactive glass as a bone graft materials.<sup>(53)</sup>

Due to the clinical importance of critical size defect in testing of biomaterials, Standardized 5mm bone defects were created bilateral to the midline at submental area in this study to see the difference in

the bone defect repair between the groups received the strontium ranelate and control groups.<sup>(54, 55)</sup>

According the study done by **Passeri et al**, which reported that bone healing includes fibrous connective tissue and the immature bone and so it was easily to evaluate the effect of strontium ranelate on healing process so we made histological evaluation after 2, 4 and 6 weeks as the best tissue quality could be evaluated within the shortest time period assuming that the bone will be remodeled throughout animal life.<sup>(56)</sup>

In this study, the histological findings on control group A, normal bone healing sequence was observed.

Starting with condensed granulation tissue filling the cavity with discrete areas of new bone formation in the osseous defect. These results are in accordance with the study done by **Tawfik and El-Hawary** in the detection of effect of OSTEON II in healing of induced bony cavity in guinea pigs, they noticed formation of wide area of granulation tissue in their control group after 2 weeks.<sup>(57)</sup>

On the other hand, the study group A showed a cavity wound with an empty space and a condensation of granulation tissue at the cavity opening. These results correlate with previous study made by **Basu et al**, who reported that this was no significance difference between SrRan treated and non SrRan treated groups when he evaluate the formation of bone tubercular bone inside the cavity after 2 week.<sup>(58)</sup>

In contradict with the **Lourenço et al**, who studied the effect of different injectable hybrid system for strontium local delivery in enhancement of bone repair in rat critical size defect, his study reported that there are major significance difference between the histological evaluation as seen higher cell invasion was observed at the sr-hybrid group than the other groups at 15 days.<sup>(59)</sup>

After 4 week, the histological findings of control group B reported partial closure of the cavity wound with bone trabecule. This agrees with **Louis's et al**, study while he was studying the effect of non-selective and selective cyclooxygenase-2 non-steroidal anti-inflammatory drugs, he reported that bone healing is highly significant in control group in comparison to study group.<sup>(60)</sup>

According to **Tian et al**, who study the effect of porous strontium-doped calcium polyphosphate scaffolds as a bone substitute, they noticed that after 4 weeks there was highly significant difference with strontium group due to active bone formation, which was evident by large number of osteoblasts and osteocytes.<sup>(61)</sup>

Also the results of the study made by **li et al**, which reported that filling of the fracture gaps and endochondral ossification lead to bone regeneration showed greater and denser newly forming trabeculae than the nontreated OVX animals.<sup>(62)</sup> in contradict to this result the histological outcome of study group B informed that bone bridge formation at wound opening and granulation tissue seen at cavity center, this result is agreed with **Cebesoy et al**, who studied effect of strontium ranelate on fracture healing in rat tibia that reported no significant differences in callus formation or bone union could be demonstrated between the strontium group and the control group radiologically while in histopathology it was seen the fractures healed normally in both group with no significant difference during the first 4 weeks after the fracture.<sup>(63)</sup>

After 6 week, the histological finding on control group C showed complete obliteration of the wound cavity with mature bone trabecule. However, the histological results of study subgroup C showed showing heavy condensation of granulation tissue this result is agreed with study made by **Mohamed et al**, who found that the treatment with strontium ranelate may interfere with the mineralization process as through the tasted blood parameter showed no significant difference in serum level of calcium and phosphate at 1,3,6 weeks while there were high statistical significance difference of serum alkaline phosphatase and osteocalcin between treated group with strontium and control group with dropping lower of serum ALP so he conclude that strontium ranelate has slow action onest on the bone marker and also that strontium ranelate might delay bone formation during the early stage of fracture healing with bone gap.<sup>(54)</sup>

In contrast to our study result, **Thormann et al**, studied the effect the strontium modified calcium phosphate bone cement on bone formation, he reported that there was higher significance difference in bone formation between experimental and control group.<sup>(64)</sup>

## Conclusion

Healing of the induced jaw osseus defects in guinea pigs was delayed with topical application of Strontium Ranelate

## References

1. Reichert JC, Saifzadeh S, Wullschlegel ME, Epari DR, Schütz MA, Duda GN, et al. The challenge of establishing preclinical models for segmental bone defect research. *Biomaterials*. 2009;30(12):2149-63.
2. Gerstenfeld LC, Cullinane DM, Barnes GL, Graves DT, Einhorn TA. Fracture healing as a post-natal developmental process: Molecular, spatial, and temporal aspects of its regulation. *Journal of cellular biochemistry*. 2003;88(5):873-84.
3. Lutolf MP, Weber FE, Schmoekel HG, Schense JC, Kohler T, Müller R, et al. Repair of bone defects using synthetic mimetics of collagenous extracellular matrices. *Nature biotechnology*. 2003;21(5):513-8.
4. Bruder SP, Kurth AA, Shea M, Hayes WC, Jaiswal N, Kadiyala S. Bone regeneration by implantation of purified, culture-expanded human mesenchymal stem cells. *Journal of Orthopaedic Research*. 1998;16(2):155-62.
5. Holstein J, Orth M, Scheuer C, Tami A, Becker S, Garcia P, et al. Erythropoietin stimulates bone formation, cell proliferation, and angiogenesis in

- a femoral segmental defect model in mice. *Bone*. 2011;49(5):1037-45.
6. Yang Y, Hallgrímsson B, Putnins EE. Craniofacial defect regeneration using engineered bone marrow mesenchymal stromal cells. *Journal of Biomedical Materials Research Part A*. 2011;99(1):74-85.
  7. Ammann P, Shen V, Robin B, Mauras Y, Bonjour JP, Rizzoli R. Strontium ranelate improves bone resistance by increasing bone mass and improving architecture in intact female rats. *Journal of bone and mineral research*. 2004;19(12):2012-20.
  8. Buehler J, Chappuis P, Saffar J, Tsouderos Y, Vignery A. Strontium ranelate inhibits bone resorption while maintaining bone formation in alveolar bone in monkeys (*Macaca fascicularis*). *Bone*. 2001;29(2):176-9.
  9. Delannoy P, Bazot D, Marie P. Long-term treatment with strontium ranelate increases vertebral bone mass without deleterious effect in mice. *Metabolism*. 2002;51(7):906-11.
  10. Hott M, Deloffre P, Tsouderos Y, Marie P. S12911-2 reduces bone loss induced by short-term immobilization in rats. *Bone*. 2003;33(1):115-23.
  11. Marie PJ, Hott M, Modrowski D, De Pollak C, Guillemain J, Deloffre P, et al. An uncoupling agent containing strontium prevents bone loss by depressing bone resorption and maintaining bone formation in estrogen - deficient rats. *Journal of bone and mineral research*. 2005;20(6):1065-74.
  12. Baron R, Tsouderos Y. In vitro effects of S12911-2 on osteoclast function and bone marrow macrophage differentiation. *European journal of pharmacology*. 2002;450(1):11-7.
  13. Canalis E, Hott M, Deloffre P, Tsouderos Y, Marie P. The divalent strontium salt S12911 enhances bone cell replication and bone formation in vitro. *Bone*. 1996;18(6):517-23.
  14. Takahashi N, Sasaki T, Tsouderos Y, Suda T. S 12911 - 2 inhibits osteoclastic bone resorption in vitro. *Journal of bone and mineral research*. 2003;18(6):1082-7.
  15. Barbara A, Delannoy P, Denis B, Marie P. Normal matrix mineralization induced by strontium ranelate in MC3T3-E1 osteogenic cells. *Metabolism*. 2004;53(4):532-7.
  16. Bonnelye E, Chabadel A, Saltel F, Jurdic P. Dual effect of strontium ranelate: stimulation of osteoblast differentiation and inhibition of osteoclast formation and resorption in vitro. *Bone*. 2008;42(1):129-38.
  17. Capuccini C, Torricelli P, Sima F, Boanini E, Ristoscu C, Bracci B, et al. Strontium-substituted hydroxyapatite coatings synthesized by pulsed-laser deposition: in vitro osteoblast and osteoclast response. *Acta Biomaterialia*. 2008;4(6):1885-93.
  18. Marie P, Garba M, Hott M, Miravet L. Effect of low doses of stable strontium on bone metabolism in rats. *Miner Electrolyte Metab*. 1985;11(1):5-13.
  19. Meunier PJ, Roux C, Seeman E, Ortolani S, Badurski JE, Spector TD, et al. The effects of strontium ranelate on the risk of vertebral fracture in women with postmenopausal osteoporosis. *New England Journal of Medicine*. 2004;350(5):459-68.
  20. Ni G, Chiu K, Lu W, Wang Y, Zhang Y, Hao L, et al. Strontium-containing hydroxyapatite bioactive bone cement in revision hip arthroplasty. *Biomaterials*. 2006;27(24):4348-55.
  21. Qiu K, Zhao XJ, Wan CX, Zhao CS, Chen YW. Effect of strontium ions on the growth of ROS17/2.8 cells on porous calcium polyphosphate scaffolds. *Biomaterials*. 2006;27(8):1277-86.
  22. Sila-Asna M, Bunyaratvej A, Maeda S, Kitaguchi H, Bunyaratavej N. Osteoblast differentiation and bone formation gene expression in strontium-inducing bone marrow mesenchymal stem cell. *Kobe J Med Sci*. 2007;53(1-2):25-35.
  23. Lenart BA, Lorch DG, Lane JM. Atypical fractures of the femoral diaphysis in postmenopausal women taking alendronate. *New England Journal of Medicine*. 2008;358(12):1304-6.
  24. Maimoun L, Brennan TC, Badoud I, Dubois-Ferriere V, Rizzoli R, Ammann P. Strontium ranelate improves implant osseointegration. *Bone*. 2010;46(5):1436-41.
  25. Li Y, Li X, Song G, Chen K, Yin G, Hu J. Effects of strontium ranelate on osseointegration of titanium implant in osteoporotic rats. *Clinical oral implants research*. 2012;23(9):1038-44.
  26. Rizzoli R, Reginster J-Y. Adverse drug reactions to osteoporosis treatments. *Expert review of clinical pharmacology*. 2011;4(5):593-604.
  27. Shen Y, Liu W, Wen C, Pan H, Wang T, Darvell BW, et al. Bone regeneration: importance of local pH—strontium-doped borosilicate scaffold. *Journal of Materials Chemistry*. 2012;22(17):8662-70.
  28. Fernandez-Yague MA, Abbah SA, McNamara L, Zeugolis DI, Pandit A, Biggs MJ. Biomimetic approaches in bone tissue engineering: Integrating biological and physicomaterial strategies. *Advanced drug delivery reviews*. 2015;84:1-29.

29. Marie PJ, Ammann P, Boivin G, Rey C. Mechanisms of action and therapeutic potential of strontium in bone. *Calcified tissue international*. 2001;69(3):121-9.
30. Marie P. Strontium ranelate: a physiological approach for optimizing bone formation and resorption. *Bone*. 2006;38(2):10-4.
31. Canalis E, Hott M, Deloffre P, Tsouderos Y, Marie PJ. The divalent strontium salt S12911 enhances bone cell replication and bone formation in vitro. *Bone*. 1996;18(6):517-23.
32. Marie PJ, Hott M. Short-term effects of fluoride and strontium on bone formation and resorption in the mouse. *Metabolism*. 1986;35(6):547-51.
33. Grynepas M, Marie P. Effects of low doses of strontium on bone quality and quantity in rats. *Bone*. 1990;11(5):313-9.
34. Delmas PD, Vergnaud P, Arlot ME, Pastoureau P, Meunier PJ, Nilssen M. The anabolic effect of human PTH (1–34) on bone formation is blunted when bone resorption is inhibited by the bisphosphonate tiludronate—is activated resorption a prerequisite for the in vivo effect of PTH on formation in a remodeling system? *Bone*. 1995;16(6):603-10.
35. Aslam MZ, Khan MA, Chinoy M, Jillani S, Sultan S, Ahmed S. Significance of strontium ranelate in healing of surgically fixed tibial diaphyseal fractures treated with strontium ranelate vs placebo; a randomised double blind controlled trial. *JPMMA The Journal of the Pakistan Medical Association*. 2014;64(12 Suppl 2): S123-6.
36. Neves N, Linhares D, Costa G, Ribeiro C, Barbosa M. In vivo and clinical application of strontium-enriched biomaterials for bone regeneration. *Bone and Joint Research*. 2017;6(6):366-75.
37. Tan S, Zhang B, Zhu X, Ao P, Guo H, Yi W, et al. Deregulation of bone forming cells in bone diseases and anabolic effects of strontium-containing agents and biomaterials. *BioMed research international*. 2014;2014.
38. Zacchetti G, Dayer R, Rizzoli R, Ammann P. Systemic treatment with strontium ranelate accelerates the filling of a bone defect and improves the material level properties of the healing bone. *BioMed research international*. 2014;2014.
39. Taylor BA, Bezuhyly M, Brace M, Carter M, Hong P. Effect of strontium citrate on bone consolidation during mandibular distraction osteogenesis. *The Laryngoscope*. 2017.
40. Grosso A, Douglas I, Hingorani A, MacAllister R, Smeeth L. Post - marketing assessment of the safety of strontium ranelate; a novel case - only approach to the early detection of adverse drug reactions. *British journal of clinical pharmacology*. 2008;66(5):689-94.
41. Bolland MJ, Grey A. Ten years too long: strontium ranelate, cardiac events, and the European Medicines Agency. *BMJ: British Medical Journal (Online)*. 2016;354.
42. Atteritano M, Catalano A, Santoro D, Lasco A, Benvenga S. Effects of strontium ranelate on markers of cardiovascular risk in postmenopausal osteoporotic women. *Endocrine*. 2016;53(1):305-12.
43. Reid IR. Efficacy, effectiveness and side effects of medications used to prevent fractures. *Journal of internal medicine*. 2015;277(6):690-706.
44. Yang C-Y, Chen C-H, Wang H-Y, Hsiao H-L, Hsiao Y-H, Chung W-H. Strontium ranelate related Stevens–Johnson syndrome: a case report. *Osteoporosis International*. 2014;25(6):1813-6.
45. Cacoub P, Descamps V, Meyer O, Speirs C, Belissa-Mathiot P, Musette P. Drug rash with eosinophilia and systemic symptoms (DRESS) in patients receiving strontium ranelate. *Osteoporosis International*. 2013;24(5):1751-7.
46. Gupta P, Gupta K. Correspondence Address. 2014.
47. Hahn GS. Topically administered strontium-containing complexes for treating pain, pruritis and inflammation. *Google Patents*; 2016.
48. Tian A, Zhai J-j, Peng Y, Zhang L, Teng M-h, Liao J, et al. Osteoblast Response to Titanium Surfaces Coated with Strontium Ranelate–Loaded Chitosan Film. *International Journal of Oral & Maxillofacial Implants*. 2014;29(6).
49. Kirschnack C, Wolf M, Reicheneder C, Wahlmann U, Proff P, Roemer P. Strontium ranelate improved tooth anchorage and reduced root resorption in orthodontic treatment of rats. *European journal of pharmacology*. 2014;744:67-75.
50. Baier M, Staudt P, Klein R, Sommer U, Wenz R, Grafe I, et al. Strontium enhances osseointegration of calcium phosphate cement: a histomorphometric pilot study in ovariectomized rats. *Journal of orthopaedic surgery and research*. 2013;8(1):16.
51. Jernberg GR. Delivery of agents and method for regeneration of periodontal tissues. *Google Patents*; 2000.
52. Skoloudik L, Chrobok V, Kalfert D, Koci Z, Sykova E, Chumak T, et al. Human Multipotent Mesenchymal Stromal Cells in the Treatment of Postoperative Temporal Bone Defect: An Animal Model. *Cell transplantation*. 2016;25(7):1405-14.

53. Jang CH, Cho YB, Bae CS. Evaluation of bioactive glass for mastoid obliteration: a guinea pig model. *in vivo*. 2007;21(4):651-5.
54. Ibrahim MRM, Singh S, Merican AM, Raghavendran HRB, Murali MR, Naveen SV, et al. The effect of strontium ranelate on the healing of a fractured ulna with bone gap in rabbit. *BMC veterinary research*. 2016;12(1):112.
55. Develioglu H, Saraydin SÜ, Dupoirieux L, Sahin ZD. Histological findings of long - term healing of the experimental defects by application of a synthetic biphasic ceramic in rats. *Journal of Biomedical Materials Research Part A*. 2007;80(2):505-8.
56. Passeri G, Cacchioli A, Ravanetti F, Galli C, Elezi E, Macaluso GM. Adhesion pattern and growth of primary human osteoblastic cells on five commercially available titanium surfaces. *Clinical oral implants research*. 2010;21(7):756-65.
57. Ihghaf NON, Tawfik MA-M, El-Hawary YM, Mansour NA. OSTEON II VERSUS BIOGEN IN HEALING OF JAW BONE DEFECTS. *DENTAL JOURNAL*. 2015;61(4045):4053.
58. Basu B, Sabareeswaran A, Shenoy S. Biocompatibility property of 100% strontium - substituted  $\text{SiO}_2\text{-Al}_2\text{O}_3\text{-P}_2\text{O}_5\text{-CaO-CaF}_2$  glass ceramics over 26 weeks implantation in rabbit model: Histology and micro - Computed Tomography analysis. *Journal of Biomedical Materials Research Part B: Applied Biomaterials*. 2015;103(6):1168-79.
59. Lourenço AH, Neves N, Ribeiro-Machado C, Sousa SR, Lamghari M, Barrias CC, et al. Injectable hybrid system for strontium local delivery promotes bone regeneration in a rat critical-sized defect model. *Scientific Reports*. 2017;7.
60. Gerstenfeld LC, Thiede M, Seibert K, Mielke C, Phippard D, Svagr B, et al. Differential inhibition of fracture healing by non - selective and cyclooxygenase - 2 selective non - steroidal anti - inflammatory drugs. *Journal of Orthopaedic Research*. 2003;21(4):670-5.
61. Tian M, Chen F, Song W, Song Y, Chen Y, Wan C, et al. In vivo study of porous strontium-doped calcium polyphosphate scaffolds for bone substitute applications. *Journal of Materials Science: Materials in Medicine*. 2009;20(7):1505-12.
62. Li Y, Luo E, Feng G, Zhu S, Li J, Hu J. Systemic treatment with strontium ranelate promotes tibial fracture healing in ovariectomized rats. *Osteoporosis International*. 2010;21(11):1889-97.
63. Cebesoy O, Tutar E, Kose KC, Baltaci Y, Bageci C. Effect of strontium ranelate on fracture healing in rat tibia. *Joint bone spine*. 2007;74(6):590-3.
64. Thormann U, Ray S, Sommer U, ElKhassawna T, Rehling T, Hundgeburth M, et al. Bone formation induced by strontium modified calcium phosphate cement in critical-size metaphyseal fracture defects in ovariectomized rats. *Biomaterials*. 2013;34(34):8589-98.

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