Journal of American Science

Websites: http://www.jofamericanscience.org http://www.sciencepub.net

Emails: editor@sciencepub.net sciencepub@gmail.com



Screening Donated Blood for Transfusion-Transmissible Cytomegalovirus Infection Among Libyans

Farag Bleiblo^{1,4*}, Abdelhakim Eljaki¹, Khaled Elwaheishi¹, Eman Almismary, Mabroka Aljlale¹, Rabea Alghazal², and Mohammed Abraheem³

 ¹ Department of Microbiology, Faculty of Science, University of Benghazi, Libya
² Immunology & Virology Laboratory, Benghazi Children Hospital, Libya
³ Department of Botany, Faculty of Arts and Science/Alkufra, University of Benghazi, Libya
⁴ Department of Chemistry and Biochemistry and the Biomolecular Sciences Programme, Laurentian University, Sudbury, ON, Canada, P3E 2C6

*fx bleiblo@laurentian.ca

Abstract: Human cytomegalovirus (HCMV) is a ubiquitous DNA-containing herpesvirus causes severe and fatal diseases in immunocompromised patients and a prevalent cause of virus-associated birth defects. Blood and apheresis donations intended for transfusion to immunosuppressed individuals, neonates and pregnant women should be screened for evidence of CMV infection prior to the release for clinical use. The effective national programmes for quality-assured screening of donated blood have not yet been fully established, hence this study was undertaken to assess whether any blood borne-CMV infections pose a significant threat to the safety of the blood supplies owing to its seroprevalence in the general population. A total of 200 voluntary blood donor subjects admitted to the Blood Bank of Benghazi/Libya were screened for transfusion-transmissible CMV (TT-CMV) using a highly sensitive CMV total IgG and IgM antibody enzyme immunoassay as well as CMV pp65 antigenemia assays. We determined that the overall seropositivity for IgG antibodies (80.50 %) was higher than that of IgM antibodies (39.00 %), but only 2 (1.00 %) individuals out of these donors were seropositive for the CMV-antigenic protein pp65. The frequency of CMV infection based on gender was incomparable due to the small population number of females. According to age, there was not influence of various age groups on prevalence of anti-CMV IgG antibodies, while a progressive increase in seropositivity of CMV-IgM antibodies with age was detected. The age groups were not significantly associated with CMV prevalence. In contrast, only 2 (1.00 %) patients were shown to be positive for all three performed assays indicating a recurrent infection. Our findings prove a risk of primary transfusion-associated transmission of CMV and may provide a policy guidance on ensuring safe blood supplies accessible to all patients who require transfusion.

[Farag Bleiblo, Abdelhakim Eljaki, Khaled Elwaheishi, Eman Almismary, Mabroka Aljlale, Rabea Alghazal, and Mohammed Abraheem. Screening Donated Blood for Transfusion-Transmissible Cytomegalovirus Infection Among Libyans. J Am Sci 2019;15(12):104-108]. ISSN 1545-1003 (print); ISSN 2375-7264 (online). http://www.jofamericanscience.org. 11. doi:10.7537/marsjas151219.11.

Keywords: CMV, Seroprevalence, Blood Transfusion, TT-CMV, Enzyme Immunoassays

Introduction

Human cytomegalovirus (HCMV) is a ubiquitous virus infection distributed worldwide. This virus is the most common infectious cause of birth defects and congenital diseases, the most significant and difficult opportunistic pathogen affecting immunocompromised patients (Cannon *et al.* 2010, Mocarski *et al.* 2014). HCMV infects an overwhelming majority of the population, transmitted efficiently throughout life and universally through contacts with bodily secretions. After the initial acquisition of HCMV, the virus replicates and causes a systemic infection, sometimes detected as a leukocyte-associated viremia, and disseminates to secretory organs such as salivary glands and kidney where replication produces virus found in secretions (Brooks *et al.* 2010). Viruses may

be shed in any body fluids, including urine, saliva, tears, semen, and cervical secretions, and persistent shedding may continue for months to years, depending on age and immune status of the host (Brooks *et al.* 2010, Trkulic *et al.* 2000). Like other human herpesviruses, HCMV is never completely cleared and remains latent for the life of the host. Persistently and sporadically shed virus is an important recurrent source of virus for transmission. Susceptibility to HCMV disease is associated with a compromised immune system, particularly related to defects in cell-mediated CD4 and CD8 T-cell functions (Mocarski *et al.* 2014). Despite potential antiviral drugs aimed to control the overall disease burden, the HCMV remains an important etiologic agent of opportunistic

infections and disease in immunocompromised individuals following organ transplantation and hematopoietic cell allografting, immunosuppressive therapies, and genetic or acquired immunodeficiency (Cannon *et al.* 2010, Cavlek *et al.* 2015, Mocarski *et al.* 2014).

HCMV circulates in the leucocytes and free in plasma during active infection. It subsequently persists latently in leucocytes as well as in other noncirculating body cells and may be released into the bloodstream following reactivation of latent virus. It is thus readily transmitted by blood transfusion, although transmission is generally a concern only when immunocompromised individuals, transfusing neonates, and pregnant women. In populations with a higher incidence of CMV, there is a correspondingly higher risk of blood being donated by viremic individuals. In such cases, leucodepletion will not prevent transmission. Thus, for the majority of countries, anti-CMV screening is still central to the prevention of post-transfusion CMV (van Prooijen HC et al. 1994, Roback JD. et al. 2002).

The provision of safe and efficacious blood and blood components for transfusion involves several processes including testing of blood donation samples for transfusion-transmissible infections (TTIs) to exclude blood donations at risk of transmitting infection from donors to recipients is a critical part of blood safety and availability. Therefore, this study aimed to screen donated blood for evidence of CMV infection prior to the release of blood for clinical use to establish an effective national blood screening programme to protect the recipients of blood transfusion from TTIs CMV (van Prooijen HC *et al.* 1994, Roback JD. *et al.* 2002).

Materials and Methods Patient Population

Two hundred subjects admitted to the Blood Bank of Benghazi/Libya were recruited in this study. Informed consent was obtained from all patients and the protocol was approved by the Blood Services Committee. Demographic Ethics information including age, sex, marital status, literacy status, residential status, socioeconomic status were obtained by means of referring to medical records and personal interviews. Throughout their tenure on the Blood Unit, a 5 ml of blood was drawn from donors by vein puncture and placed in plastic disposable tubes; it was left to stand at room temperature (20-25°C) to allow for clotting, then the sera was separated by centrifugation 10,000 rpm for 5 minutes. Sera samples were stored at -20 °C and later tested for antibodies cytomegalovirus by serological investigation.

Detection of CMV IgG

Microplate-based enzyme-linked immunoassay (ELISA) was used for the qualitative and quantitative detection of anti-CMV IgG antibodies to cytomegalovirus in donors' sera according to the manufacturer's instructions (Autobio Diagnostics Co. LTD). The assay is based upon the two steps indirect enzyme immunoassay (EIA). Briefly, in the first step, sample and recombinant CMV coated microwells are combined. During the incubation, the anti-CMV antibodies present in the sample bind to the antigen coated in the wells. After the washing, in the second step enzyme-conjugate is added to the reaction mixture. The mouse anti-human IgG in the enzymeconjugate is allowed to react with the anti-CMV IgG attached to the solid phase in the first step. Then, a complex is generated between the solid phase, the anti-CMV IgG within the sample, and the mouse antihuman IgG in the enzyme conjugate. Then, the substrate A and substrate B are added and catalyzed by this complex. The resulting chromogenic reaction is measured in absorbance. The color intensity, which corresponds to the amount of CMV IgG antibodies present in the specimens, is measured with a microplate reader at 450/630 nm and the results were according to the interpreted manufacturer's instructions. Samples with optical densities 10% or more below the cutoff were recorded as negative. those with optical densities between 10% below and 10% above the cutoff were equivocal, and all others were positive.

Detection of CMV IgM

To determine if the blood donors have acute or primary infection, serum samples were assayed for CMV-specific IgM using a CMV IgM ELISA (Autobio Diagnostics Co, LTD) and the results were interpreted according to the manufacturer's instructions. Samples giving an absorbance less than the cut-off value were considered negative for the presence of CMV-specific IgM antibodies whereas those giving an absorbance equal to or greater than the cut-off value were considered reactively positive.

Detection of CMV Antigenemia

To quantitate the viral antigenemia, we performed Sandwich- ELISA to assay CMV lower matrix protein pp65 in donors' sera according to the manufacturer instructions (Sung long Biotech). The microtiter stripplate provided with this assay are precoated with antibodies specific to CMV pp65. Standards and samples were added to stripplate wells and combined to the specific antibody and then a horseradish CMV pp65 (HRP)-conjugate specific for CMV pp65 was added to each microelisa stripplate well and incubated. The TMB substrate solution was added to each after washing away the free components. The wells containing the CMV pp65 and

HRP-conjugated CMV pp65 antibody appeared blue and then turned yellow after adding the stop solution. The optical density (OD) was measured by microtiter plate reader at 450 nm. The concentration of CMV pp65 in the sample was calculated by comparing the OD of the sample to the standard curve according to the manufacturer instructions.

Results and Discussion

Two hundred subjects of blood donors were followed during the course of the study. Our results

revealed that 161 (80.50 %) out of these subjects were seropositive for CMV-IgG indicating a past exposure to infection, while 78 (39.00 %) individuals were seropositive for CMV-IgM indicating a recent or primary infection (Table 1). Based on the laboratory diagnosis of CMV-pp65, we determined that only 2 (1.00 %) donors had CMV viremia. These two donors ere shown to be positive for all three performed assays indicated the presence of recurrent infection.

Table 1: Prevalence of CMV-IgG, CMV-IgN	, and CMV-pp65 Antibodies in Blood Donors.
---	--

Total No. of Patients	CMV-IgG		CMV-IgM		CMV-pp65	
	No	%	No	%	No	%
200	161	80.50	78	39.00	2	1.00

Cytomegalovirus is considered as one of the most encountered opportunistic viral pathogens distributed universally among human population affecting particularly immunocompromised patients. Several epidemiological studies on seroprevalence of HCMV infection in blood donors has been reported. It seems that the prevalence of CMV-IgG antibodies among blood donors varies according to the national infection rates of the general population and the socioeconomic characteristics of the blood donors. In agreement with our findings, several studies conducted in different regions of India showed a significantly high CMV seropositivity among blood donors population ranged between 90% and 100% (Kothari A et al. 2011, Pal SR et al. 1972, Madhavan HN et al. 1974, Mukundan P et al. 1977). The different contributing factors for high seroprevalence may be related to low socioeconomic status and closeness of contact within population, environmental factors and climatic conditions. The overall seroprevalence of CMV among Japanese blood donors was 76.6 % and the CMV DNA was detected in the cellular fraction of some donors (Yasumi Furui et al. 2013). In Nigeria, prevalence of anti-CMV IgG antibodies was 96.2% and that of IgM was 2.6% (Bawa MK et al. 2019). In a German study, the seroprevalence of CMV among blood donors ranged from 30% and increased up to about 80% in donors older than 65 years (Hecker et al., 2004). However, young donors already had a distinctly higher seroprevalence (about 70%) in Australia than was reported for Europe or North America (Lancini et al., 2016). Ethnicity has been suggested also to be an important factor for CMV seroprevalence. Low rates are reported for non-Hispanic Whites (about 50%) and very high rates for South Asians [289% of South Asian UK-born women and 98% among women born in South Asia and living in the UK (Pembrey et al., 2013). IgM appears first in response to a CMV primary infection CMV or reactivations. In general, IgM antibodies might be detectable both prior to IgG antibodies (Ziemann et al., 2010) or shortly after IgG seroconversion, and remain positive for several months. Studies on the IgM seroprevalence in blood donors are less frequent than those about IgG seroprevalence. Most studies reported that IgM seroprevalence is much lower comparing to that of IgG (Furui et al., 2013). In this study, we could not compare the overall seropositivity of CMV according to gender because the female participants of voluntary blood donors were very small. However, several lines of evidence including both genders consistently reported prevalence rates that are slightly higher in women than in men. Population-based evaluation of HCMV prevalence that have been derived from the National Health and Nutrition Examination Survey (NHANES), starting with a 1988 to 1994 set and continuing with 1999 to 2004 set (6 years of age or older) estimated that more female patients (63.5%) than male (54.1%) were infected (Bate et al. 2010, Staras et al. 2006). According to the age-related prevalence of CMV infection, the rates of CMV-IgG antibodies was detected to be comparable in different age groups that have been investigated except for the people within ≥ 60 years where there was no infection detected (Table 2). Regarding the IgM seropositivity, we found that the age group within 45-59 years of old had the highest rate (51.28 %) followed by those within the age group 30-44 years (39.80 %). The CMV-pp65 positivity rate was determined to be higher within the age group younger than 30 years (1.92 %). Although, CMV seroprevalence did not differ significantly between age groups (p > 0.05).

Age	No. of Patients	CMV-IgG		CMV-IgM		CMV-pp65	
Group		No	%	No	%	No	%
< 30	52	43	82.69	16	30.76	1	1.92
30 - 44	103	85	82.52	41	39.80	1	0.97
45 - 59	39	32	84.61	20	51.28	0	0
≥60	6	0	0	1	16.66	0	0
Total	200	160	80.50	78	39.00	2	1.00
Р		> 0.05		> 0.05		> 0.05	

Table 2: Distribution of CMV- IgG, CMV-IgM, and CMV-pp65 Antibodies According to Age Groups

Although the prevalence of CMV infection increases with age in many age groups that have been studied, the overall prevalence of infection and the age vary greatly according to living circumstances and socioeconomic status based on personal interview. In general, prevalence of infection is greater, and virus is acquired earlier in life in developing countries compared to the United States and Europe, where prevalence is greater in the lower socioeconomic strata, among nonwhites, and in immigrant populations from developing countries (Grifiths et al. 1984, Stadler et al. 2010). Based on a large samplings study derived from NHANES, it has been shown that the overall age-adjusted prevalence of HCMV infection in the United States remains about 59%, with about one-half of the U.S. population aged 6 to 49 infected (Bate et al. 2010, Staras et al. 2006). This study reported that the age-related prevalence of HCMV infection was greater within groups aged greater than 30 years. It has been demonstrated that the seroprevalence of blood donors aged between 18 and 30 years was about 30% and increased gradually up to about 80% in German donors older than 65 years (Hecker et al., 2004). Similarly, older donors also had an increased seroprevalence among Australian populations. However, young donors already had a distinctly higher seroprevalence in Brazil and Australia than was reported for Europe or North America (Souza MA et al. 2010, Lancini et al., 2016). Based on these findings, we highly recommend a quality-assured screening of all donated blood for transfusion transmissible infections, including CMV to establish effective national programmes to easily prevent the unacceptable risk of acquiring lifethreatening diseases.

Corresponding Author:

Dr. Farag Bleiblo Department of Microbiology University of Benghazi Benghazi, Libya Telephone: + 218 944077157 E-mail: <u>fx_bleiblo@laurentian.ca</u>

References

- 1. Bate SL, Dollard SC, Cannon MJ. Cytomegalovirus seroprevalence in the United States: the national health and nutrition examination surveys, 1988–2004. Clin Infect Dis 2010;50(11):1439–1447.
- Brooks, G.F., Carroll, K.C., Butel, J.S., Morse, S.A. and Mietzner, T.A. 2010. Jawetz, Melnick and Adelberg's Medical Microbiology. 25th ed. McGraw–Hill companies. Inc. United States of America. pp: 467-488.
- Cannon, M.J., Schmid, D.S. and Hyde, T.B. 2010. Review of cytomegalovirus seroprevalence and demographic characteristics associated with infection. Rev Med Virol, 20(4), pp: 202-213.
- Cavlek, T., Kolaric, B., Sternak, S., Kos, M., Kaic, B. and Galinovic, G. 2015. Prevalence and dynamics of Cytomegalovirus infection among patients undergoing chronic hemodialysis. Indian J. Nephrol, 25(2), pp: 95–98.
- Furui, Y., Satake, M., Hoshi, Y., Uchida, S., Suzuki, K. & Tadokoro, K. (2013) Cytomegalovirus (CMV) seroprevalence in Japanese blood donors and high detection frequency of CMV DNA in elderly donors. *Transfusion*, 53, 2190–2197.
- Grifiths PD, Baboonian C. A prospective study of primary cytomegalovirus infection during pregnancy: inal report. BJOG 1984;91:307–315.
- Hecker, M., Qiu, D., Marquardt, K., Bein, G. & Hackstein, H. (2004) Continuous cytomegalovirus seroconversion in a large group of healthy blood donors. *Vox Sanguinis*, 86, 41– 44.
- Kothari A, Ramachandran VG, Gupta P, Singh B, Talwar V. Seroprevalence of Cytomegalovirus among Voluntary Blood Donors in Delhi, India. J Health, Populat Nutr 2011;20(4):348–51.
- Lancini, D.V., Faddy, H.M., Ismay, S., Chesneau, S., Hogan, C. & Flower, R.L. (2016) Cytomegalovirus in Australian blood donors: seroepidemiologic and seronegative red blood cell component inventories. *Transfusion*, 56(6 Pt 2), 1616–1621.

- Madhavan HN, Prakash K, Agarwal SC. Cytomegalovirus infection in Pondicherry (South India) – a serological survey. Indian J Med Res 1974;62(2):297–300.
- 11. Marli Adelina Souza, Ana Maria Passos, Arício Treitinger and Celso Spada. Seroprevalence of cytomegalovirus antibodies in blood donors in southern, Brazil. Revista da Sociedade Brasileira de Medicina Tropical 43(4):359-361.
- Mocarski, ES, Shenk, Griffiths PD, Pass RF: Cytomegaloviruses. In: David M. Knipe and Peter M. Howley. Field Virology, 6th ed. Lippincott Williams & Wilkin, 2013, 1960-2014.
- 13. Mukundan P, Jadhav M, John TJ. Prevalence of cytomegalovirus antibody in young children in Vellore. Indian J Med Res 1977;65(5):589–92.
- 14. Musa Kalamullah Bawa, Aisha Mamman, Adebola Olayinka, Saheed Gidado, Ndadilnasiya Endie Waziri, Muhammad Shakir Balogun, Kabir Ibrahim Getso, Mahmood Muazu Dalhat, Peter Nsubuga, Nuruddeen Aliyu, Hussaini Bala, Hauwa Muhammad, Suleiman Haladu, Usman Lawan Shehu, Patrick Mboya Nguku. The Pan African Medical Journal. 2019;32 (Supp 1):6., 1-6.
- 15. Pal SR, Chitkara NL, Krech U. Seroepidemiology of cytomegalovirus infection in and around Chandigarh (Northern India). Indian J Med Res 1972 Jul;60(7):973–8.
- Pembrey, L., Raynor, P., Griffiths, P., Chaytor, S., Wright, J. & Hall, A. J. (2013) Seroprevalence of cytomegalovirus, Epstein Barr virus and varicella zoster virus among pregnant

11/22/2019

women in Bradford: a cohort study. *PLoS ONE*, 8, e81881.

- 17. Roback JD. CMV and blood transfusions. *Reviews in Medical Virology*, 2002, 12(4):211–219.
- Stadler, L. P, Bernstein, D. I., Callahan, S. T. et al. 2010. Seroprevalence of Cytomegalovirus (CMV) and risk factors for infection in adolescent males. Clin Infect Dis, 51 (10), pp: e76-e81.
- 19. Staras SA, Dollard SC, Radford KW, et al. Seroprevalence of cytomegalovirus infection in the United States, 1988–1994. Clin Infect Dis 2006;43(9):1143–1151.
- Trkulic, M., Jovanovic, D., Ostojic, G., Kovacevic, Z. and Taseski, J. 2000. Cytomegalovirus infection in patients with kidney diseases. Vojnosanit Pregl, 57(5), pp: 63-67.
- 21. van Prooijen HC, Visser JJ, van Oostendorp WR, de Gast GC, Verdonck LF. Prevention of primary transfusion-associated cytomegalovirus infection in bone marrow transplant recipients by the removal of white cells from blood components with high-affinity filters. *British Journal of Haematology*, 1994,;87(1):144-147.
- 22. Yasumi Furui, Masahiro Satake, Yuji Hoshi, Shigeharu Uchida, Ko Suzuki, and Kenji Tadokoro. Transfusion 2013;53:2190-2197.
- 23. Ziemann, M., Unmack, A., Steppat, D., Juhl, D., Görg, S. & Hennig, H. (2010) The natural course of primary cytomegalovirus infection in blood donors. *Vox Sanguinis*, 99, 24–33.