#### Live Body Weight Changes and Physiological Performance of Barki Sheep Fed Salt Tolerant Fodder Crops under the Arid Conditions of Southern Sinai, Egypt

#### Shaker, Y.M.

Animal and Poultry Physiology Department, Animal and Poultry Production Division, Desert Research Center,

Mataria, Cairo, Egypt

yousrimshaker@yahoo.com

**Abstract**: This experiment was conducted at Ras Sudr Research Station belonging to Desert Research Center to elucidate the effect of feeding a mixture of salt tolerant fodder crops (*Atriplex nummularia, Sorghum bicolor* and *Pearl millet*) on their physiological performance under arid and salinity conditions of Southern Sinai, Egypt. Eighteen Barki sheep, 3- 3.5 years old with an average body weight  $28.50\pm1.02$  kg were randomly divided into two equal groups (9 each). The first group (G1) was fed berseem hay and served as control. The second group (G2) was fed a mixture of *Atriplex nummularia, Sorghum bicolor* and *Pearl millet* at percentage of 50, 25 and 25%, respectively. Both groups were supplemented with concentrate feed mixture (CDM). The mean values of total proteins (TP), albumin (AL), glucose (GLU), cholesterol (CH), total lipids (TL), triiodothyronine (T<sub>3</sub>), thyroxine (T<sub>4</sub>), insulin, aldosterone (AL), alanine amino transferase (ALT), aspartate amino transferase (AST), alkaline phosphatase (ALP), urea, creatinine, sodium (Na) and potassium (K) were determined in plasma. The values of globulin (GL), albumin/ globulin (A/G) ratio, Na/ K ratio and Na index were calculated. Live body weight and average daily gain were recorded.

The results revealed that animals fed salt tolerant plants mixture had a comparable live body weight and average daily gain with the control group. On the other hand, feeding salt tolerant plants elevated (P<0.05) ALT and creatinine levels in addition to Na, K and aldosterone concentrations (P<0.01). However, salt tolerant plants mixture group had lower (P<0.01) levels of GLU, CH, TL, insulin, ALP, thyroid hormones and urea than control group. Moreover, there were not significant differences between the two experimental groups of total proteins, albumin, globulin, A/G ratio and aspartate amino transferase (AST) values.

It could be concluded that feeding a mixture of salt tolerant plants (*Atriplex nummularia*, *Sorghum bicolor* and *Pearl millet*) without serious physiological hazards, would be an avenue to overcome the problem of feedstuff shortage prevailed under arid and salinity conditions of Sinai, Egypt.

[Shaker, Y.M. Live Body Weight Changes and Physiological Performance of Barki Sheep Fed Salt Tolerant Fodder Crops under the Arid Conditions of Southern Sinai, Egypt. J Am Sci 2014; 10(2s):78-88]. (ISSN: 1545-1003). <u>http://www.jofamericanscience.org.11</u>

Key words: Barki sheep, *Atriplex nummularia*, *Sorghum bicolor*, *Pearl millet*, plasma biochemical parameters, liver function, kidney function, electrolytes

# 1. Introduction

Many arid and semi-arid regions in the world particularly, in North Africa and the Near East regions, have soils and water resources that are too saline for most of the common conventional crop systems. Saline soils of various degrees occupy over 80 million ha in the Mediterranean basin (Anon., 2006). Halophytes and other salt-tolerant plants can constitute a major portion of the feeding program of livestock in the arid and semi-arid regions (Squires and Ayoub, 1994 and El Shaer, 2010).

Encouraging results have been found in some countries in the region with rehabilitation of potential halophytic species (Nemati, 1976 and El Shaer *et al.*, 2005). Several studies recommended cultivating of salt and drought tolerant fodder shrubs (e.g. *Atriplex spp.*) and salt and drought tolerant grasses and legumes such as sorghum and *Pearl Millet* which might fill the gab in feed production in arid and saline areas (Hanafy *et al.*, 2007 and El Shaer, 2010).

Atriplex nummularia has great potentialities since it is known to be tolerant to salinity and drought in addition to high content of crude protein, fiber and mineral contents (El Shaer, 2010 and Ben Salem *et al.*, 2010). *Pearl millet* and sorghum are among the most potential salt-tolerant grass species as good quality fodders for small ruminants in Egypt and other countries in the Near East (Anon., 2009). Sorghum is becoming increasingly important fodder crop as it tolerates well low rainfall, high salinity and high temperatures (Al Khalasi *et al.*, 2010).

However, high salt content of the salt tolerant plants is perhaps the major negative component. Furthermore, some anti-nutritive factors (ANFs) like lignin, oxalates and nitrates could restrict the utilization of some halophytes and salt-tolerant forages in livestock feeding mainly when they are used as sole diets. The presence of these compounds forms insoluble complexes with essential minerals, proteins and carbohydrates, lowering the nutritive value of the product (El Shaer, 2010). Therefore, appropriate mixing of these species, based on their complementary roles, could dilute the negative effects of these ANFs, thus improves animal performances (Masters *et al.*, 2001, Abd El-Rahman, 2003, ICBA, 2006, Anon., 2009 and El Shaer, 2010).

The insufficient information about the effect of feeding salt-tolerant plants on the physiological functions was the motive of this study to focus on liver and kidney functions and body weight changes in addition to the metabolic state of Barki sheep fed on a mixture of *Atriplex nummularia*, *Sorghum bicolor* and *Pearl millet*.

#### 2. Materials and Methods

This study was carried out at South Sinai Station (Ras Sudr) which belongs to Desert Research Center, Ministry of Agriculture and Land Reclamation, South Sinai Governorate, Egypt. This study was conducted so as to monitor physiological and biochemical parameters changes of Barki sheep as a result of feeding a sun- dried chopped mixture of *Atriplex nummularia* (50%)+ *Sorghum bicolor* (25%) + *Pearl millet* (25%) for three months.

Eighteen Barki ewes, aged 3- 3.5 years old with an average body weight 28.50±1.02 kg, were divided into two equal groups. The first group (G1) was fed berseem hay (Trifolium alexandrinum, 4th cut) and served as control, while the second group (G2) was fed a mixture of Atriplex nummularia, sorghum bicolor and pearl millet at rate of 50, 25 and 25 %, respectively. Both groups were provided by concentrate feed mixture (CFM). A11 experimental animals were fed their nutrient requirements according to Kearl (1982). The chemical composition of Atriplex nummularia, sorghum bicolor and pearl millet and berseem hay (Table 1) was determined according to A.O.A.C. (1985).

Table (1): Chemical composition of feed mixture of the two experimental diets fed to Barki ewes (Dry matter basis)

	DM	OM	СР	CF	EE	NFE	Ash
Berseem hay	100.00	88.95	12.22	28.55	1.18	47.00	11.05
Atriplex nummularia	94.27	78.27	12.07	20.15	10.28	35.78	21.72
Sorghum bicolor	92.17	86.29	9.68	22.99	6.31	47.31	13.71
Pearl millet	91.93	83.81	9.18	21.40	8.25	44.98	16.19
Salt tolerant mixture*	92.59	83.41	10.91	19.72	13.68	39.10	16.59
Concentrate feed mixture	93.76	89.20	16.72	12.78	4.11	55.59	10.80
	CD	1	a				

DM, dry matter CP, crude protein CF, crude fiber

EE, ether extract NFE, nitrogen free extract OM, organic matter

\*, Atriplex nummularia (50%)+ Sorghum bicolor (25%) + Pearl millet (25%)

Fresh water was available twice daily over the experimental period. The experimental animals were housed in a semi-closed pen, roofed with and walled in four directions with concrete. All animals were weighed biweekly up to end of the experiment.

Just before offering ration and water, jugular blood samples were withdrawn into clean heparinized tubes and centrifuged for 30 minutes at a speed of 3000 r.p.m. for plasma separation.

Assay of total proteins (TP) and albumin (AL) was carried out according to Biuret method after Gornal et al. (1949) and Doumas et al. (1971), respectively. Values of globulin (GL) were calculated by subtracting the value of albumin from the total protein whereas A/G ratio was calculated according to results of albumin and globulin. Plasma total lipids (TL), total cholesterol (CHO) and glucose (GLU) concentrations were determined according to Schmillet (1964), Roeschlau et al. (1974) and Tietz (1986), in order. Alanine (ALT) and aspartate (AST) amino transferases levels were analyzed according to Reitman and Frankel (1957), while alkaline phosphatase (ALP) was determined according to Belfield and Goldberg (1971). Plasma urea and creatinine concentrations as indicators for kidnev function were determined using biodiagnostic kits according to Fawcett and Soctt (1960) and Schirmeister *et al.* (1964), respectively. Concentrations of sodium (Na) and potassium (K) were determined according to Trinder (1951) and Sunderman and Sunderman (1958), respectively.

Direct radioimmunoassay technique (RIA) was performed for plasma insulin, triiodothyronine  $(T_3)$ , thyroxine  $(T_4)$  and aldosterone hormones using ready antibody coated tubes kits manufactured by Immunotech, Beckman Counter Company, France.

Experimental data were analyzed using General Linear Model Procedure (SAS, 2004).

#### **3. Results and Discussion** Live body weight:

The results revealed that live body weight was not affected significantly by feeding salt tolerant fodder crops mixture (*Atripex halimus*, *Sorghum bicolor* and *Pearl millet*). The animals fed control diet (Control group) had insignificant higher final body weight and daily gain as compared to their counterparts of treatment (Table 2). These results underlined the potentiality of such salt tolerant fodder mixture to fulfill the animal requirements to maintain their body weight. In similar, working on Barki lambs, Shaker *et al.* (2008) reported that feeding mixture of atriplex and acacia resulted in non-significant differences in body weight with control group. Furthermore, Zarkawi *et al.* (2005) reported that Awassi ewes fed *Sesbania aculeate* which grown on salty soil and irrigated by saline water had comparable live body weight of ewes for control group. This slight reduction in daily gain of animals fed on *Atriplex nummularia*, *Sorghum bicolor* and *pearl millet* mixture might be attributed to the tannins content which was found to reduce the digestibility of protein and dry matter (Priolo *et al.*, 2000 and El-Shaer, 2010). A combination of reduced intake and low true digestibility of protein cause the negative effect of tannins on growth rate (Ben Salem *et al.*, 2010, El- Shaer *et al.*, 2005 and El- Shaer 2010). These results were in agreement with those reported by Azamel (1997) and Nasr *et al.* (2002). Badawy *et al.* (2002) found that lambs fed fresh *Atriplex halimus* or *Acacia saligna* had lower average daily gain than the control group. Additionally, Shehata *et al.* (1988) reported that body weight gain was reduced in sheep and goats fed *Atriplex nummularia*.

Item	G1	G2	± SE	Overall mean
Initial body weight <sup>NS</sup> (kg)	28.39	28.61	±1.49	28.50±1.02
Final body weight <sup>NS</sup> (kg)	36.31	35.83	±2.03	36.07±1.39
Body weight changes <sup>NS</sup> (kg)	7.92	7.22	±1.03	7.57±0.71
% of initial body weight <sup>NS</sup>	28.51	25.90	±3.11	26.71±2.14
Average daily gain <sup>NS</sup> (gm)	52.81	48.15	±10.49	50.48±8.91
NC non significant				

NS, non-significant

G1, animals fed berseem hay + CFM (Control group)

G2, animals fed a mixture of Atriplex nummularia (50%)+ Sorghum bicolor (25%)+ Pearl millet (25%)+ CFM

#### **Total proteins concentration:**

Animals fed salt tolerant plants mixture of Atriplex nummularia, Sorghum bicolor and Pearl millet had insignificantly lower mean values of total proteins (4.80%), albumin (7.32%), globulin (2.17%) and albumin/ globulin ratio (19.94%) in comparison with control ones (Table 3). These results were in agreement with those reported by Askar (1998), Badawy et al. (2002), Abdel- Halim (2003) and Shaker et al. (2008). This decrease of TP, Al, Gl and A/G ratio values in treated animals (G2) might be attributed to the high content of tannins in such salt tolerant plants where tannins were reported to decrease the digestibility of crude protein (Muller et al., 1989, Reed et al., 1990 and Mahmoud, 2001). Coles (1986) found that poor absorption of dietary constituents from the

intestinal tract leads to hypoproteinemia. On the other hand, higher salt intake might be another reason to introduce such decrement in TP, Al, Gl and A/G ratio levels (Weeth and Haverland, 1961 and Badawy, 1999). In accordance, Tata and Widnell (1966) observed a decrease in total protein concentration due to drinking saline water which might possibly reduce hepatic synthesis of RNA which in turn depressed the incorporation of amino acids for protein synthesis.

From another point of view, the slight decrease in total proteins fraction could be attributed to the lower crude protein recorded in ration of salt tolerant plants mixture (Table 1). Yousef and Zaki (2001) and Shahen *et al.* (2004) reported a positive correlation between dietary protein and plasma protein concentrations.

 Table (3): The mean values of total proteins, albumin and globulin concentrations and albumin/ globulin ratio of Barki ewes as affected by feeding salt tolerant plants mixture

Tatlo of Darki ewes as affected by feeding sait tolerant plants mixture							
Item	G1	G2	± SE	Change	Overall		
Total protein <sup>NS</sup> (g/dl)	5.90	5.63	$\pm 0.370$	- 0.27 (4.80%)	$5.77 \pm 0.264$		
Albumin <sup>NS</sup> (g/dl)	3.08	2.87	$\pm 0.234$	- 0.21 (7.32%)	$2.98 \pm 0.214$		
Globulin <sup>NS</sup> (g/dl)	2.82	2.76	$\pm 0.306$	- 0.06 (2.17%)	$2.79 \pm 0.198$		
A/ G ratio <sup>NS</sup>	1.48	1.24	$\pm 0.303$	- 0.21 (16.94%)	$1.36 \pm 0.242$		

NS, non-significant

G1, animals fed berseem hay + CFM (Control group)

G2, animals fed a mixture of Atriplex nummularia (50%)+ Sorghum bicolor (25%)+ Pearl millet (25%)+ CFM

#### **Glucose concentration:**

Feeding salt tolerant plants lowered (P<0.01) plasma glucose concentration by 17.88% which might be attributed to the tannins content in these plants. Tannins were reported to reduce digestibility of protein and carbohydrate by inhibiting digestive enzymes and/ or by altering permeability of the gut wall (Streeter *et al.*, 1993). Moreover, Ortiz *et al.* (1993) reported that tannins could adversely influence digestibility and

absorption of nutrients such as proteins and amino acids, carbohydrates and lipids and also the activity of digestive enzymes.

On the other hand, high salt in these salt tolerant forages might reduce glucose concentration in group two (G2). This theory would be confirmed by results reported by Assad *et al.* (1997) when working on rams receiving saline water. In consistence, Kewan (2003) reported that the

glucose level in camels fed acacia plus atriplex was lower than that of control groups.

#### **Cholesterol concentration:**

Animals fed salt tolerant plants mixture had lower (P<0.01) cholesterol concentration by 9.43% than the control ones (Table 4). Similar results were reported by Shaker *et al.* (2008) in Barki lambs and Fayed (2009) in Barki rams. This significant reduction in plasma cholesterol concentration could be due to the presence of tannins which interfere in lipids digestibility by complexing with fatty acids (Romero *et al.*, 2000), decreasing cholesterol absorption and increasing fat excretion (Bravo *et al.*, 1993). On the other hand, this decrease in cholesterol levels might be owing to saponins contents (Salem *et al.* 2004, El- Shaer *et al.*, 2005, Fayed *et al.*, 2010 and Ben Salem *et al.*, 2010). Saponins were found to lower serum cholesterol levels in animals (Matsuura, 2001). The hypocholesterolaemic action of saponins was previously reported (Francis *et al.*, 2002) through inhibiting the cholesterol absorption causing reduction in plasma high- density lipoprotein cholesterol fraction (Morehouse *et al.*, 1999) and/ or delaying the intestinal absorption of dietary fat by inhibiting pancreatic lipase activity (Han *et al.*, 2000). Finally, through low digestibility of fats in ruminants which is limilleted by the lack of emulsifying agents in the rumen (Cheeke, 1999).

 Table (4): The mean values of glucose, cholesterol, total lipids, insulin and thyroid hormones concentrations of Barki ewes as affected by feeding salt tolerant plants mixture

concentrations of barki ewes as affected by feeding san tolerant plants mixture							
Item		G1	G2	$\pm$ SE	Change	Overall	
Glucose <sup>**</sup>	(mg/l)	46.93a	39.81b	± 1.307	-7.12 (17.88%)	43.37±1.596	
Cholesterol **	(mg/dl)	102.57a	93.73b	$\pm 1.978$	-8.84 (9.43%)	98.15±1.544	
Total lipids **	(g/l)	3.03a	2.54b	$\pm 0.068$	-0.49 (19.29%)	$2.78 \pm 0.106$	
Insulin <sup>**</sup>	(µU/ml)	12.56a	9.24b	±0.136	-3.32 (35.90%)	10.90±1.703	
T3 **	(pg/ml)	5.05a	3.87b	±0.053	-1.18 (30.49%)	4.46±1.762	
T4 **	(ng/ml)	11.99	8.18	±0.65	-3.81(46.57%)	10.09±1.445	

\*\*, p<0.01

In the same row, any two means in a certain item having different letters differ significantly.

G1, animals fed berseem hay + CFM (Control group)

G2, animals fed a mixture of Atriplex nummularia (50%)+ Sorghum bicolor (25%)+ Pearl millet (25%)+ CFM

#### **Total lipids concentration:**

Lipids virtually have an important role in all aspects of biological life, serving as hormones precursors, aiding in digestion, providing energy storage and metabolic fuel and acting as functional and structural components in biomembranes (Tietz, 1990).

Plasma total lipids (TL) levels had the same trend of glucose and cholesterol levels where treated group had lower (P<0.01) total lipids values than control by 19.29 % (Table 4). These lower values of TL could be due to the reported effects of tannins and saponins on fat metabolism (Cheeke, 1999, Han *et al.*, 2000, Salem *et al.* 2004, El- Shaer *et al.*, 2005 and Fayed *et al.*, 2010). Similar results were reported by Abdelhameed *et al.* (2004& 2006) who reported a significant decrease in total lipids and lipid fraction as a result of feeding on fresh saltbush forage plants. However, Shaker *et al.* (2008) demonstrated that feeding a mixture of atriplex and acacia led to a non-significant decrease in total lipids of Barki lambs.

#### Insulin concentration:

The present data revealed that insulin level decreased (P<0.01) in animals fed salt tolerant forages (G2) by 35.90% in comparison to their counterparts of control group (Table 4). Similar results were reported by Pearce *et al.* (2008) who reported that there was a significant effect of diet on plasma insulin concentration (P< 0.05) where animals fed saltbush diet had significantly lower

plasma insulin concentration than sheep fed the control diet. In agreement, Digby *et al.* (2010 a&b) and Blache *et al.* (2007) suggested that decreasing in secretion of insulin could be in response to high salt which affect feed intake affecting the energy balance, defined as the difference between energy expenditure and the sum of energy intake and energy reserves, pushing towards a negative value. Moreover, in humans, plasma insulin decreased by up to 47% in response to an increase in salt intake, via saline infusion or consumption of salt in diet (Goodfriend *et al.*, 1991).

#### Thyroid activity:

Thyroid hormones play a major effect on growth and development of animal. These hormones are in correlation with metabolism of protein, carbohydrate, fat (Zanouny *et al.*, 2013). In addition, normal function of the control nervous system is very dependent on normal output of  $T_4$ . Therefore, reduced thyroid secretion will ultimately results in reduced metabolism of such nutrients (Trenkle, 1978 and Hart *et al.*, 1981).

The obtained results of triiodothyronine  $(T_3)$ and thyroxine  $(T_4)$  hormones declared that the salt tolerant forages mixture group (G2) showed significant lower (P<0.01) mean values comparing to the control group by 30.49 and 46.57% for  $T_3$ and  $T_4$ , respectively (Table 4). This decrement might be attributed to the high content of salt. Metwally (2001) reported that saline water treatment in camels decreased both of  $T_3$  and  $T_4$  suggesting that to the decreased of feed intake, so that the metabolism process decreased. In addition, Ayyat *et al.* (1991) and Ahmed (1996) reported that drinking saline water depressed  $T_3$  and  $T_4$  levels in rabbits.

#### Liver function:

Most plasma enzymes come from different tissues of animal. Its activity level had a relation with metabolism and functional status of certain organs. Body's ability to adjust and adapt depends on the function of tissues and organs largely (Shi-Gang *et al.*, 2010). Concentrations of its enzymes alanine amino transferase (ALT), aspartate amino transferase (AST), alkaline phosphatase (ALP) and gamma glutamyl transferase (GGT) are those conventionally used for diagnosing hepatic damage. Transaminases are widely distributed in plasma, bile, cerebrospinal fluid and saliva but none is found in urine unless a kidney lesion is present (Norbert, 1987).

Alanine amino transferase is particularly useful in measuring hepatic necrosis and increased in serum when cellular degeneration or destruction occurs (Lessard *et al.*, 1986).

The levels of alanine amino transferase (ALT) of salt tolerant forages mixture group exceeded (P< 0.05) their counterparts of control ration by 17.22% (Table 6) which might be

attributed to the high tannins in these shrubs (Tripathy *et al.*, 1984) or to the high content of salt as reported by Ibrahim (1995) in goats, Hussein (1987) in sheep, Ibrahim (2001) in camels and Ibrahim *et al.* (1991) in male buffalo- calves. Consistently, Shaker *et al.* (2008) found that the activity of ALT was higher (P<0.01) in animals fed fresh or silage form of atriplex and acacia mixture comparing to control group. Badawy *et al.* (2002) noticed that feeding lambs on fresh acacia increased significantly the level of ALT by 14.1 %.

Aspartate amino transferase (AST), which is extrahepatic present in tissues including myocardium and kidney, can be used as a good indicator of hepatic injury of sheep (Lessard et al., 1986). The mean values of AST exhibited similar trend as ALT where animals fed on the salt tolerant forages mixture (G2) got higher insignificant activity of AST (10.14%) than those recorded for the control group (Table 5). This slight elevation of AST activity could be due to high contents of tannins in these shrubs might be the main reason of AST elevation (Badawy et al., 2002 and Tripathy et al., 1984). In a harmony with the present results, Shaker et al. (2008) found a significant increase in activity of AST in lambs fed on fresh atriplex and acacia mixture as compared to their control group.

Table (5): The mean values of alanine amino transferase (ALT), aspartate amino transferase (AST), alkaline phosphatase (ALP), urea and creatinine concentrations of Barki ewes as affected by fooding solt to logont plonts minutes

feed	ling salt tole	rant plants i	mixture			
Item	L	G1	G2	± SE	Change	Overall
ALT <sup>*</sup>	(IU/l)	23.69	27.77	± 1.24	+4.08 (17.22%)	25.73+1.126
AST <sup>NS</sup>	(IU/l)	26.04	28.68	$\pm 3.355$	+2.64 (10.14%)	27.36+2.581
ALP **	( <b>IU/l</b> )	166.89	151.81	$\pm 1.856$	-15.08 (9.93%)	159.25+3.940
Urea **	(mg/dl)	42.73	36.78	$\pm 0.802$	-5.95 (16.17%)	39.75+1.218
Creatinine *	(mg/dl)	1.06	1.40	$\pm 0.093$	+0.34 (32.07%)	$1.22 \pm 0.073$
*, P<0.05			**, p<0.01		NS, non	significant

In the same row, any two means in a certain item having different letters differ significantly.

G1, animals fed berseem hay + CFM (Control group)

G2, animals fed a mixture of Atriplex nummularia (50%)+ Sorghum bicolor (25%)+ Pearl millet (25%)+ CFM

Silanikove et al. (1996) demonstrated that alkaline phosphatase is used to detect the bile obstruction (i.e. mild and progressive damage to the liver). The present results of alkaline phosphatase (ALP) activity demonstrated that animals fed salt tolerant forages mixture diet had lower (P< 0.01) values than those of control group (Table 5). This decline in alkaline phosphatase activity might be attributed to the existence of antinutritional factor in such salt tolerant plants. Tannins, which react with this enzyme, were reported to reduce its activity (Horigome et al., 1988 and Abde- Halim, 2003). In addition, excess of oxalate is an inhibitor for that liver enzyme (McComb et al., 1979). Abu- Zanat et al. (2003) reported that Atriplex spp. contain high concentrations of oxalate which can be toxic to livestock. In agreement, Shaker et al. (2008)

reported that ALP levels decreased (P<0.01) in animals fed either fresh or silage forms of acacia and atriplex mixture.

# **Kidney function:**

Urea and creatinine are the two main nitrogenous compounds eventually excreted by kidney. Accordingly, any change of their concentration would reflect impaired glomerular filtration and/ or insufficiency of renal tubules (Coffin, 1955, Miller, 1966 and Kaneko, 1989). Furthermore, urea is the major nitrogen- containing metabolic product of protein catabolism accounting for more than 75% of the non- protein nitrogen excreted. Also, production is also too dependent on several non- real variables such as a diet and hepatic synthesis.

The present results revealed that lower (P<0.01) mean concentration of urea was observed

in animals fed salt tolerant forages mixture diet (G2) in comparison with control group (Table 5). These results were in agreement with those of Shaker et al. (2008) when Barki lambs fed atriplex and acacia mixture. This reduction in urea concentrations (16.17 %) could be owing to the presence of tannins, which reduce the ruminal proteins degradation (Mashudi et al., 1997). In accordance, Waghorn et al. (1994), Patra et al. (2002) and Cook et al. (2008) reported that plasma urea nitrogen was lowered in sheep and goats fed legumes that high in tannins. Pearce et al. (2008) reported that treatment had a significant effect on plasma urea concentrations (P < 0.001) where sheep fed the control diet had significantly higher plasma urea concentrations than the saltbush group.

Azamel (1997) found that the blood urea was significantly lowered in two groups of growing lambs fed *Atriplex nummularia* in comparison with control groups. However, Badawy *et al.* (2002) found that animals fed fresh *Atriplex nummularia* had high urea values comparing with control group.

Feeding a mixture of Atriplex nummularia, Sorghum bicolor and *Pearl millet* forages (G2) increased (<0.05) the mean values of creatinine comparing to the control group (Table 5) in a reverse trend with that of urea results. Consisitenty, Badawy et al. (2002) and Shaker et al. (2008) found that animals fed fresh atriplex or acacia had higher mean values of creatinine than their counterparts fed on control ration. This increment in creatinine levels could be as a result of antinutritional factors and/ or high salt in such salt tolerant plants. Consistently, Zhu et al., (1992) and Zhu and Filippich (1995) noticed that plasma creatinine was significantly increased in sheep dosed by 0.1 g tannic acid/ kg body weight intraperitoneally. Clark and Clark (1978) reported that feeding on Atriplex species caused destruction of renal nephrons since such plants contain high amounts of oxalates. Cheeke (1995) reported that

the common effect of oxalate is to cause kidney damage owing to blocking of tubules by crystals of calcium oxalate. This does not necessarily cause death, but the kidney damage remains and subsequently ingestion of oxalate- containing plants may have fatal results.

On the other hand, high content of salt in such salt tolerant forages mixture might be the clue to understand the increment observed in creatinine level. Receiving salt load resulted in alteration in kidney function of cattle's (Nelson *et al.*, 1995 and Weeth and Lesperance, 1965). Hussein (1987) reported that drinking saline water caused highly significant increase in sheep plasma creatinine. Assad *et al.* (1989) noted a significant elevation of creatinine in serum of the ewes and their lambs received diluted sea water (1.3% TDS). Similarly, Abdel- Gawad (1993) demonstrated that drinking saline water by sheep, goats and camels increased plasma creatinine.

#### **Blood electrolytes:**

Minerals play an important role in the regulation of body fluids, acid base balance and metabolic processes (Milne, 1996).

The present findings of sodium (Na) level values demonstrated that animals fed salt tolerant forages (G2) had higher (P<0.01) value than control group (Table 7). The increase in sodium concentration might be due to atriplex which contains high content of sodium (6.45%) and chloride (7.03%) as reported by Mohamed (1996). Ben Salem et al. (2010) reported that most of the salt in oldman saltbush is sodium chloride and potassium chloride. Similar results were obtained by Rasool et al. (1996) in sheep. Badawy et al. (2002), Nasr et al. (2002) and Shaker et al. (2008) reported that serum Na<sup>+</sup> levels increased significantly in animals fed atriplex. Rasool et al. (1996) reported that there was significant (p < 0.01) increase in the sodium contents of blood serum at high levels of saltbush containing diet.

Table (6): The mean values of sodium, potassium, aldosterone concentrations, Na/ K and of Na index of Barki ewes as affected by feeding salt tolerant plants mixture

Dark twes as anceted by recurd sait toterant plants inixture							
Item		G1	G2	± SE	Change	Overall	
Na <sup>**</sup>	(ppm)	6150	7183	±36.955	+1033 (16.80%)	6666.67±117.286	
K **	(ppm)	122.67	170.67	$\pm 2.298$	+48 (39.13%)	146.66±6.253	
Na/ K <sup>**</sup>		50.47	42.70	±1.137	-7.77 (18.20%)	46.57±1.246	
Na/ Na+ K **		0.98	0.97	$\pm 0.0004$	-0.01 (1.03%)	0.97±0.473	
Aldosterone **	(pg/ ml)	614.58	800.39	±12.71	+185.81 (30.23%)	706.85±69.394	

\*\*, p<0.01

In the same row, any two means in a certain item having different letters differ significantly.

G1, animals fed on Berseem hay + CFM (Control group)

G2, animals fed on a mixture of Atriplex nummularia (50%)+ Sorghum bicolor (25%)+ Pearl millet (25%)+ CFM

Concerning of potassium concentration, animals fed salt tolerant forages mixture (G2) had higher (P<0.01) mean plasma potassium concentration value than the control group (Table 6). In agreement, Badawy *et al.* (2002) and Shaker *et al.* (2008) reported that animals fed on atriplex

had higher  $K^+$  concentration. This pattern of potassium was in complete harmony with that of sodium. Wegner and Schuh (1974) and Wichell (1976) noted a relationship between increasing Na<sup>+</sup> intake; which is already high in such salt tolerant forages, and high concentration of K<sup>+</sup> in plasma.

Rasool *et al.* (1996) reported a significant (P < 0.01) increase in the potassium contents in the blood from medium to very high levels of the saltbush in the diet Sodium and potassium are known to be found in high concentrations in halophytes and salt tolerant forages (NRC, 1975, Kearl, 1982 and El Shaer, 1981), however, excess intake of these electrolytes is accompanied by their excess excretion through the kidneys (Neathery, 1980).

The obtained results revealed that values of Na/K ratio and Na, K index values were lower (P<0.01) in treated group than control group (Table 6). In agreement, Shaker *et al.* (2008) reported that animals fed fresh form of atriplex and acacia mixture had insignificant Na/K ratio and Na, K index values. This reduction in Na/K ratio and Na, K index values might be attributed to the higher level of potassium in plasma (Table 6).

# Aldosterone concentration:

Results of aldosterone level showed that animals fed salt tolerant forages mixture (G2) had higher (P<0.05) aldosterone level than that of control group (Table 6). This increment in aldosterone concentration might be attributed to the high content of salt in such desert shrubs. It is well known that aldosterone secretion increases hen blood potassium increased and this is evident in the present results (Table 6). In accordance, Shaker et al. (2008) reported that feeding atriplex and acacia mixture increased aldosterone concentration. Furthermore, Hamdi et al. (1982) clarified the association of aldosterone and  $Na^+/Na^+ + K^+$  index in the blood suggesting that it might be attributed to an increase in the aldosterone release in response to drinking the diluted sea water. In addition, demonstrated Aboulnaga (1987) that the aldosterone secretion and concentration of sodium and potassium in urine and blood plasma would be changed when the ratio of sodium and potassium intake was changed.

# Conclusion

From aforementioned results, it could be concluded that introducing salt tolerant plants *Atriplex nummularia, sorghum bicolor* and *pearl millet* for sheep could be an attempt to reduce feed shortage for livestock in arid and saline conditions prevailed in Southern Sinai, Egypt and to increase the utilization of the available unpalatable salt tolerant plants as animals feeds specially for sheep and goats.

# Corresponding author Shaker, Y.M.

# Associate Professor

Animal and Poultry Physiology Department, Animal and Poultry Production Division, Desert Research Center, Mataria, Cairo, Egypt vousrimshaker@vahoo.com

# Acknowledgment

The author would thank Prof. Dr. El-Shaer, the coordinator and Prof. Dr. Badawy, the key

person of the regional project titled "Adaptation to climate changes in WANA marginal environments through sustainable crop and livestock diversification" which is funded by International Center for Biosaline Agricultural (ICBA), UAE for their finical support to achieve this work.

# References

- A.O.A.C. (1985). Official Methods of Analysis. Association of Official Analytical Chemistry. Washington, D. C., USA.
- Abd El-Rahman, H. H. (2003). Constraints and possibilities for their alleviation to improve utilization of desert natural range plants for grazing ruminants. Ph.D. Thesis, Faculty of Agriculture, Cairo University, Cairo, Egypt.
- Abdel- Gawad, M. M. H. (1993). Some nutritional aspects of camels offered saline water. M. Sc. Thesis, Fac. of Agric., Cairo Univ.
- Abdel- Halim, A. M. (2003). Studies of some antinutritional factors affecting utilization by ruminants. Ph. D. thesis, Faculty of Science, Ain Shams University.
- Abdelhameed, A, A. E.; Shawket, M. S. and Hafez, I. M. (2006). Physiological studies on the effects of feeding salt plants in ewes under semi- arid condition. Society of Physiological Science and Their Applications, Fourth Scientific Conference, 29- 30 July, 2006, pp. 113-132.
- Abdelhameed, A. A. E.; Shawket, M. S. and Kewan, Kh. Z. E. (2004). Effect of feeding *Atriplex halimus* and *Acacia saligna* on some biochemical parameters of growing camels under desert environmental conditions. Society of Physiological Science and their Application, Second Scientific Conference, July.
- Aboulnaga, A. I. A. (1987). The role of aldosterone in improving productivity of heat stressed farm animals with different techniques. Ph. D. Thesis, Fac. of Sci., Zagazig Univ., Egypt.
- Abu-Zanat, M. M.; Al-Hassanat, F. M.; Alawi, M. and Ruyle, G. B. (2003). Oxalate and tannins assessment in *Atriplex halimus L*. and A. nummularia L. J. Range Manage 56, 370–374.
- Ahmed, M. H. (1996). Effect of saline drinking water on productive performance of rabbits. M. sc. Thesis, Fac. Of Agric., Zagazig Univ., Egypt.
- Al Khalasi, S. S.; Mahgoub, O.; Kadim, I. T.; Al-Marzouqi, W. and Al-Rawahi, S. (2010). Health and performance of Omani sheep fed salttolerant sorghum (*Sorghum bicolor*) forage or Rhodes grass (*Chloris gayana*). Small Ruminant Research 91 (2010) 93–102.
- Anon (2006). Electronic Conference on Salinization: Extent of Salinization and Strategies for Salt-Affected Land Prevention and Rehabilitation, 6 February–6 March 2006. Organized and coordinated by IPTRID

(International Programme for Technology and Research in Irrigation and Drainage), FAO.

- Anon (2009). Introduction of salt-tolerant forage production systems to salt-affected lands in Sinai Peninsula in Egypt: A pilot demonstration project. Final Report, DRC, Egypt—ICBA, UAE.
- Askar, A. R. T. (1998). Effect of feeding some halophytic plants on nutritional and reproductive performance for growing sheep in Sinai. M. Sc. Thesis Faculty of Agriculture, Ain Shams University.
- Assad, F.; Bayoumi, M. T; El- Begowyi, M. and Deeb, S. (1989). Pathological changes in ewes and their lambs drinking sea water. Egypt. J. Comp. Pathol. Clin. Pathol, 2 (2): 61-76.
- Assad, F. M.; Nasser, A. M.; Hussein, N. and Abdelmageed, S. M. (1997). Effect of saline water on some biochemical parameters in sheep/ Egypt. J. Applied Sci., 12: 11- 31.
- Ayyat, M. S.; Habeeb, A. A. and Bassumg, S. M. (1991). Effect of water salinity on growth performance, carcass traits and some physiological aspects of growing rabbits in summer season. Egypt. J. of Rabbit Science, 1: 21-34.
- Azamel, A. A. (1997). Physiological responses and daily gain of growing Barki lambs fed on *Atriplex halimus*, a natural desert shrubs. Egypt. J. Appl. Sci., 12 (2): 1-9.
- Badawy, M. S. M. (1999). Digestive function and heat regulation in Saidi sheep. M. Sc. Thesis, Assiut University, Egypt.
- Badawy, M. T.; Gawish, H. A. and Younis, A. A. (2002). Some physiological responses of growing Barki lambs and Baladi kids fed natural desert shrubs. International Symposium on Optimum Resources Utilization in Salt -Affect Ecosystems in Arid and Semi- arid Regions. Cairo, 8- 11, April, 496- 503.
- Belfield, A. and Goldberg, D. M. (1971). Revised assay for serum phenyl phosphayatase activity using 4- amino- antipyrine. Enzyme. 12: 561-573.
- Ben Salem, H.; Norman, H.C.; Nefzaoui, A.; Mayberry, D.E.; Pearce, K.L. and Revell, D.K. (2010). Potential use of oldman saltbush (*Atriplex nummularia Lindl.*) in sheep and goat feeding. Small Ruminant Research, v.91, p.13-28.
- Blache, D.; Grandison, M. J.; Masters, D. G.; Dynes, R. A.; Blackberry, M. A.; Martin, G. A. (2007). Relationships between metabolic endocrine systems and voluntary feed intake in Merino sheep fed a high salt diet. Aust. J. Exp. Agric., 47: pp. 544–550.
- Bravo, L.; Manas, E. and Calixto, F. S. (1993). Dietary non extractable condensed tannins as indigestible compound: Effect on fecal weight

and protein and fat excretion. J. Sci. Food Agric., 63: 63-68.

- Cheeke, P. R. (1999). Actual and potential applications of yucca schidigera and Quillajasponaria saponins in human and animal nutrition. Proceedings of the American Society of Animal Sci., pp: 1-10.
- Cheeke, P. R. (1995). Endogenous toxins and mycotoxins in forage grasses and their effect on livestock. J. Anim. Sci., 73: 909- 918.
- Clark, E. G. C. and Clark, M. L. (1978). Veterinary toxicology. Bailliere Tindall, London.
- Coffin, D. L. (1955). Manual of veterinary clinical pathology. 3<sup>rd</sup> ed., Cornell Univ. Press.
- Coles, E. H. (1986). Veterinary Clinical Pathology.
  4<sup>th</sup> Ed. Sunders W. B. Co., Philadelphia, London.
- Cook, R. W.; Scott, C. B. and Hartmann, F. S. (2008). Short-term Mesquite pod consumption by goats does not induce toxicity. Rangeland Ecology & Management 61 (5): 566–570.
- Digby, S. N.; Blache, D.; Masters, D. G. and Revell, D. K. (2010a). Responses to saline drinking water in offspring born to ewes fed high salt during pregnancy. Small Ruminant Research 91: pp. 87–92.
- Digby, S. N.; Masters, D. G.; Blache, D.; Hynd, P. I.; Revell, D. K. (2010b). Offspring born to ewes fed high salt during pregnancy have altered responses to oral salt loads. Animal, 2010 4(1): 81- 88.
- Doumas, B. T.; Watson, W. A. and Biggs, H. G., (1971). Albumin standards and the measurement of serum albumin with bromcresol green. Clinca Chemica Acta, 31: 87-96.
- El Shaer, H. M. (2010). Halophytes and salttolerant plants as potential forage for ruminants in the Near East region. Small Ruminant Research 91: pp. 3–12.
- El Shaer, H. M. (1981). A comparative nutritional study on sheep and goats grazing in Southern Sinai desert range with supplements. Ph. D. Thesis, Fac. of Agric. Ain Shams Univ., Cairo, Egypt.
- El Shaer, H. M.; Ali, F. T.; Morcos, N. Y. S.; Emam, S. S. and Essawy, A. M. (2005). Seasonal changes of some anti-nutritional factors contents of some halophytic shrubs and the effect of processing treatments on their utilization by sheep under desert conditions of Egypt. Egyptian J. Nutr. Feeds, 8 (1): Special Issue: 417-431.
- Fawcett, J. K. and Soctt, J. E. (1960). A rapid precise method for the determination of urea. J. Cline., Path. 13: 156.
- Fayed, A. M. (2009). In vitro and In vivo evaluation of biological treated salt plants. American-Eurasian J. Agric. & Environ. Sci., 6 (1): 108-118.

- Fayed, A. M.; El- Essawy, A. M.; Eid, E.Y.; Helal, H. G.; Abdou, A. R. and El Shaer, H. M. (2010). Utilization of berseem and atriplex for feeding sheep under saline conditions of South Sinai, Egypt. Journal of American Science, 6 (12): 1447- 1461.
- Francis, G.; Kerem, Z.; Makkar, H. P. S. and Becker, K. (2002). The biological action of saponins in animal system: a review. British J. Nut., 88: 587-605.
- Goodfriend, T. L.; Ball, D. L.; Weinberger, M. H.; Moore, T. J.; Weder, A. B. and Egan, B. M. (1991). Salt loads raise plasma fatty acids and lower insulin. Hypertension 17, 958–964.
- Gornal, A. C.; Bardawill, C. J. and David, M. M., (1949). Kit protein Egyptian American Co. for Laboratory Services. J. Biol. Chem., 177: 751-755.
- Hamdi, H.; Abdel- Rahman, Y.; Malek, A.; El-Bagoumy, I.; Ibrahim, W. Z. and Makasem, F. (1982). Fundamentals of human physiology. Kidney and body fluid. Vol (7). Altas Press, Cairo.
- Han, L. K.; Xu, B. J.; Kimura, Y.; Zheng, Y. N. and Okuda, H. (2000). Platycodi radix affects lipid metabolism in mice with high fat diet induced obesity. J. Nutrition, 130: 2760- 2764.
- Hanafy, M.A.; Fahmy, A.A.; Farghaly, M.S. and Afaf, A.; El Sheref, (2007). Effect of using some fodder plants in diets on goats' performance under desert conditions of Sinai. Egyptian J. Nutr. Feeds 10, 151–163.
- Hart, I. G.; Morant, S. V. and Roy, H. B. (1981). A note on the variability of hormone concentration in twice-weekly blood samples taken from heifers calves during the first 110 days of life. J. Anim. Prod. 32: 215.
- Horigome, T.; Kumar, R.; Okamoto, K. (1988). Effect of condensed tannins prepared from leaves of fodder plants on digestive enzymes *in vitro* and in the intestine of rat. British J. of Nutrition 60: 275.
- Hussein, N. M. (1987). Hematological studies on sheep drinking salty water. M. Sc. Thesis, Faculty of Science, Al- Azhar University, Cairo, Egypt.
- Ibrahim, I. A.; Abd- Alla, O. A. and El- Nahia, A. M. (1991). Biochemical and hematological studies in male buffalo- calves drinking well's water. Egypt. J. Comp. Pathol. Clin. Pathol., 4 (2): 237-244.
- Ibrahim, N. H. M. (2001). Studies on some physiological and behavioural aspects in camels. M. Sc. Thesis. Fac. of Agric., Minufiya Univ.
- Ibrahim, S. M. N. (1995). Clinicopathological studies in goats drinking salty water under desert conditions. M. V. Sc., Cairo Univ.

- ICBA(2006). Biosalinity news. Newsletter of the International Center of Biosaline Agriculture (ICBA) 9 (July (2)), 2006.
- Kaneko, J. J. (1989). Clinical Biochemistry of Domestic Animals.4th Edn. Academic Press.
- Kearl, I. C. (1982). Nutrients requirements in developing countries. Utah Agric. Exp. Stat., Utah State Unviersity, Logan, USA.
- Kewan, Kh. Z. M. (2003). Studies on camel nutrition. Ph. D. Thesis, Fac. of Agric., Alexandria University.
- Lessard, P.; Wilson, W. D. and Olander, H. J. (1986). Clincopathologic study on horses surviving P. A. (*Senecio vulgaris*) toxicosis. Aim. J. Vet. Res., 47: 1779- 1780.
- Mahmoud, H. A. (2001). Physiological and nutritional studies on sheep feeding certain halophytic plants in Sinai. M. Sc. Thesis, Fac. of Sci., Cairo University, Giza, Egypt.
- Mashudi, Brooke, I. M.; Holmes, C. W. and Wilson, G. F. (1997). Effect of *Mimosa bark* extracts containing condensed tannins on rumen metabolism in sheep and milk production by grazing cows. Proceeding o the New Zealand Society of Anim. Prod., 57: 126.
- Masters, D.G.; Norman, H.C. and Dynes, R.A. (2001). Opportunity and limilletations for animal production from saline land. Asian-Aust. J. Anim. Sci. 14, 199–211.
- Matsuura, M. (2001). Saponin in garlic as modifiers of the risk of cardiovascular disease. J. Nut., 131: 1000-1005.
- McComb, R. B.; Bower, G. and Posen, S. (1979). Alkaline phosphatase. New York, Plenum Press.
- Metwally, N. H. (2001). Studies on some physiological and behavioral aspects in camels.M. Sc. Thesis, Fac. of Agric., Minufiya University, Egypt.
- Miller, S. E. (1966). Textbook of clinical pathology. 7<sup>th</sup> ed. The Williams and Wilkins Co., Baltimore.
- Milne, D. B. (1996). Trace elements. In: Tietz Fundamentals of Clinical Chemistry. Burtis, C. A. and Ashwood, E. R. (Eds.) (1996) 4th Edition, pp. 485- 496.
- Mohamed, M. I. (1996). Studies on desert roughages on camels and small ruminants nutrition. Ph. D. Thesis, Faculty of Agriculture, Cairo University.
- Morehouse, L. A.; Bangerter, F. W.; Deninno, M. P.; Inskeep, P. B.; McCarthy, P. A.; Pettini, J. L.; Savoy, Y. E.; Sugarman, E. D.; Wilkims, R. W.; Wilson, T. C.; Woody, H. A.; Zaccaro, L. M. and Chandler, C. E. (1999). Comparison of synthetic saponin cholesterol absorption inhibitors in rabbits: evidence for nonstoichiometric, intestinal mechanisms of action. J. Lipid Res., 40: 464- 474.

- Muller, H. M.; Leinmuller, E. and Rittner, U. (1989). Effect of tanniferous plant material on protein and carbohydrate degradation in rumen fluid *in vitro*. In Recent Advance of Research. In Antinutritional Factors in Lequme Seeds. (Huisman, J.; Van der Poel, T. F. B. and Liener, I. E. eds.), pp. 156-159, Wageningen.
- Nasr, S. M; Ibrahim, E. A.; Bakeer, A. and Dessouky, M. I. (2002). Clinicopathological and histopathological studies in goats fed on *Atriplex halimus* raised in the Egyptian desert. International Symposium on Optimum Resources Utilization in Salt - Affect Ecosystems in Arid and Semi- arid Regions. Cairo, 8- 11, April, 515- 525.
- National Research Council (NRC) (1975). Nutrient requirements of domestic animals No. 5, Nutrient requirements of sheep. 5th ed. National Academy of Science. National Res. Coun., Washington.
- Neathery, M. W. (1980). Chloride metabolism in cattle. In: Proceedings of the Georgia Nutrition Conference, pp. 137.
- Nelson, A. B.; Macvicor, R. W.; Archer, J. R. W. and Meiske, J. C. (1995). Effect of high salt intake on the digestibility of ration constituents and on nitrogen sodium and chloride retention. J. Anim. Sci., 14: 528-832.
- Nemati, N. (1976). Range rehabilitation problems in the steppe zone of Iran. J. Range Manage. 30, 339–342.
- Norbert, W. (1987). Fundamentals of clinical chemistry. 3<sup>rd</sup> edition. W. B. Saunders Company Philadelphia.
- Ortiz, L. T.; Centeno, C. and Tervino, J. (1993). Tannins in faba bean seeds: effect on the digestion of protein and amino acids in growing chicks. Animal Feed Science and Technology, 41: 271- 278.
- Patra, A. K.; Sharma, K.; Dutta, N. and Pattanaik, A. K. (2002). Effect of partial replacement of dietary protein by a leaf meal mixture containing *Leucaena leucocephala*, *Morus alba* and *Azadirachta indica* on performance of goats. Asian-Aust. J. Anim. Sci., 5 (12): 1732-1737.
- Pearce, K. L.; Pethick, D. W. and Masters, D. G. (2008). The effect of ingesting a saltbush and barley ration on the carcass and eating quality of sheep meat. Animal, 2 (3), pp. 479–490.
- Priolo, A.; Waghorn, G. C.; Lanza, M.; Biondi, L. and Pennisi, P. (2000). Polyethylene glycol as means for reducing the impact of condensed tannins and carob pulp: effects on lambs growth performance and meat quality. J. Anim. Sci., 78 (4): 810.
- Rasool, E.; Rafique, S.; Haq, I. U.; Khan, A. G. and Thomson, E. F. (1996). Impact of fourwing saltbush on feed and water intake and on blood

serum profile in sheep. A.J.A.S., 9 (2): pp. 123-126.

- Reed, J. D.; Soller, H. and Woodward, A. (1990). Fodder tree and straw diets for sheep: intake, growth, digestibility and the effects of phenolics on nitrogen utilization. Anim. Feed Sci. Tecknol. 30: 39.
- Reitman, S. M. D. and Frankel, S. (1957). A colorimeter method for determination of serum glutamic oxaloacetic acid and glutamic pyruvic acid transfereases. Am. J. Clin. Path., 28: 56-63.
- Roeschlau, P.; Bernt, E. and Gurber, W. (1974). Enzymatic determination of total cholesterol in serum. Zklin. Chem. Klin. Biochem. 12 (5) 226.
- Romero, M. J.; Madrid, J.; Hernandez, F. and Ceron, J. J. (2000). Digestibility and voluntary intake of Vine leaves (*Vitis vinifera L.*) by sheep. Small Ruminant Res., 38, pp. 191-195.
- Salem, A. F. Z. M.; Gohar, Y.; El- Adawy, M. M. and Salem, M. Z. M. (2004). Growth inhibitory effect of some anti-nutritional factors extracted from *Acacia saligna* leaves on intestinal bacteria activity in sheep. Proc. 12th Scientific Conf., Egypt. Soc. Amin. Prod., 41: 283- 300.
- SAS Institute (2004). Statistical Analysis System, STAT/ user's guide, Release 9.1, SAS Institute, Cary NC. USA.
- Schirmeister, J.; Willmann, H.; Kiefer, H. And Hallauer, W. (1964): Fuer und wider die brauchbarkeit der endogenen hrea tininclearance der fanktionellen nierendiagnostik. Dtsch. Med. Woschr 89 (9) pp. 1640- 1647.
- Schmillet, J. M. (1964). Thesis, Lyon.
- Shahen, G. F., Zaki, A. A. and Yousef, H. M. (2004) Effect of feeding level on growth nutrient digestibility and feed efficiency for buffalo calves. Egyptian Journal of Nutrition and Feeds, 7, 11-.
- Shaker, Y. M.; Abou El-Ezz, S. S. and Hashem, A. L. (2008). Physiological performance of Barki male lambs fed halophytes under semi-arid conditions. J. Agric. Sci. Mansoura Univ., 33 (9): 6393- 6408.
- Shehata, E.; El- Sayed, I.; Heider, A.; El- Serafy, A. M. and El- Gallad, T. (1988). Effect of supplementing *Atriplex nummularia* with minerals, barley or both on dry matter and water intakes and nutrients metabolism. Proc. 2<sup>nd</sup> Conf. Agric. Develop. Res. Fac. Agric., Ain Shams Univ., Cairo, Egypt, 17- 19 December, Vol (1), Animal Production and Genetics, pp. 111-119.
- Shi-Gang, YU.; Hai-Ying, Li.; Chao, F.; Chong-Kai, Qin and Wu-Jun, Liu. (2010). Effect of early weaning on biochemical indicator in serum of Xinjiang local sheep breeds in

China.J. of Anim. and Vet.Advan., 9(20):2659-2664.

- Silanikove, N.; Gilboa, N.; Perevolotsky, A. and Nitsan, Z. (1996). Goats fed tannin- containing leaves do not exhibit toxic syndromes. Small Ruminant Res., 21 (3): 195- 201.
- Squires, V.R. and Ayoub, A.T. (1994). Halophytes as a Resource for Livestock and for Rehabilitation of Degraded Lands. Kluwer Academic Publisher, Dordrecht Boston, London, 315 p.
- Streeter, M. N.; Hill, G. M.; Wagner, D. G.; Owens. F. N. and Hibberd, C. A. (1993). Effect of bird resistant and non bird resistant sorghum grain on amino acid digestion by beef heifers. J. Anim. Sci., 71: 1648- 1656.
- Sunderman, F. W. Jr. and Sunderman, F. W. (1958). Sm. J. Clin, Pathol, 29: 95.
- Tata, J. R. and Widnell, C. C. (1966). Ribonucleic and synthesis during the early action of hormone. Biochemistry, 98: 604.
- Tietz, N. W. (1986). Textbook of clinical chemistry. W. B. Saunders Co. London, Philadelphia, pp. 796.
- Tietz, N. W. (1990). Clinical guide to laboratory tests. 2<sup>nd</sup> ed. W. B. Saunders Co.
- Trenkle, A. (1978). Relation of hormonal variations to nutritional studies and metabolism of ruminants. J. Dairy Sci., 61: 281-293.
- Trinder, P. (1951). A rapid method for the determination of sodium in serum. Analyst, (907): 596- 599.
- Tripathy, K. C.; Sahu, B. K.; Panda. N. C. and Nayak, B. C. (1984). Toxicity of tannin acid in goats. Indian J. Anim. Sci., 54- (11): 1091-1093.
- Waghorn, G. C.; Shelton, I. D.; NcNabb, W. C. and McCutcheon, S. N. (1994). Effect of condensed tannins in *Lotus pednuculatus* on its nutritive

value for sheep. 2- Nitrogenous aspects. J. Agric. Sc., 109-120.

- Weeth, H. J. and Haverland, L. H. (1961). Tolerance of growing cattle for drinking water containing sodium chloride. J. Anim. Sci., 20: 518-521.
- Weeth, H. J. and Lesperance, A. L. (1965). Renal function of cattle under various water rand salt loads. J. Anim. Sci., 24 (2): 441- 447.
- Wegner, T. N. and Schuh, T. D. (1974). Effect of highly mineralized livestock dairy caws. J. Dairy Sci., 57 (5): 608.
- Wichell, A. R. (1976). Relationships between individual difference in salt appetite of sheep and their plasma electrolyte sates. De. Med. Royal- Vet., 17: 215.
- Yousef, H. M. and Zaki, A. A. (2001). Effect of barley radical feeding on body weight gain and some physiological parameters of growing Friesian crossbred calves. Egyptian Journal of Nutrition and Feeds, 6 (Special Issue), 465.
- Zanouny, A. I.; Abd-El Moty, A. K. I.; El –Barody, M. A. A.; Sallam, M. T. and Abd El –Hakeam, A. A. (2013). Effect of supplementation with *Nigella sativa* seeds on some blood metabolites and reproductive performance of Ossimi male lambs. Egyptian Journal of Sheep and Goat Sciences, Vol. 8 (1), P: 47-56.
- Zarkawi, M.; Al-Masri, M.R. and Khalifa, K. (2005). Nutritive value of *Sesbania aculeate* grown on salty soil and its effect on reproductive parameters of Syrian Awassi ewes. Aust. J. of Anim. Res., 56 (8): pp. 819-825.
- Zhu, J. and Filippich, L. J. (1995). Acute intraabomasal toxicity of tannic acid in sheep. Vet. Hum. Toxical., 37 (1): 50- 54.
- Zhu, J.; Filippich, L. J. and Alsalami, M/ I. (1992). Tannic acid intoxication in sheep and mice. Res. Vet. Sci., 53 (3): 280-292.