# Multiparametric Flow Cytometric Analysis of Fine Needle Aspirate of Enlarged Lymph Nodes: Validation with Histopathology

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Abstract: Objectives: The section of Hematopathology at King Fahd Specialist Hospital-Dammam (KFSHD) has expertise in diagnostic evaluation of patients with hematologic malignancy, interpretation of morphology of blood, bone marrow, and lymph node and other hematologic specimens as well as the application of ancillary techniques, especially flow cytometry (FCM). Using the four-color multiparameteric analysis, we are able to detect and immunophenotype malignant hematopoietic cells in lymph node aspirates allowing appropriate classification. We designed the current study to evaluate the validity of multiparameteric FCM as a diagnostic test for hematologic diseases in fine needle aspirated samples from patients with lymphadenopathies. Methods: We evaluated the validity of multiparametric FCM in diagnosing oncohematologic disease in 89 consecutive lymph node fine needle aspirate (FNA) specimens from patients with lymphadenopathy. All cases had excisional lymph node biopsy where histopathological evaluation and FCM when possible was performed for confirmation. Results: Flow cytometric diagnosis of non-Hodgkin's lymphomas on FNA specimens showed 100% correlation with the final histopathological interpretation. In addition, FCM enabled NHL sub-classification in all cases. The FCM interpretation was faster than histopathological examination, allowing quicker therapeutic decisions. FCM could not establish the diagnosis of our Hodgkin lymphoma cases since all these cases revealed unremarkable FCM features. Conclusion: Utilizing FCM is reliable and an accurate in the evaluation of lymphadenopathy in FNA material. We were able to validate our FCM technique in our laboratory for the evaluation of lymphoid malignancies.

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Key words: multiparametric flow cytometry, fine needle aspiration, lymphoma, lymphadenopathy

#### 1. Introduction:

Traditionally, the technique of choice for the diagnosis of lymph node pathology was based on histopathological studies of paraffin-embedded formalin-fixed biopsy specimens, where excisional biopsy of enlarged lymph nodes was considered the gold standard for the evaluation lymphoproliferative disorders (1, 2). Currently, the addition of immunohistochemistry for the detection of cell-specific antigens is essential in the classification of hematological lymphoid diseases (3). In the context of lymphoma diagnosis, immunohistochemistry (IHC) has the advantage over FCM that the cells of interest are identified morphologically and that these studies can be applied retrospectively on archived fixed tissue specimen (4).

However, immunophenotyping by immunohistochemical techniques has few limitations, which include subjectivity, limited reproducibility, prolonged turnaround time and the lack of consensus in quantifying antigen expression and defining positive and negative results (5, 6). Moreover, fixation might lead to loss of some cells and/or the cellular antigenicity, and there is reported difficulty in

demonstrating cytoplasmic immunoglobulin light expression (4,6).Because aforementioned limitations, immunophenotyping of lymphoid malignancies by FCM has become an essential diagnostic tool in the accurate classification of lymphoproliferative disorders. FCM offers many advantages. It is fast, qualitative as well as quantitative method in the evaluation of determining different cell antigens simultaneously using the multiparametric analysis (7). Meanwhile, FCM suffers few limitations such as variability in antigen signature of expression and cellular fragility during processing (8). In the current era of cost containment, fine needle aspiration (FNA) became a valid initial diagnostic tool for the evaluation of different masses including lymphadenopathy particularly when conventional biopsy is not feasible (9).

It is well-established that FNA as a diagnostic approach is reliable, associated with fewer complications, and more cost effective than excisional biopsy, especially when deeply seated lymph nodes are involved (10, 11). FNA became an extremely helpful technique to help distinguish lymphoma and metastatic malignancies from benign lymphoid

After obtaining an approval from the Internal

Review Board, prospective analysis of FCM

performed on 89 patients with lymphadenopathy was

carried out. The aspiration was performed by a

pathologist or a radiologist when localization by image

guidance was needed. The gauge of the needles used

ranges between 21-23 and the number of passes varied

from 2-4. Immediate adequacy evaluation of these

aspirates was performed on all cases utilizing Diff

proliferations (10-12). This option especially became very attractive when dealing with patients who are very sick or unstable enough to undergo surgical excisional biopsy procedures requiring general anesthesia (12).

However, there are few limitations for FNA as an initial diagnostic approach. These shortcomings are mostly related to sample adequacy and inability to evaluate architectural morphological Additional problems and complications with FNA are very rare and they may include bleeding, infections, and rarely pneumothorax (13). Nevertheless, the adequacy and complication of FNA procedures is operator dependent and improvements are achieved when they are performed by well-trained clinicians

Currently, many centers utilize the combined approach where the morphological cytopathological features in conjunction with the FCM findings to establish an accurate diagnosis on enlarged lymph nodes (2). While limited markers are routinely used on tissue sections with relative subjectivity, FCM allows a more precise definition of individual cell types. This is achieved, since cells of interest are identified by a combination of physical characteristics and the use of multiple antibodies directly conjugated fluorochromes (15). It also has the ability to assess monoclonality through detection of immunoglobulin light chain expression with an attractive speed (16).

## 2. Material and Methods:

Table	1: List of the antibodies up	sed in FCM pa	ınel.				
FITC	clone	PE	clone	PerCp	clone	APC	clone
IgG	X40	IgG	X40	CD45	2D1	IgG	X40
CD19	4G7	CD23	EBVCS-5	CD45	2D1	CD5	L17F12
CD7	M-T701	CD2	S5.2	CD45	2D1	CD5	L17F12
CD103	BER-ACT8	CD11c	S-HCL	CD45	2D1	CD25	2A3
CD34	8G12	CD33	P67.6	CD3		CD45	2D2
TdT	HT1,HT4,HT8,HT9	CD10	HI10a	CD45	2D1	CD34	8G12
Lambda	1-155-2	KAPPA	T28-2	CD45	2D1	CD19	SJ25C1
CD3	SK7	CD8	SK1	CD45	2D1	CD4	SK3

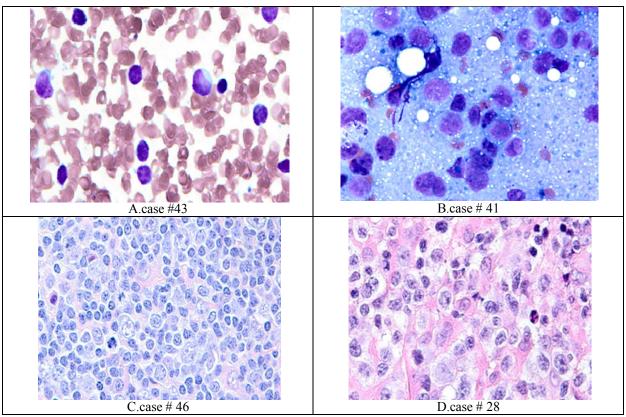
All specimens were tested for viability; samples with less than 60% viability were excluded and those with less than 85% viability were examined with caution. After examination of the forward and side scatter, cells within the lymphocyte gate were examined, excluding granulocytes and monocytes. All concomitantly cases underwent detailed the morphological cytological evaluation using Hematoxylin and Eosin stain on cell blocks. Standard cell block preparation was done using fixation in 10% neutral buffered formalin and embedding the tissue in paraffin. Routine IHC stains on cell block were done

Quick stain by pathologists on all cases. Material for FCM was collected into Roswell Park Memorial Institute (RPMI) medium for cell preservation and to the hematology &flowcytometry transfer laboratory. All samples were transported immediately at room temperature and stored at refrigerator (2-8°C) where testing was performed within 48 hours. They were transferred to a 5 ml tube and centrifuged for 2 minutes at approximately 400Xg. The supernatant was discarded and the cells were lysed in 2ml ammonium chloride solution then washed with 2 ml PBS-BSA. The cells were gently vortexed, incubated at room temperature for 10 minutes and then centrifuged again for 2 minutes. Leukocytes were resuspended in 1ml PBS and a basic panel of directly conjugated monoclonal antibodies was employed after capping with normal mouse IgG(Invitrogen Corporation, USA) and further extended according to the number of cells available (Table 1). The initial morphology of the preliminary slide is examined and patient's previous diagnosis if any. Samples were analyzed using the BD FACS-Canto II machine (Becton & Dickenson).

according to the morphology, also utilized for correlation with the flowcytometry.

### **Results:**

A total of 89 consecutive Fine Needle Aspiration (FNA)/ lymph node (LN) samples from 53 males and 36 females were collected with male to female ratio of 1.5:1. Their ages ranged from 4 to 76 years with a median of 30 and a mean of 34. All 89 FNA/LN samples were deemed adequate for FCM analysis and interpretation and all patients had a follow up excisional biopsy of the same lymph node within 1 month.



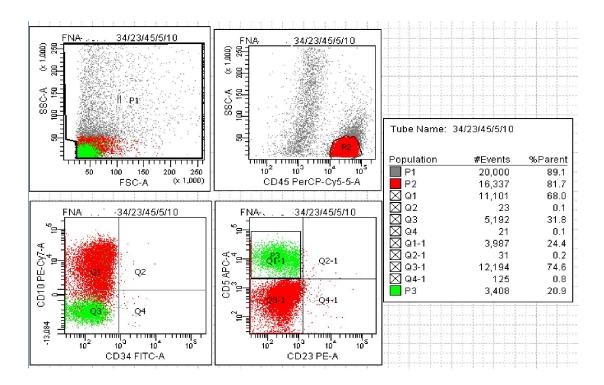
**Figure 1**: A: B-Non-Hodgkin's Lymphoma .B: Large B cell lymphoma with monoclonal lambda light chain restriction. C: Normal Lymph Node. D: Diffuse Large B-cell Lymphoma with Lambda light chain restriction

Table 2: List of reagents used & Clones used in histopathology

 Antibody
 Clone
 Company
 Antibody

Antibody	Cione	Company
CD45	RP2/18	Ventana, Tucson, USA
CD43	2B11 + PD7/26	Dako, Glostrup, Denmark
CD3	2GV6	Ventana, Tucson, USA
CD3	Poly	Dako, Glostrup, Denmark
CD4		Ventana, Tucson, USA
CD4	4B12	Dako, Glostrup, Denmark
CD2	MRQ-11	Ventana, Tucson, USA
CD2	AB75	Dako, Glostrup, Denmark
CD1a	EP3622	Ventana, Tucson, USA
СБТа		Dako, Glostrup, Denmark
CD7	SP94	Ventana, Tucson, USA
CD/	CBC.37	Dako, Glostrup, Denmark
CD8		Ventana, Tucson, USA
CD8	C8/144B	Dako, Glostrup, Denmark
CD19		Ventana, Tucson, USA
CD19	LE-CD19	Dako, Glostrup, Denmark
CD20	L26	Ventana, Tucson, USA
CD20	L26	Dako, Glostrup, Denmark
CD79a	SP18	Ventana, Tucson, USA
CD/9a	JCB117	Dako, Glostrup, Denmark
CD43	L60	Ventana, Tucson, USA
CD43	DF-T1	Dako, Glostrup, Denmark
CD23	SP23	Ventana, Tucson, USA
CD25		Dako, Glostrup, Denmark
CD15	MMA	Ventana, Tucson, USA
CDIS	Carb-3	Dako, Glostrup, Denmark
CD30	Ber-H2	Ventana, Tucson, USA
CD30	Ber-H2	Dako, Glostrup, Denmark
Ei	55k-2	Ventana, Tucson, USA
Fascin		Dako, Glostrup, Denmark

Antibody	Clone	Company
Pax5	SP34	Ventana, Tucson, USA
Paxs		Dako, Glostrup, Denmark
Ki-67	30-Sep	Ventana, Tucson, USA
	MIB-1	Dako, Glostrup, Denmark
CD138	B-A38	Ventana, Tucson, USA
CD138	MI15	Dako, Glostrup, Denmark
Vonno	Poly	Ventana, Tucson, USA
Kappa	Poly	Dako, Glostrup, Denmark
Lambda	Poly	Ventana, Tucson, USA
Lambua	Poly	Dako, Glostrup, Denmark
Cyclin D1	SP4-R	Ventana, Tucson, USA
Cyclili D1		Dako, Glostrup, Denmark
Alk1	ALK01	Ventana, Tucson, USA
AIKI		Dako, Glostrup, Denmark
Bcl-2	124	Ventana, Tucson, USA
DCI-2	124	Dako, Glostrup, Denmark
Bcl-6	GI191E/A8	Ventana, Tucson, USA
DCI-0	PG-B6p	Dako, Glostrup, Denmark
Cytokeratin	AE1/AE3 & PCK26	Ventana, Tucson, USA
Суюкетанн	AE1/AE3	Dako, Glostrup, Denmark
Cytokeratin	Cam 5.2	Ventana, Tucson, USA
Суюкстанн		Dako, Glostrup, Denmark
CD68	KP-1	Ventana, Tucson, USA
CD08	KP-1	Dako, Glostrup, Denmark
CD68		Ventana, Tucson, USA
CD06	PG-M1	Dako, Glostrup, Denmark
Epithelial Membrane	E29	Ventana, Tucson, USA
Antigen	E29	Dako, Glostrup, Denmark



**Figure2:** (Case 43)B-lymphoproliferative Disorder co-expressing CD19 (not shown) & CD10. These cells where negative for both kappa (not shown) & lambda (not shown)

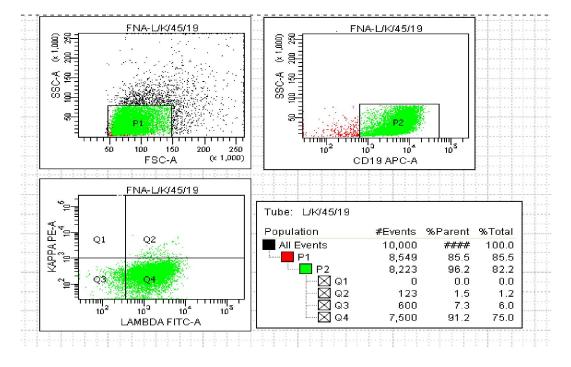


Figure 3: (Case 41) Large B cell Lymphoma Expressing CD19 & Lambda light chain restriction

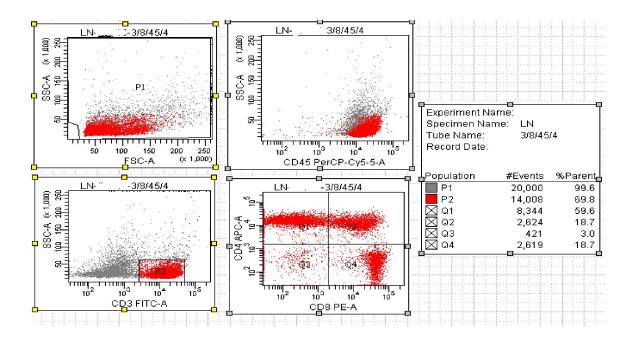


Figure 4: (Case 46) 16% of the population co-expressing CD4& CD8 suggesting normal peripheral cells.

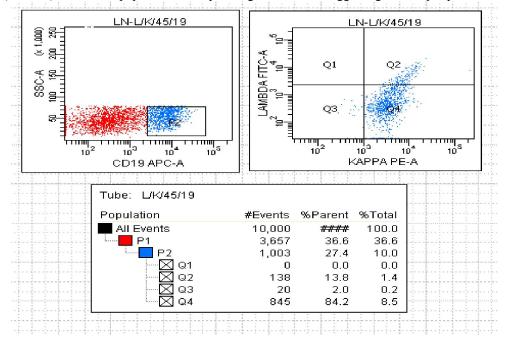


Figure 5: (Case 28) Diffuse Large B cell Lymphoma expressing CD19 & Kappa light chain restriction

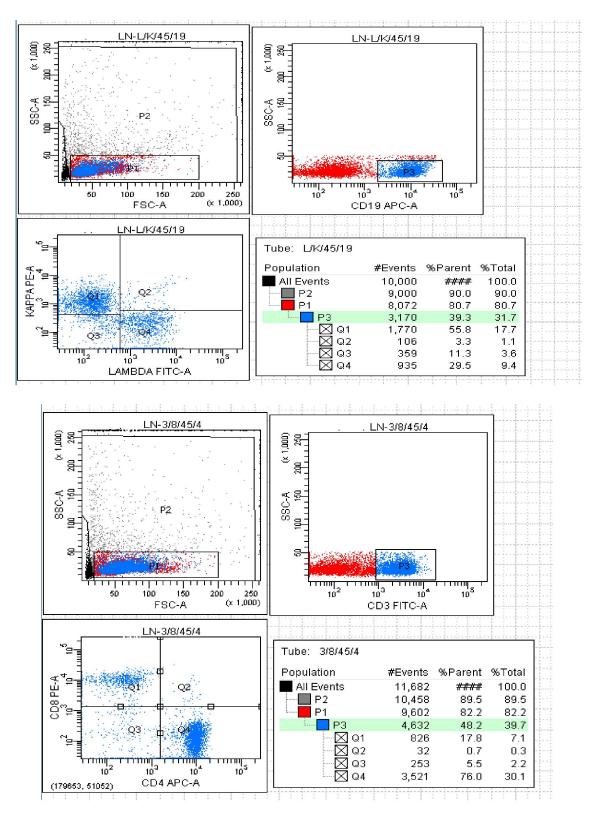


Figure 6: Normal Lymph node.

### 4, Discussion:

The results of this study confirm the reliability and validity of utilizing FNA/LN as an initial diagnostic tool where combining morphology and FCM analysis aids in reaching an adequate diagnosis (17). In all our cases, the technique provided an accurate diagnosis of non Hodgkin lymphomas (NHL) and the ability to differentiate benign from malignant lymphoid diseases. Similar to previous reports the accuracy of this approach provides an alternative diagnostic approach and saves patients unnecessary more invasive surgical biopsy intervention (17).

In the present study, combined FNA and FCM allowed an unequivocal distinction between non lymphomatouslymph node pathology(57/89) and NHL in all cases (32/89). FCM was thus 100% reliable in the diagnosis of reactive hyperplasia (Figures 1C, 4 & 6) and the exclusion of non-lymphomatous lymph node pathology such as necrotizing granulomatous inflammation, dermatopathic lymphadenopathy and Castleman's disease. In one case, the FCM on the FNA material was more accurate than the FCM that was performed on the fresh lymphoid tissue. In this case and as shown in the representative figures, the T-cell anaplastic large cell lymphoma involved certain areas of the lymph node where the aspirated material were more representative and most likely was not affected by the dilutional effect of the normal surrounding lymphoid cells.

Light chain restriction allowed an unequivocal diagnosis of B cell lymphoma to be made in most studied cases (12/14 of B-celllymphoproliferative disorders) (Figures 1B, 1D, 3 & 5). Two cases diagnosed as follicular lymphoma and diffuse large Bcell Lymphoma, were both positive for CD10 however not showing K/L chain restriction. Those two cases werediagnosed by morphology, supported by the presence of CD10; a marker which shouldn't have been present normally. Negativity of kappa and lambda thus doesn't exclude malignancy; however monoclonality proves its presence. Dunphy et al reported a concordance rate of 94.1% between FCM and histopathology while Mandacova et al. reported a concordance of 89% between the two techniques in cases suspected to be lymphoma (4,18). However, their number of cases was much higher than ours and Mandacova included bone marrow samples in their cases.

As it is well-known and was heavily investigated in the literature and simply because the number of malignant cells in Hodgkin lymphoma (HL) is very low compared to the surrounding benign lymphoid cells, all our cases of HL showed no monoclonality in FCM. The supportive element to the presence of Hodgkins Lymphoma is the disturbed ratio between CD4 & CD8. Although our FCM lymphoma panel lacks CD 30 and CD 15, we do FCM in HL suspected cases to exclude B cell lymphoproliferative disorders such as T cell rich B cell lymphoma.

The histological examination combined with immunohistochemistry confirmed the diagnosis of HL. It is worthwhile noting that our FCM lymphoma panel lacks CD 30 and CD 15, while they are available in our immunohistochemistry antibody panel to characterize HL diagnosis.

The present study demonstrates that the application of ancillary methods to cytological specimens such as IHC and FCM leads to precise subclassification of lymphomas in the majority of cases. In most cases typing of non-Hodgkin lymphoma was according to the WHO classification and it agrees in this contest with the results of other investigators (19-22).

Correct selection of cases submitted for FCM is crucial and enhances diagnostic accuracy of the cytomorphological interpretation. As recommended by many researchers, combining cytomorphology with immunophenotyping by FCM in the context of the appropriate clinical data is very helpful to reach a specific diagnosis on lymphadenopathy thru FNA.

In conclusion, the combination of FCM analysis and cytomorphology on FNA material for lymphadenopathy samples is a valuable technique for providing rapid, precise and minimally invasive tool for diagnostic evaluation of lymphoid cell population. In large centers FCM should be incorporated as a routine diagnostic method for lymphadenopathy. Similar to other centers, we were able to validate this diagnostic approach in our laboratory.

#### **Conflict of interest:**

We declare there is no conflict of interests which may bias our study; and no financial fund has been given from any company. The usage of different reagents was upon, the experience of our pathologists or, the availability in the market.

# Acknowledgement:

We acknowledge all the financial support to King Fahad Specialist Hospital –Dammam; which is a tertiary hospital and cancer center, a governmental non profitable organization. This work is part of our work to ensure a better quality, faster and accurate service to the cancer patients.

			L	YMPH NODE VALIDATION FLOW CYTO	MET	RY vs H	IISTOF	PATHOLOGY			
	name	MRN	sample	date diagnosed		Gender	Age		Flowctometry	Histopathology	Comment
1	Mariam Al-Najrani	45205	LN	diagnosed with lymphadenitis	1	F	37y	Positive	CD19, 20,23,K,L,45,2,3,4,5,7,8	No IHC	Histopathology
				on 17-10-2009				Negative	CD10,25, 103, 11c, TdT No Malignancy/monoclonality	No IHC Castleman's Diseas,plasma cell	report was depending on
								Diagnosis	detected	variant,Negative for malignancy	morphology
2	Ahmed Al_Samti	29886	LN	isly diagnosed with dermatopathic lymphader	2	M	27y	Positive	CD3,4,partial5,8,19,20,K,L,34,TdT	No IHC	
				in 25-09-2011				Negative	CD23,103, 11C	No IHC	
								Diagnosis	No Malignancy/monoclonality detected	LymphadenopathyNegative for malignancy	
3	Ahlam Al-Khoufi	53586	LN	previously diagnosed with HL	3	F	46y	Positive	CD2,3,4,5,7,8,19,K,L	CD30 & fascin	
Ĭ				in 4-9-2012			,	Negative	CD10,34,20	CD3, 20,15	No availibility of CD30 by FCM
								Diagnosis	detected	variant	,
4	Ahmed Goma	53940	LN	previously established micronodular cirrhosis	4	M	54v	Positive	CD2.3.4.5.7.8.19.22.45	No IHC	Histopathology
				on 26-1-2012				Negative	CD10,20,23,25,slgM,103,lgG,K,L,11c, 34,TdT	No IHC	report was depending on
				01120-1-2012				Diagnosis	detected	Inflamation	morphology
5	Sattam Yousef	60978	LN	newly diagnosed as Burkitts lymphoma	5	M	33y	Positive	CD19,20,22,slgM,K, 45	CD20,Mum-1,Bcl-6, Faint CD10	
				20 0 2044				Negative	CD2,3,4,5,7,8,10,23,25,L,11c,34,HLA-	ODS Del S Ovelle D4 TelT	
				on 20-9-2011				Diagnosis	DR Burkitt Lymphoma	CD3,BcI-2,CyclinD1, TdT Burkitts lymphoma	
6	Saleha Harobi	50236	LN	previously diagnosed with DLBCL	6	М	44y	Positive	CD2,3,4,5,7,8,19,20,23,45	No IHC	Histopathology
Ĭ				10/03/2010				Negative	CD10,103,K,L,11c,34,TdT	No IHC	report was
								Diagnosis	No Malignancy/monoclonality	inflamationpredominently non-	depending on
_		00500			_				detected	necrotizing	morphology
-/	Abdulla Al-Nasser	60598	LN	previously diagnosed with b-cell lymphoma on 19-09-2011	7	M	24y	Positive Negative	CD2,3,4,5,7,8,25,19,20,K,L TdT, CD34,23	No IHC No IHC	Histopathology report was
		1		011 10-05-2011					No Malignancy/monoclonality	histocytosis No	depending on
								Diagnosis	detected	Malignancy/monoclonality detected	morphology
8	Rabab Al-Ema	60930	ovary	newly diagnosed 6-9-2011	8	F	24y	Positive	CD3,4,8,19,20,K,L	faintCD117, PLAP,CK	
		1						Negative Diagnosis	CD10,CD117	CD30, AFP, HCG lymph node Negative for metastasis	
a	Naif Al-Otaibi	60554	FNA	newly diagnosed on 23-08-2011	9	М	30y	Positive	detected CD3,4,5,7,	CD3,5,7,4,30	
3		55554			,	.**	July	Negative	CD2,8,10,19,20,25,103.K,L,11c,34,	4=3400.004	
									TdT	CD2,Alk-2, CD10 & Bcell markers	
								Diagnosis	detected	cell lymphoma	sample was dilute
10	Naif Al-Otaibi	60554	LN	newly diagnosed on 23-08-2011	10	M	30y	Positive	CD2,3,4,5,7,8,19,K,L	CD30,3,7,4	with many norma
								Negative	CD10,25,103,11c,34,TdT,30 No Malignancy/monoclonality	Alk-1,CD2,5,15,8,20 Alk-Negative T-cell anaplastic Large	cells& apparently the normal part of
								Diagnosis	detected	cell lymphoma	the LN was sent t
11	yousef AL-Bohasan	58075	LN	iously diagnosed with lymphoepithelial sialade	11	M	71y	Positive	CD2,3,4,5,7,8,19,20,23,K,L	No IHC	
				on 24-04-2012				Negative	CD103,11c,34,TdT	No IHC	
		l						Diagnosis	detected	detected	
12	Ali Al-Saheemi	54533	LN	diagnosed with DLBCL	12	M	49y	Positive	CD19,20,23,K, 11c,HLA-DR CD2,3,4,5,7,8,10,25,103,L,34,45,TdT,	Mum-1	
				on 4-12-2010				Negative	14	CD3, 10	
								Diagnosis	B-Cell Lymphoma	Diffuse Large B-cell Lymphoma	
13	Ahmed Al-Moher	61249	LN	newly diagnosed on 10-10-2011	13	M	61y	Positive	DR,slqM,L,	67	
								Negative Diagnosis	CD3.4.5,8,38,103,11c,30,K	CD3,5,21,23	22222
14	Fawzya Al-Abad	61681	LN	newly diagnosed on 24-10-2011	14	F	48y	Positive	B-Cell Lymphoma CD2,3,4,5,7,8,19,20,23,K,L	IIIa No IHC	((((
•	r unzyu / u / uuu	0.00.		nowly diagnoods on 21 to 2011	•	Ċ	.0,	Negative	CD25, 103,11c,TdT	No IHC	
								Diagnosis	detected	Atypical Large Lymphoid Cells	
15	Zahra Al-Muslem	61262	neck	newly diagnosed on 18-10-2011	15	F	18y	Positive	CD2,3,4,5,7,8,19,20,23,25,K,L	CD15,30,few CD20	No availibility of
								Negative Diagnosis	CD10,103,11c,34,TdT detected	CD3 Classical Hodgkin's Lymphoma	CD30 by FCM
									CD3,4,5,8,19,20,22,45,56,79b,HLA-	Classical Hougkins Eymphoma	
16	Abdulrahman Nagi	60928	LN	newly diagnosed on 10-10-2011	16	M	22y	Positive	DR	No IHC	
								Negative	CD23,10,25,slgM,38,103,K,L,11c,34,		
		1							TdT_14_FMC7	No IHC	
		1	<b>-</b>					Diagnosis	detected	benign Reactive lymph node occasional CD20, weak CD15 &	
17	Nawal Al-Timani	56946	LN	newly diagnosed on 18-10-2011	17	F	56y	Positive	CD2,3,4,5,7,8,19,20,K,L,	CD30	No availibility of
								Negative	CD10,23,25,103,11c,34,TdT	CD3, CD45	CD30 by FCM
		<u> </u>	L				L	Diagnosis	detected	Classic Hodgkins Lymphoma	
18	Masoma Al-Awjami	915	Thyroid	viously diagnosied with focal thyroid inflamati in 16-2-2012	18	F	55y	Positive	CD2,3,4,5,7,8,19,20,23,K,L	No IHC	report was
		1		III 10-2-2012				Negative Diagnosis	CD10,25,103,11c,34 detected	No IHC Hashimoto's Thyroditis	depending on morphology
19	Saadon Al-Sadoon	42813	LN	diagnosed with ostiosarcoma in 22-9-2009	19	М	18y	Positive		No IHC	c.priology
								Negative	CD10,25,103,11c,34,TdT,30	No IHC	
	B	40	<b>.</b>	, , ,		L	0.5	Diagnosis	Normal T & B cells, No monoclonality		
20	Rami Al-Harbi	12695	ppharyn biopsy	diagnosed in 26-10-2011	20	M	23y	Positive Negative	CD2,3,4,5,7,8,19,20,23,K,L CD10,25,103,11c,34,TdT	No IHC	
		1	υισμεγ					Diagnosis	detected	No IHC Reactive Follicular hyperplasia	
21	Fatima al-Marzoog	51134	LN	diagnosed in 29-11-2011	21	F	76y	Positive	CD10,19,20,23,L	CD20,23,10,Bcl-2, faint Bcl-6	
								Negative	CD2,3,4,5,7,8,25,K,103,11c,34,TdT	CD3,5,Cyclin D	
		+	-					Diagnosis	Follicular Lymphoma	grade	
22	Safa Alawad	53440	LN	usly diagnosed with classical Hodgkins Lymph	22	F	11y	Positive	CD2, 3, 4, 5, 7, 8, 19,20, 11c, K ,L, partial 23	CD3, CD20	
		1		in 16-8-2010			,	Negative	CD10,25,103,34,TdT	CD30, CD15	
								Diagnosis	No Malignancy/monoclonality	hyperplaia, no evedience of	
		ļ							detected	recurrent Hodgkins Lymphoma	
		55820	LN	usly diagnosed with classical Hodgkins Lymph	23	M	10y	Positive	CD2,3,4,5,7,8,19,K,L	CD30	toNebraska
23	Mujtaba Alalwan	1		lymphocytes rich in 23-3-2011				Negative Diagnosis	CD10,23,25,33,11c,34,TdT	CD15	medical center No evidence o
23	Mujtaba Alalwan						47		detected CD2,3,4,5,7,8,19,K,L,11c	Atypical Lymphoproliferative lesion CD15,30,focal CD20	INO EVIUETICE O
		64325	ΙN	iagnosed with classical Hodokins I vmphoma	24	M					
	Mujtaba Alalwan  Mohammed Al-Jaber	64325	LN	liagnosed with classical Hodgkins Lymphoma in 19-2-2012	24	M	17y	Positive Negative	CD10,23,25,33,34,TdT	CD45,3	No availibility of
		64325	LN		24	М	179		CD10,23,25,33,34,TdT detected	CD45,3 Classic Hodgkins Lymphoma	No availibility of CD30 by FCM
24	Mohammed Al-Jaber			in 19-2-2012				Negative Diagnosis	CD10,23,25,33,34,TdT detected CD3,4,5,8,19,45,20,22,23,79b,K,L,	CD45,3 Classic Hodgkins Lymphoma	
24		64325	LN		24	M	20y	Negative	CD10,23,25,33,34,TdT detected	CD45,3	

March   Art   Ar												
	26	Theiden Al Eid	44004	LNI	diagraphic 10 F 2012	20		24	Positive	CD3,4,5,8,19,K,L,11c,20,22,23,79b,	CD2 20 24 5 40	
2	20	IIIdiudii Al-Eiu	44091	LIN	diaggnosis 19-3-2012	20	IVI	24	Negative		CD3,20,21,5,10	
10.75   2.00   10.75   2.00   10.75   2.00   10.75   2.00   10.75   2.00   10.75   2.00   10.75   2.00   10.75   2.00   10.75   2.00   10.75   2.00   10.75   2.00   10.75   2.00   10.75   2.00   10.75   2.00   10.75   2.00   10.75   2.00   10.75   2.00									Diagnosis	detected		
Company   Comp	27	Hager Al-Abdulaziz	46624	LN		27	F	12y		CD3,4,8,19, partial slgM,K,L		
Part					III 20-5-2005							
20   1	28	moneera Al-Zuabi	65308	LN		28	F	48y			CD20	
2004   A.					in 28-3-2012							
Description   Control	29	Ali Al-Ashwan	58583	LN	previously diagnosed as HL on 4-5-2011	29	М	14y				
Section   Process   Proc					· · · · · ·					CD10,103, slgM		
Monach Record   1778   147	20	ferrer Al Face	40077	LNI	40/00/0040	20		20				
S Annex Policy 1778 Us 1508/00/12 37 N 6 Policy 2004 Selected 1990 No. 10 No. 1	30	layez Al-Essa	103//	LIN	18/08/2012	30	IVI	30				
									Diagnosis	detected	follow up	
20   Designed   Designed   Designed   Designed   Designed   Color   Designed   Designe	31	Ahmad Hosny	17783	LN	18/08/2012	31	М	9у				
200   1972   1												
Amount	32	Ibrahim Bo-Howaid	68612	LN	diagnosed on 19-8-2012	32	М	49y				No qualibility of
Section   Column												
March   Marc	22	Arasi Al Cabaina	60003	LN	diagraphed on 25 9 2012	22		16.				
Second A Chairs	33	Aleej Al-Gudase	00903	LIN	diagnosed on 25-6-2012	33	Г	TOY			CD30,15	
Display									Diagnosis	detected		CD30 by FCM
Dispress	34	Fatemah Al-Owiss	48560	FNA		34	F	44y				
Confusion   Conf					DEBGE germinal center type					CD 10' RIGIN' FSILIDGS		
Magazine									_		highly suspesious for lymphoma	
Second Content   Seco	35	Alyaa Al-Shameri	24042	LN	diagnose as DLBCL on 19-9-2009	35	F	64y		CD19, CD23	CD20, BCL-2, BCL-6 Ki 67	
Make   March												
Magame	36	Mujtaba Al-Alwan	55820	LN	diagnosed as EBV related lymphadenitis	36	М	10Y				
Martine A-Hall   68642   LH   diagnosed 12-9-2012   37   M   19   Positive   C02.4,8,19.64, supps, starting   C02.0,15 FMS									Negative	CD10,19,slgM, 34	CD20	
Magneties	27	Mohammed Al Lilel	68040	I NI	diagnosed 42 0 2042	27	1.4	15.				
Page	3/	worldmineu Al-Pilai	00042	LIN	ulayriosed 12-9-2012	31	IVI	ıby				No availibility of
Negative   C019									Diagnosis	detected	Sclerosis	CD30 by FCM
Object   Part   Object   Obj	38	Hattan Al-Mutairi	58049	LN	diagnosed 4-5-2011	38	M	4y			CD15,CD30,Fascin,PAX5	No availibility of
Section   Sect										<del></del>		CD30 by FCM
April	39	fatima Baaqeel	69105	FNA	diagnosed 14/10/2012	39	F	44y		CD2,3,4,5,7,8,TCRab,19,k,I,		
Add   Aber Sahbil   36747   LN   diagnosed 10-3-2000   40   M   469   Positive   Positive   Aber Sahbil   36747   LN   diagnosed 10-3-2000   40   M   469   Positive   Positive   Aber Sahbil   36747   LN   diagnosed 10-3-2000   40   M   469   Positive   Positive   Aber Sahbil   Sahber Sahbil   Sah									Negative	TCRgd, CD1a, CD34		
Add   Aber   Sahtol										00/ - ( T    OD 100		
40 Jaber Saholi									Diagnosis			
Adapter Sation   3674   LN   diagnosed 10-3-2009   40 M   469   Positive   3022 38.788 ft.1-6.5   CD2.8.45.7.8 to 10.03 188 sight N, Linex 70%												
Negative   C7, C01, 34, 55, R4, D4R   C7, C01, 34, 58, R4, D4R   C7, C01, 34, S4, R4, D4R   C7, C01, C01, C01, C01, C01, C01, C01, C01										disease.		
Alian Atlateed	40	Jaber Sahloli	36747	LN	diagnosed 10-3-2009	40	M	46y	Positive	19,20,23,38,79B,11c,45	CD20,BCL6,Mum,Ki index 70%	
Alian Atlateed									Negative	CD2.3.4.5.7.8.10.103.138.slgM.k.l.FM		
Algorithms									· ·	C7,CD14,34,56,HLA-DR		
41   Min Altaleed    68838   FNA									Diagnosis	D 0 III		
Agroup	41	Ailan Altaleedi	65838	FNA	Known case	41	М	71v	Positive			
Age   Paisal A-Mutairi   61207   LN   Dusly diagnosed Hodgkins lymphoma in 27-9   42   M   4   Positive   Colorada   1,192.KL   Colorada   5,192.KL   Co		.,						,		CD3,4,5,8,38,10,23,103,slgM,K,11c,3		
42   Faisal Al-Mutain   61207					diagnose 18-4-2012					4		
Neadline	42	Faisal Al-Mutairi	61207	IN	nusly diagnosedf Hodgkins lymphoma in 27-9	42	М	4 v				
43 Fatra Al-Hemaidan 6662 FNA newly diagnosed in 27-10-2012 43 F 56y Positive CD3.4.5.8.2.3/L.3.4 No. FtC. No availability of Diagnoses B. Coll (1979) No. FtC. No availability of Diagnoses B. Coll (1974) No. FtC. No. availability of CD3.4.5.8.2.3/L.3.4 No. FtC. No. availability of CD3.4.5.8.2.3/L.3.4 No. FtC. No. availability of CD3.4.5.8.1.3/L.1.4. No. FtC. No. availability of CD3.4.5.8.1.3/L.1.4. No. FtC. No. availability of CD3.4.5.8.1.3/L.1.4. No. FtC. No. availability of CD3.4.5.7.8.1.0.1.2.3.3.4 No. FtC. No. availability of CD3.4.5.7.8.1.0.1.2.3.3.4 No. FtC. No. availability of CD3.4.5.7.8.1.0.1.2.3.3.4 No. FtC. No. availability of CD3.4.5.7.8.1.0.1.2.3.3.4. No. FtC. No. availability of CD3.4.5.7.8.1.0.2.3.3.4. No. FtC. No. availability of CD3.4.5.7.8.1.0.2.3.4.1.3.3.4. No. FtC. No. availability of CD3.4.5.7.8.1.0.2.3.4.5.3.4. No. FtC. No. availability of CD3.4.5.7.8.1.0.2.3.4.5.3.4. No. FtC. No. availability of CD3.4.5.7.8.1.0.2.3.4.5.3.4. No. FtC. No. availabi		Taloa 7 Trioan	0.20.		Sacry dagnessarrisaginis iyinpilana iii 27 o	-		.,		CD10		
Negative										detected		
A	43	Fakra Al-Hemaidan	69628	FNA	newly diagnosed in 27-10-2012	43	F	56y				
Abderahman Ai-rabisah   32792   FNA   newly diagnosed in 12:11:2012   44   M   51y   Positive   CD34.58.19.KL   No. H°C.   No. availability of CD30 by FCM   Positive   CD34.58.19.KL   No. H°C.   No. availability of CD30 by FCM   Positive   CD3.48.78.10.19.23.24   No. H°C.   No. availability of CD30 by FCM   Positive   CD3.48.78.10.19.23.25.103.KL.33   No. H°C.   No. availability of CD3.48.78.10.19.23.25.103.KL.33   No. H°C.   No. availability of CD3.48.78.10.19.23.25.103.KL.33   No. H°C.   No. availability of CD3.48.78.10.19.23.25.103.KL.34   No. H°C.   No. availability of CD3.48.78.19.19.23.KL.45   No. H°C.   No. availability of CD3.48.78.19.23.KL.45   No. H°C.   No. availability of CD3.48.78.19.23.KL.45   No. H°C.   No. h°C.   No. availability of CD3.48.78.19.23.KL.45   No. H°C												CD30 by FCM
Name	44	Abderahman Al-rabiaah	32792	FNA	newly diagnosed in 12-11-2012	44	M	51y	Positive	CD3,4,5,8,19,K,L	No IHC	No availibility of
A												
No history of malgrancy   Negative   CD23,4,5,7,8,10,19,23,25,103,k1,33   A,T of the stead on 5-11-2012   46   M   14y   Positive   CD23,4,5,7,8,10,19,23,25,103,k1,33   A,T of the stead on 5-11-2012   46   M   14y   Positive   CD23,4,5,7,8,10,19,23,25,103,k1,33   A,T of the stead on 5-11-2012   46   M   14y   Positive   CD23,4,5,7,8,10,19,23,25,103,k1,33   A,T of the stead on 5-11-2012   46   M   14y   Positive   CD23,4,5,7,8,10,19,23,25,103,k1,33   A,T of the stead	45	Muna Al7ahrani	57471	I NI	natient with history of obstructive isundice	45	F	26V		CD45 11c		
No history of malignancy   tested on 5-11-2012   Diagnosis   Dia	43		51411	LIN	patient with history of obstructive jaulituice	40	<u>'</u>	201		CD2,3,4,5,7,8,10,19,23,25,103,k,l,33.	ророжини	
AB-ham Al-Hshem   70609   LN   newly tesed on 11-11-2012   46   M   14y   Positive   CD2,3,4,5,7,8,19,23,K,L,45   No HC									. 5	34,TdT		
Negative   CD10,25,103,33,11c,34   No HC	40	Hacham Al Haham	70000	N		AC	1.4	144	Diagnosis			
Diagnosis  There is 16% population coexpressing CD4& CD8 suggesting follicular hyperplasia, inspect the patient for viral infection coexpressing CD4& CD8 suggesting follicular hyperplasia, inspect the patient for viral infection patient for viral infecti	46	nasnem Al-HSNem	10609	LN	newly tesed on 11-11-2012	46	M	14y				
A7 Mareem alkhaldi 69182 LN 07/01/2013 F 37y Positive CD3 CD4 CD5 CD8 CD19, KAPPA LAMEDA No HC Negative for granuloma or malignancy or malignancy or granuloma or malignancy or granuloma or malignancy or granuloma or metastatic malignancy or granuloma or metastatic malignancy or granuloma or metastatic tumer, poliagnosis biagnosis holding rock page to provide the province of poliagnosis or poli											t	
A   Mareem alkhaldi								1	Diagnosis		College Lands and Aller College	
47 Mareem alkhaldi								1				
American A						$\vdash$		$\vdash$	D:r'	CD3,CD4,CD5,CD8,CD19, KAPPA,	parent for viral intection	
Diagnosis No Matignancy monoclonality detected malignancy malignan	47	Mareem alkhaldi	69182	LN	07/01/2013		F	37y		LAMBDA	No IHC	
48 Abdulla Al-Shehri 72023 LN 08/01/2013 M 111y Positive LAREDA No HC NO										CD10, CD34, TdT	No IHC	
Abdula Al-Shehri								1	Diagnosis			
No H2   No H									Positiva	CD3,CD4,CD7,CD8,CD19, KAPPA,		
Diagnosis   No Matignancy/ monoclonality   Negative for primary lymphoma, metastatic matignancy or granulomatous inflamation   CD79a, CD20, (CD3 & 5 positive on the Leets)	48	Abdulla Al-Shehri	72023	LN	08/01/2013	-	М	11y		LAMBDA		
Diagnosis   No Matignancy / monoclonality   metastatic malignancy or   detected   granulomatous inflamation									rvegative	<del> </del>		
Age								1	Diagnosis		metastatic malignancy or	
Age						<u> </u>				detected		
Negative CD3,CD4,CD8,CD20 CD10, CD30, inconclusive kappa & lambda, QNS sample was processed show 10% population of large B-cell Lymphoma with plasmabilastic features, non CcColl and Al-Harbi CD3, CD4, CD8, CD19 No HC Negative CD3, CD4, CD8, CD19 No HC Negative CD20, kappa, Lambda No HC Negative CD3, CD4, CD8, CD19 No HC Negative CD20, kappa, Lambda No HC Negative CD3, CD4, CD8, CD19 No MC Negative CD20, kappa, Lambda No HC	40	Hussain Al-Shoman	63005	I NI	13/01/2013		М	61v	Positive	CD19		
inconclusive kappa & lamidad_QNS   Sample was processed show 10%   Diffuse Large B-cell Lymphoma   with plasmatilastic features, non   G.C.B. type	49	1 100 Sall AFOIDHIAH	00990	LIN	15/01/2013		IVI	Uly	Negative			
population of large B-cells. But with plasmablastic features, non clonality couldn't be proven G.C.B type  50 Fahad Al-Harbi 70828 LN 13/01/2013 M 73y Positive CD3, CD4, CD8, CD19 No HC Negative CD20, kappa, Lambda No HC Diagnosis No Malignancy/ monoclonality polymorphic population of										inconclusive kappa & lambda. QNS		
								1	Diagnosis			
50   Fahad Al-Harbi   70828   LN   13/01/2013   M   73y   Positive   CD3, CD4, CD8, CD19   No HC								1				
Negative for metastatic turner, Diagnosis No Malignancy/ monoclonality polymorphic population of	50	Fahad Al-Harbi	70828	LN	13/01/2013		M	73y		CD3, CD4, CD8, CD19	No IHC	
Diagnosis No Malignancy/ monoclonality polymorphic population of									Negative	CD20, kappa, Lambda		
								1	Diagnosis	No Malignancy/ monoclonality		
						L	<u></u>	L	g0010			

51	Najla Al-Abdulqader	71219	LN	13/01/2013		F	33y	Positive Negative	CD19, CD20 Lambda CD3, CD4, CD8,kappa	No IHC No IHC	
								Diagnosis	QNS sample sugest B-cell	Atypical large lymphoid cells with rare formed non-necrotizing granulomas. Refer to flow result.	
52	Ariam Al-Shammari	64001	LN	15/01/2013		F	7Y	Positive	lymphoproliferative clone CD2,CD3,CD4,CD5,CD7,CD19,CD2 0,CD22,CD23, Kappa, Lambda	granulomas. Relef to flow result.	
32	Alain Ar-Silaininan	04001	LIN	1301/2013			, ·	Negative	CD103, CD10	CD3, CD20, CD30, CD15 fail to	
								Diagnosis	No Malignancy/ monoclonality detected	elict viable RS cells. No evidence of residual hodgkins lymphoma	
53	Mohammed Al-Shamma	72017	LN	20/01/2013		М	18y	Positive	CD2,CD3,CD4,CD5,CD7,CD19,CD2	CD45,CD20,EMA,Fascin, BCL6,	
								Negative Diagnosis	3 dim, Kappa, Lambda CD10,CD5,CD103,CD11c, CD34 No Malignancy/ monoclonality	CD30 CD15 CD3 Nodular Lymphocyte predominant	
								Positive	detected	hodgkin lymphoma (NLPHL)	
54	Nawal Al-Timani	56946	LN	27/01/2013		F	57y	Negative	CD3,CD4,CD8,CD19Kappa, Lambda	No IHC No IHC	
								Diagnosis	No Malignancy/ monoclonality	few large atypical cells, Hodgkins lymphoma cannot be totally	
	Khalifah Al-Hamad	72276	LN	28/01/2013		F	33v	Positive	CD2,CD4,CD5dim,CD7dim,CD8,	excluded	
50	Manan Arianau	72270	LIN	2001/2013			July		CD38,TdT dim & partial CD1a,CD3,CD10,CD19,CD20,CD23, CD25,	TdT, CD3	
								Negative	CD103,Kappa,Lambda,FMC7, CD11c,CD34	CD20	
56	Rayan Al-Eniz	72217	LN	29/01/2013		F	8y	Diagnosis Positive	Tcell Lymphoma/Leukemia CD3.CD4.CD8.	T-cell Lymphoblastic Lymphoma No IHC	
								Negative Diagnosis	CD19, Kappa,Lambda No Malignancy/ monoclonality	No IHC Tuberculous Granulomatous	
								Positive	CD3,CD4,CD8,CD19,	Inflamation	
57	Mutib Al-Harthi	2179	LN	02/02/2013		М	49y	Negative	Kappa,Lambda	No IHC	
								Diagnosis	No Malignancy/ monoclonality detected Normal T cells 26 %, & are positive	reactive lymphe node, negative for malignancy	
								Positive	for CD3,CD2,CD4,CD5,CD7,CD8, &CD38 Normal B cells 5 %, &are positive for CD19,CD20,kappa	CD15, CD30	
58	Noura Al-Arfaj	12643	LN	05/02/2013		F	77y	Negative	&lambda.  The sample is infiltrated with CD45	CD3, CD20, CD10	No availibility of CD30 by FCM
								Diagnosis	negative cells 60%, are seen in the scatter; the population is negative to CD3, CD10, CD5, CD19CD15;	Hodgkin Lymphoma.	0500 591 0111
						<b> </b>		_	morphology and flowcytometry suggestive of Hodgkin Lymphoma.		
59	Nawal Al-Timani	56946	LN	09/02/2013		F	57y	Positive Negative	CD3, CD4, CD8,CD19,kappa,lambda	No IHC	
								Diagnosis	No Malignancy/ monoclonality detected	reactive lymphe node, negative for malignancy	
60	Ahmed Mahalawi	72112	LN	12/02/2013		М	48y	Positive	CD3,cCD3, CD4, CD8, CD19, CD79a, kappa, lambda, MPO	No IHC	
								Negative Diagnosis	No Malignancy/ monoclonality	No IHC	
								_	detected	negative for malignancy	
61	Nowal Al-Timani	56946	LN	23/02/2013		F	55y	Positive Negative	CD3, CD4, CD5, CD7, CD8, CD19, CD20, CD79a, kappa, lambda, CD45 CD10, CD23, CD25, CD103, CD33, CD117, MPO, FMC7, CD11c, CD34,		
									CD64, Tdt No Malignancy/ monoclonality		
62	Modi Al-Abdulla	73283	LN	26/02/2013		F	19y	Diagnosis Positive	detected CD3, CD4, CD8	negative for malignancy	
								Negative	CD19, kappa, lambda No Malignancy/ monoclonality		
								Diagnos is Positive	detected CD3, CD4, CD5, CD8, CD19, CD20,	atypical lymphoproliferation	
63	Khatebah Al-Meshal	71826	LN	10/03/2013		F	34y	Negative	kappa, lambda CD10	No IHC No IHC	
								Diagnosis	No Malignancy/ monoclonality detected	reactive lymphe node, negative for malignancy	
64	Zafer Al-Shhrani	73544	LN	10/03/2013		М	75y	Positive	CD4, CD5, CD3, CD8, CD19, kappa, lambda	CD15, CD30,	No availibility of
								Negative Diagnosis	No Malignancy/ monoclonality detected	CD3, CD20, CD23, CD10	CD30 by FCM
GE	Modi Al-Abdulla	73283	LN	11/03/2013		F	19y	Positive	CD3, CD4, CD5, CD8, CD19, CD20, kappa, lambda	Classical Hodgkin's Lymphoma	
0.0	INDUI AI-ADUUIIA	73203	LIN	11/03/2013			159	Negative	CD10 No Malignancy/ monoclonality	No Malignancy / kikuchi Fujimoto	
								Diagnosis	detected CD19, CD5, CD20, CD23, kappa,	diseas	
66	Ali Al-Mohsen	67822	LN	13/03/2013		М	62y	Positive	partial FMC7, CD45	CD20, CD5, Zap70	
								Negative	CD3, CD4, CD7, CD8, CD10, CD25, CD103, CD38, Lambda, CD34	CD3	
67	Murtadah Al-Salem	72695	LN	26/03/2013		M	24y	Diagnosis Positive	CLL/SLL CD3, CD20, kappa, lambda	SLL No IHC	
								Negative Diagnosis	No Malignancy/ monoclonality	No IHC reactive lymphe node, negative for	
68	Moneera Al-Zuabi	65308	LN	27/03/2013		F	49y	Positive	detected CD19, CD25, CD45, kappa CD5, CD10, CD23, CD103, lambda	malignancy CD20, CD79a,	
								Negative	CD5, CD10, CD23, CD103, lambda FMC7, CD11c	CD3, CD10, CD43	
								Diagnos is Positive	relapsed B-cell Lymphoma CD3, CD4, CD5, CD7, CD8, CD19,	Marginal zone Lymphoma in transformation to large b-cell lymphoma	
69	Hitham Al-Dokhi	74265	LN	06/04/2013		М	24y	Negative	CD20 CD38 CD45, kappa, lambda CD10, CD23, CD25, CD11c	CD15, CD30, CD10, CD20, CD3, CD45	No availibility of
								Diagnosis	T-cell population show inapropriate ratio between CD4/CD8 with	Classical Hodgkin's Lymphoma,	CD30 by FCM
70	Ajab Al-Qahtani	74397	LN	15/04/2013		М	73y	Positive	increased CD4, to correlate with clinical findings.	mixed cellularity type.	
70	,			100-120-13			, Sy	Negative	CD19, CD20, kappa lambda, CD5, CD10, CD23, CD25, CD38, CD103, FMC7, CD34	CD20, CD5, CD10, CD34, CD3	
					L	L		Diagnosis	B-cell lymphoproliferative disorder	Diffus e large B-cell lymphoma, non- germinal center type	
71	Mohamed Al-Hagri	74556	LN	22/04/2013		М	2y	Positive	CD3, CD4, CD5, CD8, CD19, CD20, kappa, lambda	No IHC	
								Negative	CD10, CD23, CD25, FMC7, CD11c, CD34	No IHC	
								Diagnosis	No Malignancy/ monoclonality detected CD3, CD4, CD8, CD19, kappa,	reactive lymphe node, negative for malignancy	
72	Saleh Alkhofi	74115	LN	22/04/2013		м	30y	Positive	CD3, CD4, CD8, CD19, kappa, lambda	CD15, CD30	No availibility of
								Negative Diagnosis	No Malignancy/ monoclonality	Classical Hadakin- !	CD30 by FCM
72	Lolwah Al-Rasheed	74721	LN	06/05/2013		F	54y	Positive	CD3, CD4, CD8, CD19, kappa, lambda	Classical Hodgkins Lymphoma  No IHC	
73		/21	LIN	55-53/2013		Ĺ	Эну	Negative		No IHC polymorphic population of	
						l		Diagnosis	No Malignancy/ monoclonality	lymphocytes & rare follicular cells consistent with chronic lymphocytic	
						-			detected CD3, CD4, CD5, CD7, CD8, CD19,	thyroditis	
74	Mutlaq Al-Amohsen	24064	LN	19/05/2013		М	69y	Positive	CD20, CD23, kappa, lambda CD10, CD25, CD103, fmc7, CD11c,	CD3, CD5, CD10, CD30,	
								Negative	CD38	CD8, TdT portion of the Lymph Node partially	
								Diagnosis	No Malignancy/ monoclonality detected	infiltrated with pereipheral T-cell lymphoma	
75	Abdulmalek Al-hamed	75500	LN	20/05/2013		М	13y	Positive	CD3, CD4, CD8, CD19, kappa, lambda	CD3, CD10, TdT	
								Negative Diagnosis	No Malignancy/ monoclonality	CD20, CD5, Zap70 T-cell Lymphoblastic	
		<u> </u>		l		I			detected	Leukemia/Lymphoma	

	1			1	_	1		CD3, CD4, CD8, CD19, kappa,	T	
76 Mohammed Al-taibi	51769	LN	20/05/2013		м	5y	Positive	lambda	No IHC	
							Negative		No IHC	
							Diagnosis	No Malignancy/ monoclonality detected	reactive lymphe node, negative for malignancy	
77 Abdullah Al-Harbi	62491	LN	03/06/2013		М	29v	Positive	CD2, CD5, CD7CD3, CD4, CD8, CD19, kappa, lambda	No IHC	
7 7 Dadiidii 7 T T Li Di	02.01		00/00/2010			20,	Negative			
							Diagnosis	No Malignancy/ monoclonality	reactive lymphe node (HIV related benign lymphadenopathy), negative	
				+-				detected CD3, CD4, CD8, CD19, kappa,	for malignancy	
78 Hussain Al-Mussaly	68137	LN	10/06/2013		м	16v	Positive	lambda	No IHC	
							Negative		No IHC reactive lymphe node, negative for	
							Diagnosis	No Malignancy/ monoclonality detected	malignancy	
79 Ali Al-Ghamdi	62394	LN	17/06/2013	_	M	48y	Positive	CD19, CD20, lambda	CD10, CD20, CD5, CD3, TdT	
				-			Negative	CD19, CD20, lambda CD3, CD5, CD10, slgM, kappa Monoclonal B-cell population. To be	CD5, CD3, TdT	
							Diagnosis	correlated with histopathology	low grade follicular lymphoma	
80 Lolwah al-Bin ali	69825	LN	22/06/2013		F	56y	Positive	CD19, kappa	CD45, CD20	
							Negative	CD3, CD4, CD5, CD8 lambda	CD10, CD30,	
							Diagnosis	Large cells show B-cell monoclonality, correlate with	Diffuse Large B-cell Lymphoma,	
							Diagnosis	histopathology	non-germinal center b-cell	
81 Huda Al-Turki	37997	LN	23/06/2013	1	F	45y	Positive	CD10, CD19, Lambda	CD20, CD10	
							Negative	CD3, kappa	CD5 Follicular Lymphoma	
2011 4101	74404		07/07/0040	_			Diagnosis	Follicular Lymphoma	Follicular Lymphoma	
82 Hasna Al-Shammary	74184	LN	07/07/2013	+	f	36y	Positive Negative	CD3, CD4, CD8, CD7	<del> </del>	
							Diagnosis	No dim CD4 seen, correlate with histopathology	No difinite evidence of lymphoma seen	
83 Anas Al-Abdulatif	66055	LN	09/07/2013	1	M	21y	Positive	CD4, CD8	CD30_CD15	
							Negative		CD45, CD20, CD3	No availibility
							Diagnosis	no monoclonality proven, correlate with histopathology	Recurrent Classical Hodgkins Lymphoma	CD30 by FCN
84 Ali Al-Bouri	75352	LN	10/07/2013	+	М	7у	Positive	CD4, CD8	CD68	
							Negative			
							Diagnosis	No Malignancy/ monoclonality detected	Reactive Hyperplasia	
85 Rami Al-Harbi	12695	LN	15/07/2013		М	22y	Positive	CD3, CD4, CD8, CD19, kappa,	7	
85 Rami Al-Harbi	12695	LIN	15/07/2013		IVI	ZZY	Negative	lambda	<b></b>	
								No Malignancy/ monoclonality	reactive lymphe node, negative for	
							Diagnosis	detected	malignancy	
	=====		05/07/0040		F	=0	Positive	CD3, CD4, CD8, CD19, kappa,		
86 Dalilah Al-Haisoni	75562	LN	25/07/2013		F	53y	Negative	lambda	<b></b>	
							regative	<b></b>	<del> </del>	
							Diagnosis	necrotic sample correlate with histopathology	Necrotizing Granulomatous Inflamation. Negative for malignancy	
87 Roqayah Al-Mahdi	65728	LN	31/07/2013		М	41y	Positive	CD5,CD7,CD3, CD4, CD8,CD19, CD20, kappa, lambda	CD30, CD15	No ovoilibiit
							Negative	FMC7,CD10	CD30, CD15 CD45, CD3, CD20	CD30 by EC
							Diagnosis	No Malignancy/ monocionality detected	Classical Hodgkins Lymphoma	5500 by 1 01
							Positive	CD3, CD4, CD5, CD8, CD19, CD20,		
88 Safa Al-Awad	53440	LN	31/07/2013	-	F	12y		kappa, lambda	CD30, CD15 CD45, CD3, CD20	No availibility
							Negative	CD10, FMC7 No Malignancy/ monoclonality	CD45, CD3, CD20	CD30 by FCI
							Diagnosis	detected	Classical Hodgkins Lymphoma	
89 Khalil Al-Atevah	76804	LN	31/07/2013	1	М	26v	Positive	CD3, CD4, CD5, CD8, CD19, CD20, kappa, lambda	CD30, CD20	
	1.0007		0.0072010			20,	Negative	CD10, FMC7	CD45, CD3, CD15	No availibility CD30 by FCI
							Diagnosis	No Malignancy/ monoclonality		CD30 By FCI
							Diagnools	detected	Classical Hodgkins Lymphoma	

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