EFFECT OF VANADIUM TOXICITY IN CLARIAS LAZERA

Mona S. Zaki¹; Nevin E.Sharaf² and Mostafa H. Osfor³

¹Head of Department of Hydrobiology, National Research Center, Cairo, Egypt. ²Department of Environmental and Occupational Medicine, National, Research Center, Cairo, Egypt . ³Department of Nutrition, National, Research Center, Cairo, Egypt dr mona zaki@yahoo.co.uk

Abstract: The effect of dietary carbohydrates and vanadium toxicity on haematalogical profile, blood chemistry and hormonal level was studied in fry *Clarias Lazera*. Fry fish were divided into 3 groups (n=10) and exposed to different doses of vanadium sulfated and carbohydrate. Group1 was served as control, group 2 was fed with carbohydrate and vanadium sulfate (10 mg/ Kg diet ration), group 3 was fed with carbohydrate and vanadium sulfate (15 mg/Kg diet ration). There is a significant decrease in hemoglobin and P.C.V in group (3). There is a significant increase in serum cortisol, cholestrol, AST, ALT, urea, creatinine and alkaline phosphatase in group (3), also there is a significant decrease in serum phosphorous, sodium and potasium in treated fish. There is a significant high level of vanadium content in kidney muscles, heart and spleen in group (3) suggesting toxic effects of vanadium on cat fry fish *Clarious Lazara*. The total viable count of bacteria identified higher in fry fish fed on carbohydrate vandium. Predominate bacteria were identified as *Aeromonas, E. coli, Staph aureus. Pseudomonas, Fluorscences* and *Lacto bacilus* species. We emphasize the finding that increase in carbohydrate concentration causes harmful pathological effects which reduces humoral immune responses and enhances dietary vanadium toxicity. [Journal of American Science. 2010;6(12):291-296]. (ISSN: 1545-1003).

Keywords: Fry *Clarias Lazera*, Vanadium Pollution, Haematological, Biochemical, Clinicopathological, Bacterial count.

1. Introduction

Fry fish plays an important role, not only in human food diets but also in animal and poultry rations. It is a palatable and easily digested food which is rich in vitamins, calcium, phosphorus and iodine. In Egypt, fry fish is considered as a cheap food article if compared with other foods of animal origin. The flesh of healthy fry fish is considered as a marker for the natural aquatic environment.

Vanadium is a rare element found combined in certain minerals and used mainly to produce certain alloys. Most of the vanadium (about 80%) produced is used as ferrovanadium or as a steel additive. Mixed with aluminium in titanium alloys is used in jet engines and high speed air-frames, and steel alloys are used in axles, crankshafts, gears and other critical components. Vanadium oxide (V_2O_5) is used as a catalyst in manufacturing sulfuric acid and in making ceramics. It is added to glass to produce green or blue tint [1].

Vanadium was first discovered in 1971 as a trace element that is essential for normal growth. Since then, vanadium has been found to regulate the activity of various enzymes that induce pronounced changes in metabolic functions. Vanadium is never found unbound in nature, Vanadium occurs in carbon containing deposits such as crude oil, coal, oil shale and tar sands. Vanadium is abundant in most soils, in variable amounts, especially in areas where chemicals or petrochemicals complex were located, where these areas showed a significant increase in its concentration [2].

Humans may be exposed to excessive vanadium in several situations for example, overconsumption of vanadium-rich foods (e.g: seafood) [3], ingestion of certain dietary regimens specially that of body building, or inhalation of vanadium-rich environmental pollutants in certain occupations including boilermakers and power plant workers, who are often exposed to high levels of vanadiumrich compounds at work.

Because vanadium is vasoactive, individuals exposed to excessive vanadium may develop adverse vascular effects [4] especially pulmonary vascular diseases [5] as well as nanoparticulate of vanadium oxide potentiated vanadium toxicity in human lung cells [6] and Nickel and vanadium rich pollutant dust could be responsible for the respiratory problems reported [7]. Chronic exposure to vanadium pentoxide dust and fumes may cause severe irritation of the eyes, skin, upper respiratory tract, persistent inflammations of the trachea and bronchi, pulmonary edema, and systemic poisoning. Signs and symptoms of overexposure include; conjunctivitis, nasopharyngitis, cough, labored breathing, rapid heart beat, lung changes, chronic bronchitis, skin pallor, greenish-black tongue and an allergic skin rash [1] and [7]

In animals, vanadium causes the inhibition of certain enzymes, which has several neurological effects. Next to the neurological effects vanadium can cause breathing disorders, paralyses and negative effects on the liver and kidneys. Laboratory tests with test animals have shown, that vanadium can cause harm to the reproductive system of male animals, and that it accumulates in the female placenta. Vanadium can be found in fry fishes and many other species. In mussels and crabs vanadium strongly bioaccumulates, which can lead to concentrations of about 10^5 to 10^6 times greater than the concentrations that are found in seawater [8].

In recent years, much attention had been paid to the possible danger of metals poisoning in human as a result of consumption of contaminated fry fishes. So, the present study was carried out to elucidate the impact of vanadium on cat fry fish *Clarias Lazera*. Its haematological, biochemical and hormonal parameters were studied as well as the bacteriological and clinicopathological investigations.

2. Material and Methods

Experimental design

Thirty cat fry fish fry Clarious Lazera were used to assess the effect of vanadium sulfate. Fry fish weighing from 15-25g were obtained from Nile River and were kept in glass aquaria supplied with dechlorinated tap water at rate of one liter for each cm of fry fish's body. Fry fish were acclimated to the laboratory conditions for two weeks before the beginning of the experiment, they were fed with a commercial fry fish diet [9], the experiment was determined after 4 weeks. Fry fish were divided into 3 groups (n=10) and exposed to different doses of vanadium sulfated and carbohydrate. Group1 was served as control, group 2 was fed with carbohydrate and vanadium sulfate (10 mg/ Kg diet ration), group 3 was fed with carbohydrate and vanadium sulfate (15 mg/Kg diet ration). Mean of the initial body weight of each examined fry fish was at the beginning of the experiment then after 2-4 weeks of exposure.

Blood samples

Blood samples were collected from the caudal vein after 4 weeks of exposure. Each sample was divided into two parts the first one was heparinized for haematological investigations, while the second was centrifuged at 3000 rpm for 5 minutes to obtain serum for biochemical studies.

Hematological Analysis:

Haematological studies were performed according to Sandnes *et al* 1988 [10], where blood haemologlobin (Hb) and haematocrit (Ht) values were evaluated.

Biochemical Analysis:

The activities of alkaline phosphatase, aspartic aminotransferase (AST) and alanine aminotransferase (ALT) as well as cholesterol, urea and creatinine level were determined according to the method of Varley *et al.* [11] by using commercial kits (Bio Merieus, France).

Total serum protein was estimated according to Drupt [12]. Serum cortisol was analyzed by a Gamma counter using 125 Ι cortisol radioimmunassay Kit (Baxter Health Care Corporation USA) according to the method described by Pickering and Pottinger [13]. Potassium, Sodium and Phosphorous concentrations were determined by atomic absorption spectrophotometry [11].

Tissue analysis:

Liver, kidney and spleen samples were washed with distilled water then dried in hot air oven, sulphuric acid and hydrogen peroxide were added on samples then heated until the mixture became transparent after performing a wet ash digestion according to the method of Issac and Kerber [14].

Identification of bacteria:

The liver, kidney, spleen, muscle, stomach and gill from each examined fry fish were diluted immediately after sampling in sterile 0.9% saline and 0.1 ml volumes of appropriate dilutions and were spread over the surface of the typtic soy agar (oxoid). The plates were incubated at 22°C and inspected daily for up to 4 weeks.

The isolates were classified and identified according to Stevenson [15] and Quinn *et al.* [16].

The data were evaluated statistically according to Gad and Weil [17].

Water samples

Two water samples were collected from River Nile (Helwan) as well as two water samples free from any heavy metal pollution El-Kasr El-Eini (control) were analyzed for vanadium concentration by atomic absorption spectrophotometry.

3. Results and Discussion:

Data in table 1 showed that, the vanadium level in Helwan region was clearly higher than the maximum allowable concentration for human consumption as recommended internationally according to WHO (World Health Organization) . Nadal et, al.concluded that the occurrence of vanadium in nature and its use in various industrial processes has increased its inputs in the environment [2]. From the present study it is clear that the low vanadium levels were reported in water samples collected from areas far from industrial discharges, while high vanadium levels in the present study may be due to the collection of samples from areas subjected to industrial pollution.

In table 3 there is a significant decrease in body weight in group 3 (fry fish fed 15 mg vanadium for 4 weeks) than in group 1 (control) and group 2 (fry fish fed 10 mg vanadium), this results agree with that report [18].

The results present in table 6 showed the comparison of cholesterol levels between groups. The level was significantly increased in group 3 (fry fish fed on 15 mg vanadium) than in group 1(control). Hyperchrolestremia might be due to necrotic changes occurring in liver with liberation of cholesterol as a by-product of cell destruction. The present data suggest that impaired liver function lead to increased serum levels of alkaline phosphate, AST and ALT among group 3 (fry fish fed on 15 mg vanadium) and among group 2 (fry fish fed 10 mg vanadium) compared to group1 (control). In this concern Khalaf-Allah concluded that ALT and AST enzymes are good indices for the health status of liver parenchymatous, tissue necrosis is considered as the main source of AST and its increase in the serum of cat fry fish Clarias Lazera declared these necrotic changes [18]. In addition, exposure of fry fish to environmental pollutants might result in stimulation or depression of the enzyme activity depending on the concentration of pollutant and the duration of exposure [19] and [20].

Regarding the effect of vanadium on serum cortisol level in cat fry fish Clarias Lazera, highest level was obtained in group 3 (fry fish fed on 15 mg vanadium) then in group 2 (fry fish fed on 10 mg vanadium) as compared to that obtained in group 1 (control). The significant increase of cortisol level is probably due to the activation of hypothalamus pituitary internal axis [21].

From the data present in table 6, it is clear that elevation of vanadium level in the diets fed to Clarias

Lazera was positively correlated to hemoglobin (Hb) levels and haematocrit (Ht). A marked decrease in the Hb and Ht was recorded after feeding diet containing 15 mg and 10 mg vanadium, respectively. Reduced

reduced need for oxygen by change in blood pH. Moyle and Ceeh, Hall and Cliffs recorded, actived acetylcholinesterase of erythrocytes [22], [23]. Further more, Pickering and Dusten concluded that a consistent effect of cortisol was the reduction in the hemoglobin and iron levels as a result of decrease in appetite in rainbow trout fry fish or more likely to be the direct result of catablolic effect of cortisol in the fry fish tissues [24].

Hb may reflect metabolic adjustment according to

The mean phosphorus, sodium and potasium values in the serum of fry fish of group 3 (fry fish fed 15 mg vanadium) were significantly increased respectively than those recorded in the group 1 (control). This retention may be attributing to kidney dysfunction, whereas, the kidney is the normal pass for sodium and potassium. This kidney dysfunction may also explain the increase in serum urea and creatinine especially in group 3, but little known about the mechanisms involved in this association.

The results displayed also in table 6 showed that, there was general decrease in the mean total protein value in serum samples collected from the fry fish of group 3 and 2, respectively. The mean value of these parameters was lower than in group 1. Jagadeesh et al. estimated marked decrease in glycogen in tissues of fresh water fry fish after exposure to vanadium [25].

This experiment showed that the body weight of the examined fry fish was significantly decreased than the initial body weight after 4 weeks of exposure to 15 mg Vanadium. Also, Hilton and Better recorded a significantly reduced growth and increased mortality among the fry fish feeding diets of Vanadium (0,10,100,1000 or 10000 mg/kg) [26]. The increase in muscles and tissue lactic acid (2-12 fold) in association with decrease in pyruvic acid (72% in muscles +26% in liver) reflect a shift towards an anaerobic metabolism of fry fish following long term exposure to vanadium. [26]

Table 4 showed that, the bacterial isolates and counts were increased by feeding the fry fish with CHO and vanadium. The carbohydrates affect immunity and resistance to infection as recorded by Waagbo et al. [9]. Utility of vanadate mimetic protein phosphates inhibitors, to protect fry fish from microorganism [27]. The increase of bacterial count among the fry fish fed on vanadium may be related to the increased level of cortisol which decreases the host immunity.

In the course of experiment, a high concentration of vanadium levels has been found in

Clarias Lazera fed 15 mg vanadium table 5. This suggests that these organs could be useful as a marker for vanadium in the aquatic environment. In this concern, Ray et al. recorded a high concentration of vanadium in kidney, liver and other organs of cat fry fish as the concentration of vanadium in the tissues increased with its concentration in the aquatic environment and exposure time [28]. After exposure of fry fish to increased doses for 4 days, the vanadium content increase in the muscle then increased in all tissues [20], [25] and [26]. The capability of vanadium to be present in fry fish muscles is of particular interest in assessing the exposure of man to environmental vanadium as ingested by food.

Clinicopathological observations:

Abnormal swimming, lighting of the skin, scale loss and haemorrhages, were seen on the external body surface. In addition to congestion of gills, eyes, mouth, liver, kidney, spleen and intestine. This was noticed in fry fish exposed to vanadium sulphate 15 mg (group 3) but not in fry fish exposed to vanadium sulphate 10 mg (group 2)

In conclusion we emphasize that, the reported finding increase of carbohydrate concentrations causes harmful physiological effects, reduces humoral immune responses and enhances dietary vanadium toxicity.

Table 1: Vanadium concentration in water samples collected from two areas in Egypt.

Areas	No	Concentration of Vanadium p.p.m.
TT 1	1	1.04
Helwan	2	1.27
Al Kaan El Aini	3	0.154
Al- Kasr El- Aini	4	0.163

Table 2: Ingredients and Proximate composition of diets used in the experiment with vanadium supplementation.

Ingredient%	Diet, control	Diet 2	Diet 3
Fish meal	25	25	30
Meat and bone meal	5	5	10
Wheat bran	20	20	20
Skimmed milk	12	12	7
Yeast	10	10	15
Starch	-	10	15
Cod liver oil	2	2	2
Vitamin premix	1	1	1
Vanadium Mg	-	10	15
Crude protein%	40.35	35.95	38.89
Metabolizable energy k cal /kg	2205.4	2551.78	2315.4
Ether extract %	4.29	4.21	2.86
Crude fiber %	4.46	3.73	4.27
Ash %	5.65	6.26	10.25
Lysine %	2.13	1.88	2.29
Methionine %	0.62	0.55	0.613

Mineral and vitamin premix per/Kg of pellet food.

Vit A, 8000 g/u, vit D 900 g/u, vit E 2/u, vit K 4 mg, vit B2 3.6, niacin 20 mg., pyridoxine 0.2 mg, vit B125, Mn 70 mg, Sn 60 mg.

Table 3: Changes in body weight in cat fish fry (*Clarias Lazera*) fed on different levels of dietery carbohydrates in addition to vanadium sulphate.

Groups	Group 1	Group 2	Group 3
Intial body weight g	15±0.15	20±0.16	32±012
After 2 weeks g	18±0.45	30±0.23	38±0.7
After 4 weeks g	14±0.27	22±0.63	38±0.8*

P < 0.01

http://www.americanscience.org

No of examined fish 10 /group	Bacterial isolates	Site of isolation	Bacterial count
Group 3	-Aeromonas sp - E. coli	-Skin	$2 \text{ x} 10^3$
	- Staph Aureus	-Skin	
		- External surface,	$2x10^{2}$
	- E. coli	Stomach	
	- Aeromonas	- Gills	
	- lactobacillus	- Gills, stomach	1×10^{3}
		- Gills	
Group 2	-Enterbacter sp.	-Skin	$1 \ge 10^2$
-	-Pseudomonas	-Spleen, muscles	$2 \text{ x} 10^3$
	-Fluroscences	-Stomach	$1 \ge 10^{6}$
	-Lactobacillus	-Skin	

Table 4: Bacterial isolates recovered from the examined fry fish.

Table 5: The mean vanadium concentation in the organs of the fry fish mg/g net weight.

groups	muscles	spleen	heart	kidneys	Liver
Group 1	0.18±0.13	0.30±0.82	0.28±0.18	23.15±0.72	1.17±0.59
Group 2	0.17±0.24	0.20±0.70	0.13±0.20	4.0 0±0.83	2.10±0.60
Group 3	0.28±0.27*	$0.12 \pm 0.40 *$	0.17±0.12*	$6.00 \pm 0.74*$	5.21±0.15*
p < 0.01		* Significant			

Table 6: Some haematological, biochemical and hormonal parameters in cat fry fish *Clarias Lazera* fed on different levels of dietery carbohydrates in addition to vanadium sulphate.

Groups	Group 1	Group 2	Group 3
Parameters		-	-
Hemoglobin g/dl	7.5±0.20	7.52±0.14	6.10±0.12*
H.CT %	36.00±0.26	37.4±0.13	30.5±0.20*
Cortisol ng/ml	0.72±0.20	0.83 ± -0.08	1.30±0.66*
Phosphorous mg/dl	9.5±0.64	9.2±0.27	8.11±0.65*
Soduim M.E.Q	121±1.24	115±0.75	102±0.14*
Potasuim M.E.Q	7.0±0.82	7.03±0.44	6.4+/-0.74*
AlkPhosphatase U/L	21.5±3.2	22±0.60	27±0.70*
AST U/L	124±L40	131±0.85	144±0.25*
ALT U/L	22±0.15	24±0.70	37±0.20*
Cholestrol mg	144±0.25	149±0.13	170±0.54*
Total protein g/dl	9.20±0.75	9.01±0.80	8.01±0.62*
Urea mg/dl	3.3±0.68	3.3±0.78	4.8±0.20*
Creatinine mg/dl	0.77±03	0.73±0.75	0.92±0.42*

P < 0.01 * Significant

n = 30 Fish 10 / group.

References

1. Copyright © 1998-2007 Lenntech Water treatment & air purification HoldingB.V.Rotterdamseweg 402 M

2629 HH Delft, The Netherlands e-mail: info@lenntech.com

2. Nadal M., Schulmacher M., Domingo J.L. 2007: Levels of metals, PCB's PCN's and PAH's in soils of highly industrialized chemical/petrochemical area, chemosphere, 66, 267-76.

- Bu- Olayan A.H. and Al-Yakoob S. 1998: Lead, Nickel and Vanadium in sea food an exposure assessment for Kuwait consumers. Sci Environ. 2-3, 81-86.
- Huang Y.C., Ghio A.J. 2006: Vascular effects of ambient pollutant particles and metals, curr. Vasc. Pharmacol; 4(3): 199-203.
- Li Z., <u>Carter</u> J.D., <u>Dailey</u> L. A., <u>Huang</u> Y.C.T. 2004: 4-Vanadyl sulfate inhibits NO production via threonine phosphorylation of eNOS, Environ Health Perspect 112:201-206.
- Worle J.M., Kern K., Schelh C., Helmy A.C., Feldman C. and Krug H.F. 2007: Nanoparticulate vanadium oxide potentiated vanadium toxicity in human lung cells, Environ. Sci. Technol., 41,331-6.
- Figuero D.A., Rodriquez-Sierra C.J. and Jimenez-velez B.D. 2006: Toxicol & health, 22; 87-99
- Bu- Olayan A.H. and Subranmanyam M.N.V. 1996: Trace metal in fish from Kuwait coast using microwave acid digestion technique. Environ-Inter., vol. 22,753-758.
- 9. Waagbo R., Glette J., Sandnes K.and Hemre G.F., 1994: J. Fish Dis., 17, 245.
- Sandnes K., Lee Q., and Waagabo R., 1988: J. of Fish Biol., 32.129
- 11. Varley, H., Gwenbek, A. H. and Bell, M. 1980: Practical clinical chemistry, vol. I General I top's commener test, 5th ed. London, William medical books Ltd.
- 12. Drupt F. 1974: Pharm Biol, 9,77.
- 13. Pickering A.D. and Pottinger J. 1983: Gen Comp Ender., 49.232
- 14. Issac R.A. and kerber 1971: Amer Madison. 17
- Stevenson P. 1987: "Field Guide Systematic Bacteriology". University of Cuelphontaria, Canada pp. 280.

- Quinn PJ, Carter M.E., Markey B.K. and Carter G.R. 1994: "Clinical Veterinary Microbiology", Wolf Publishing Mosby, Year Book Europe limited.
- Gad S.C. and Weil C.S. 1986: Statistics for toxicologists. In, Hages A.W. (2nd ed.), "Principles and Methods of toxicology", Raven Press, New York, PP. 273-32.
- Khalaf-Allah 1998: Screening the effect of water pollution with some pesticides on the immune response in Oreochromis Niloticus fish. Vet. Mid. J. Giza; 46 (4-3); 883-393.
- Venberg F.G. and Venberg W.B. 1974: Pollutiion and Physiology of marine organisms. Academic Press. New York. pp. 59.
- Edel J. and E. Sabioni J. 1993: Accumulation and distribution of mussel myailus edulis and the gold fish carassius auratus. Sci.total-Environ. 133 (12), 139-151.
- Carballo M.J., M.J., Torroba, C.Munoz, D.V.Sanchez, J. D. Tarazona, and J.Dominguez 1992:J. Fish and Shell Fish Immunology. Z, 121
- Moyle P.B. and Cech J.J. 1982: Blood and Its Circulation in Fish-An Introduction to Ichthyology" (Ed. by, P.B. Moyle, J.H. Cech) pp-52-73 Prentice.
- 23. Hall I. and Cliffs E. 1982: Aspects and energy response in an Indian catfish Batrachus, Biometais. 11, 95-100.
- 24. Pickering A.P. and Duston J. J. Fish Biol., 23, 163, (1983).
- 25. Jagadeesh K.B., Shaffi S.A. and Jeelani S. 1989: Acta Physiologia Hungarica, 74, 43.
- 26. Hitton J.W.and Bettgeo W.G. 1988: Aquatitic Toxicology. 12,63.
- 27. Evans-Donald L. and Jaso. Friedmann L. 2001: Protection of Teleost fish. Biotclr, Nav. 15, 777.
- Ray D., Panerjeo S.K. and Chatterjee M. J.1999: Bioaccumulation of Nickel and Vanadium in tissues of Catfish Batracchus. J. Inorg-Biochem 38, 169-173.

6/22/2010